

# Les Syndromes myélodysplasiques

L'apport de la cytométrie en flux

*Corata Belgique 2015, Beaune*

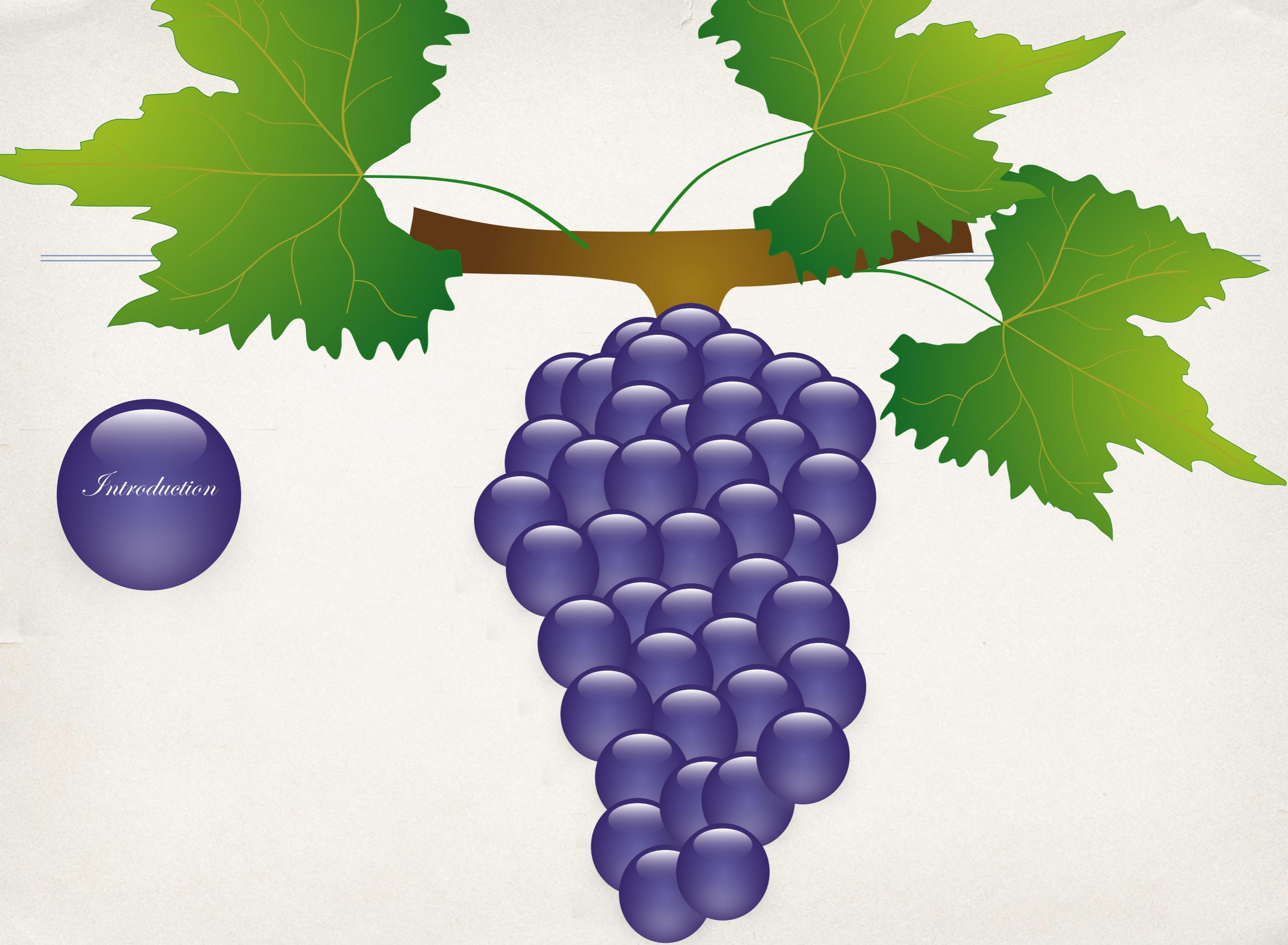
# Myélodysplasie

SMD/MDS

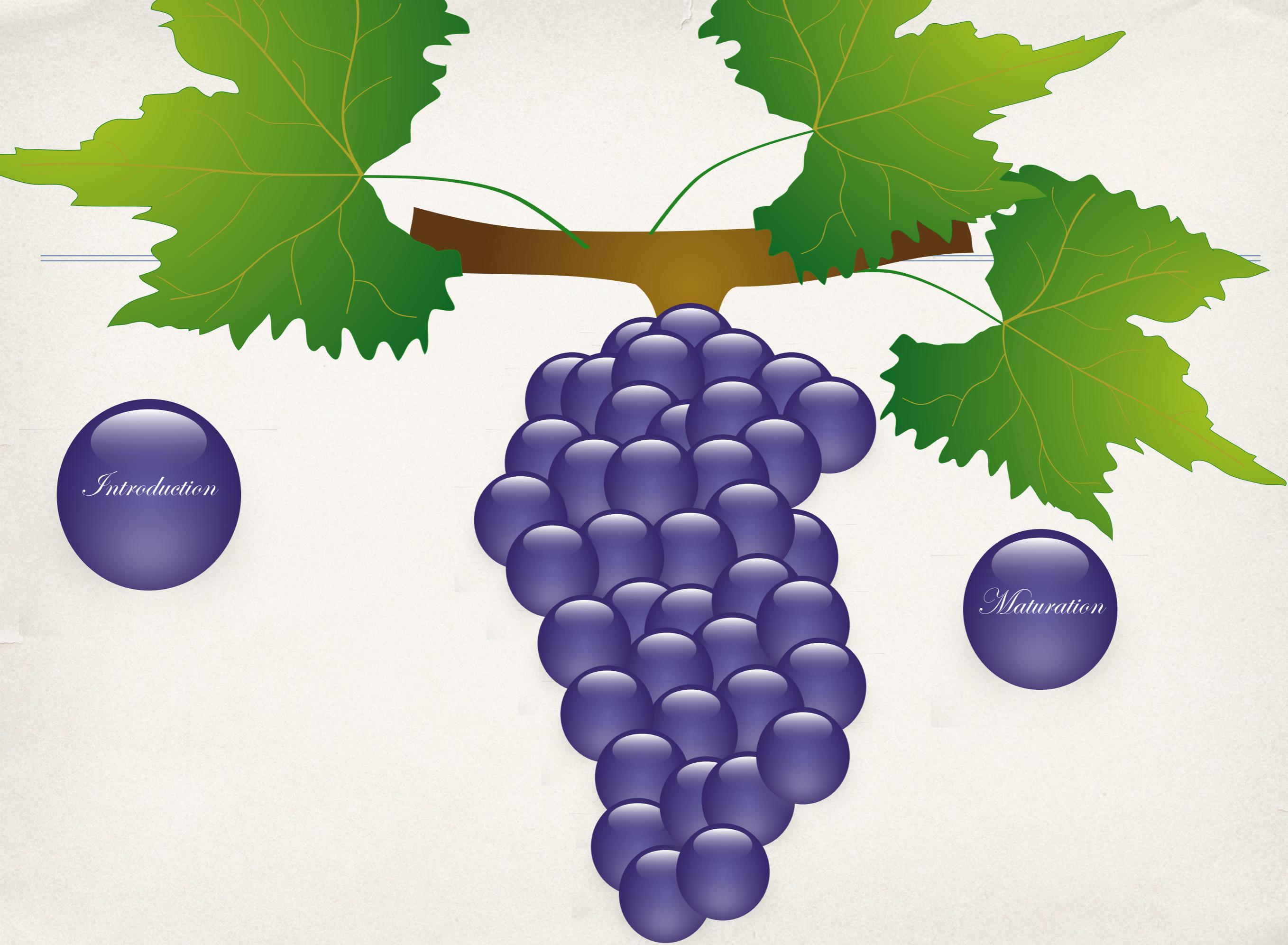


«The myelodysplastic syndromes are a group of clonal haematopoietic stem cell diseases characterized by cytopenia(s), dysplasia in one or more of the major myeloid cell lines, ineffective haematopoiesis, and increased risk of development of acute myeloid leukaemia»



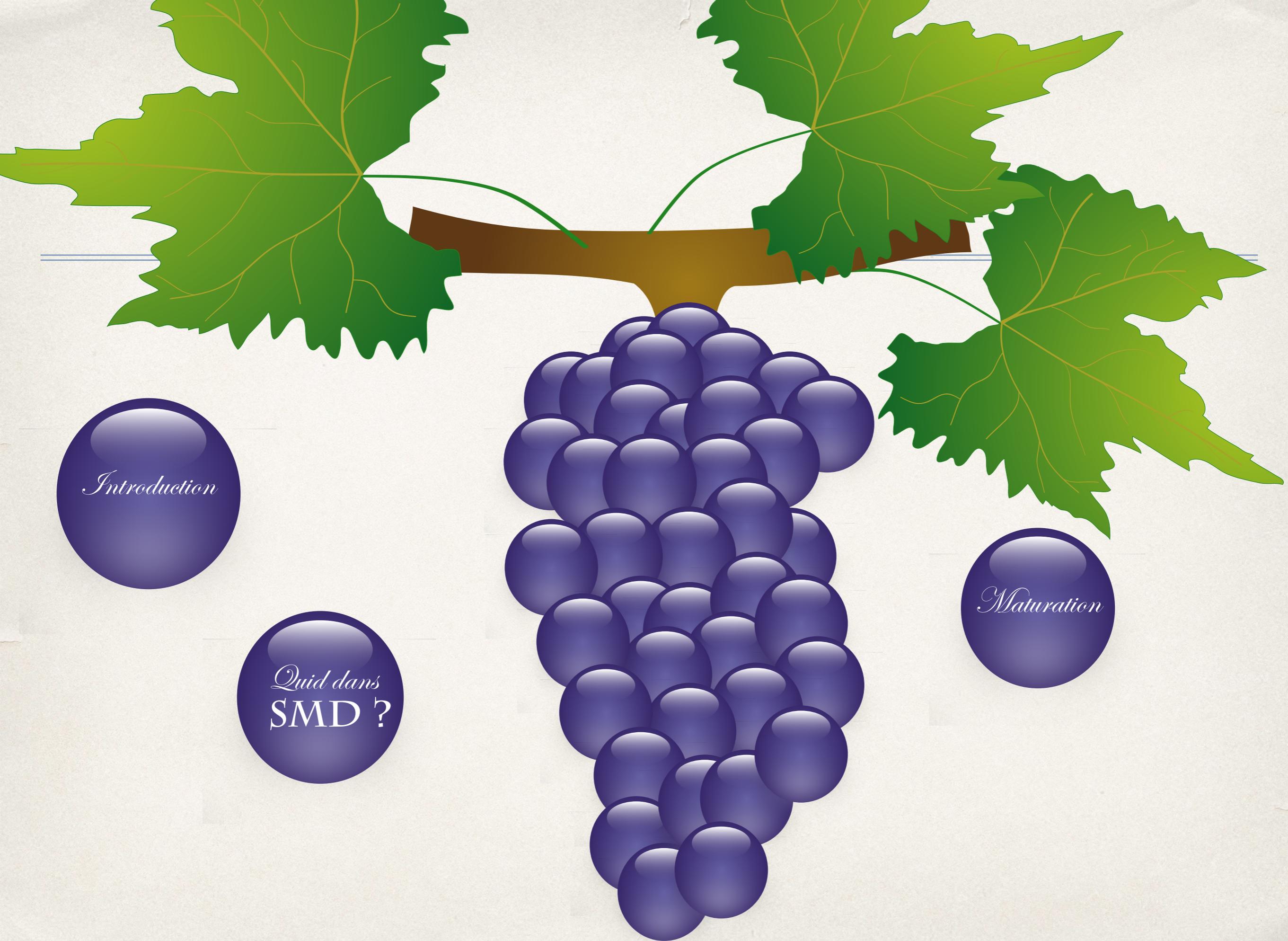


*Introduction*



*Introduction*

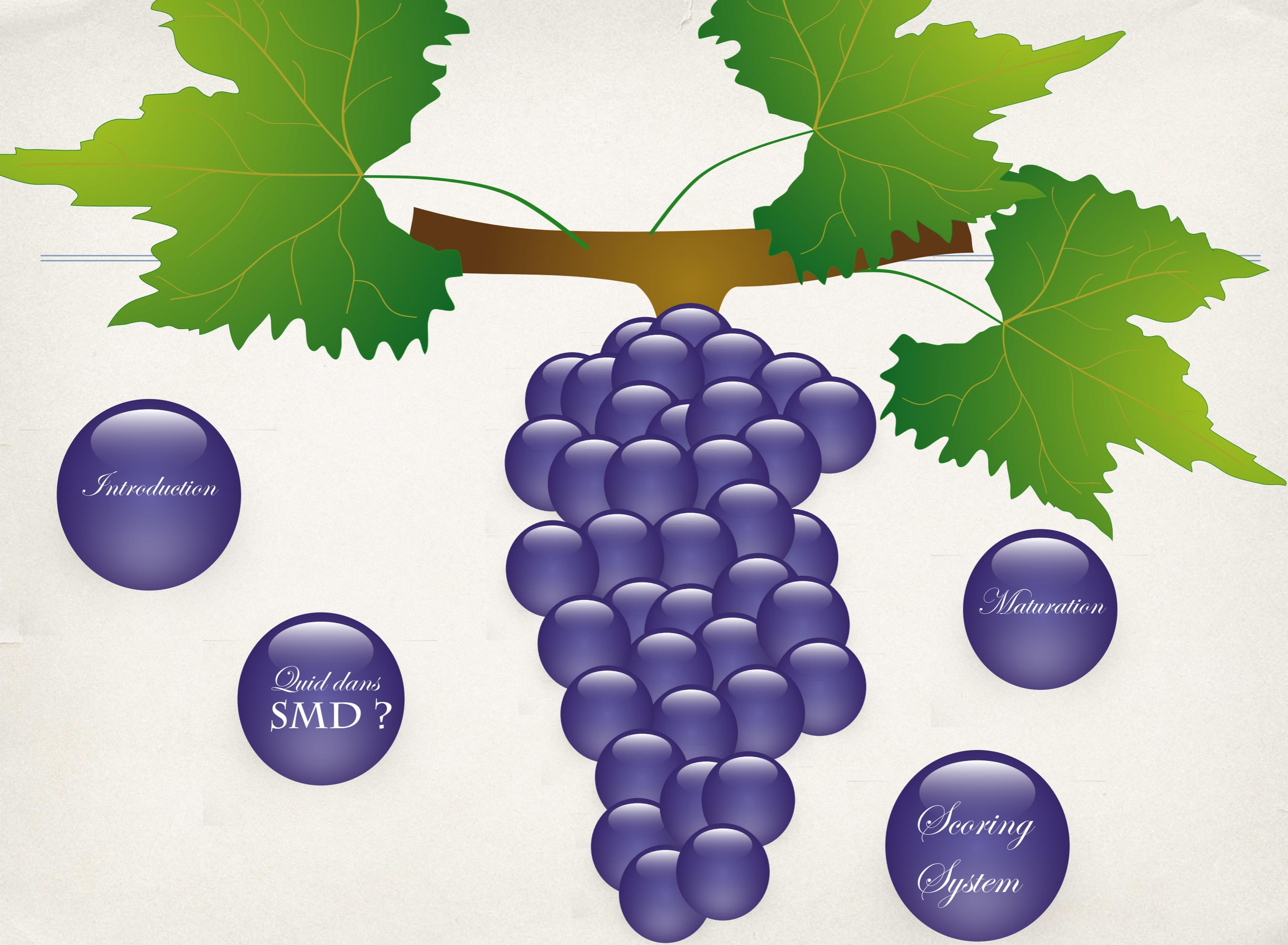
*Maturation*



*Introduction*

*Quid dans  
SMD ?*

*Maturation*



*Introduction*

*Quid dans  
SMD ?*

*Maturation*

*Scoring  
System*



*Introduction*

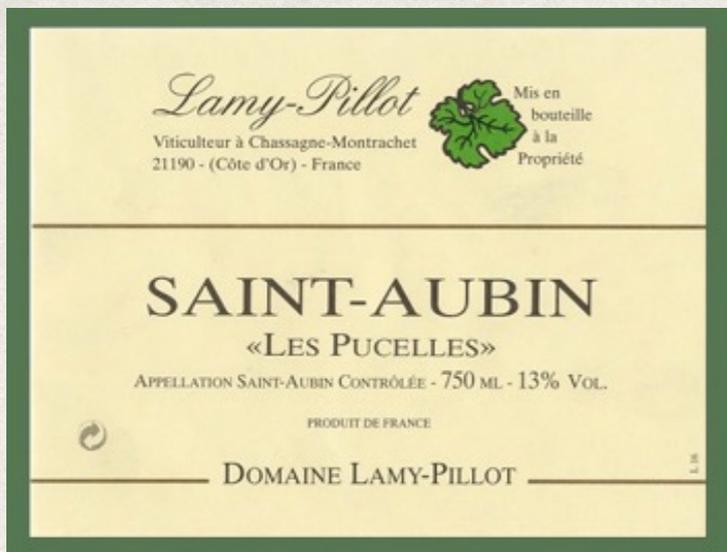
*Quid dans  
SMD ?*

*Mise en  
pratique*

*Maturation*

*Scoring  
System*

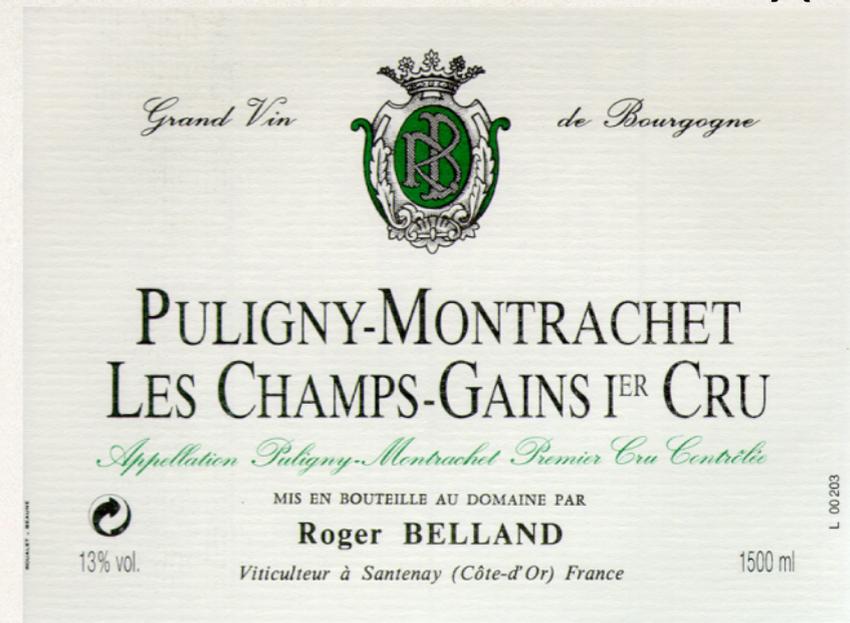




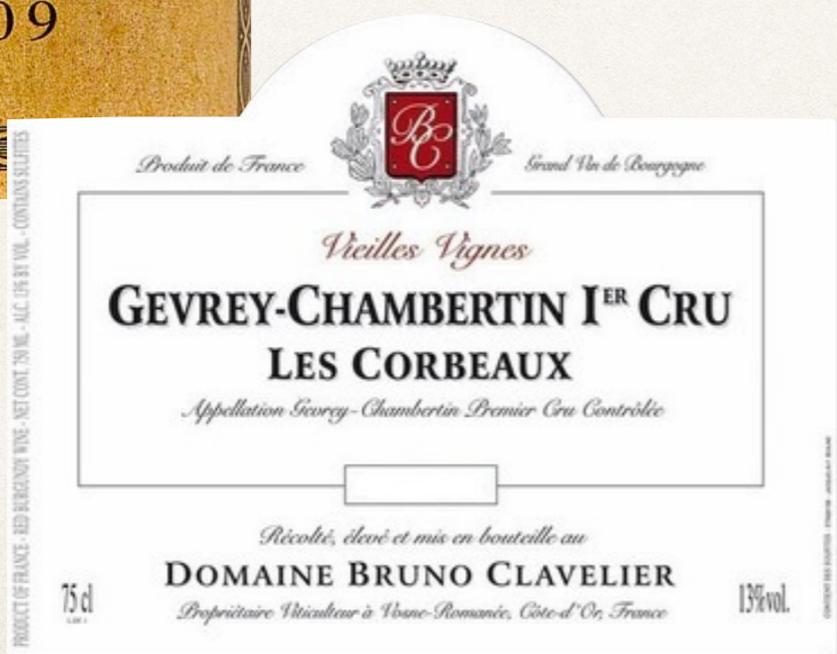
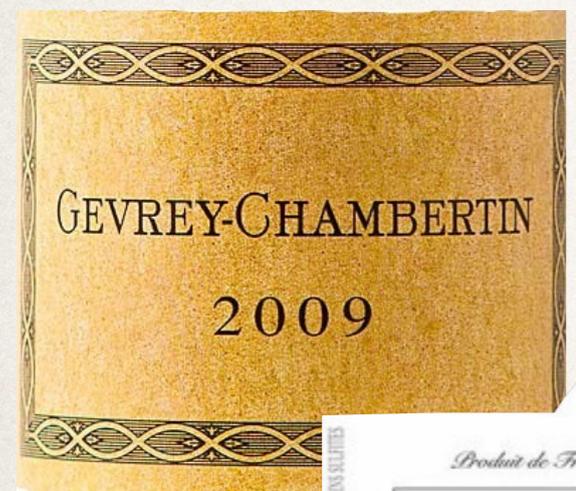
Refractory cytopenia with unilineage dysplasia (RCUD)



Refractory cytopenia with multilineage dysplasia (RCMD)



Refractory anemia with ring sideroblasts (RARS)



Refractory anemia with excess blasts : RAEB-1 or RAEB-2



Myelodysplastic syndrome with isolate 5q-



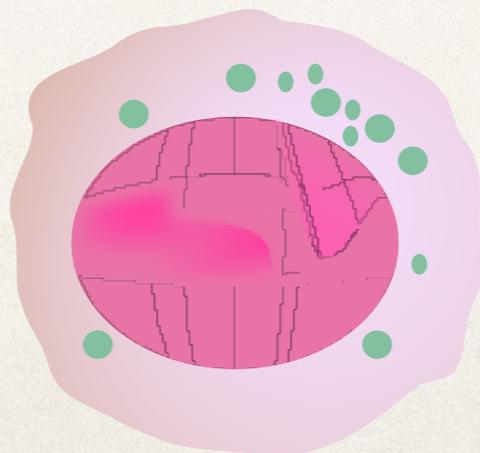
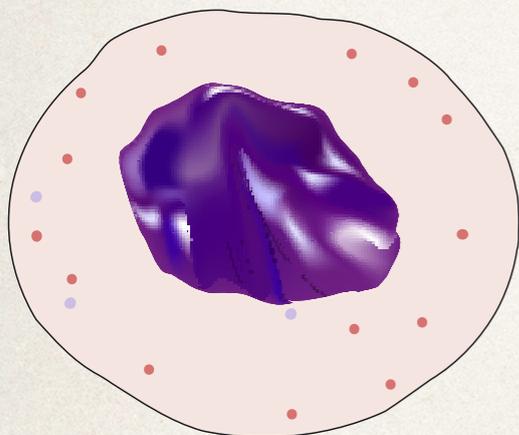
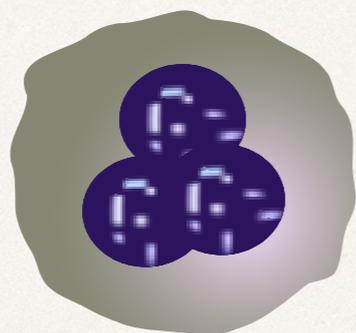
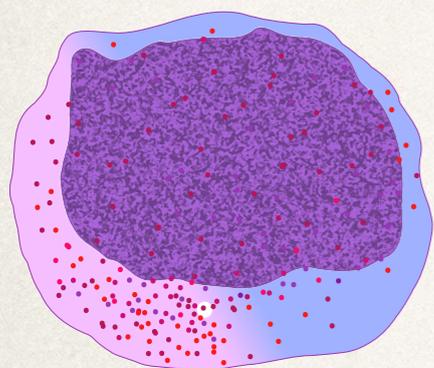
Myelodysplastic syndrome unclassifiable

# SMD ?



- \* Etiologie : maladies hétérogènes. Sélection clonal d'une cellule souche hématopoïétique (comment ?), avantage de croissance.
- \* Physiopathologie : anomalies des voies apoptotiques, instabilités génétiques et génomiques, épigénétique, ...
- \* Epidemiologie
  - \* 2-10 / 100.000, augmente avec l'âge, prédominance masculine.
- \* IPSS (*International Prognostic Scoring System*) : %blastes, cytopénies, cytogénétique
- \* WPSS (WHO 2008) : WHO categorie, Caryotype, besoins transfusionnels

# Outils au laboratoire



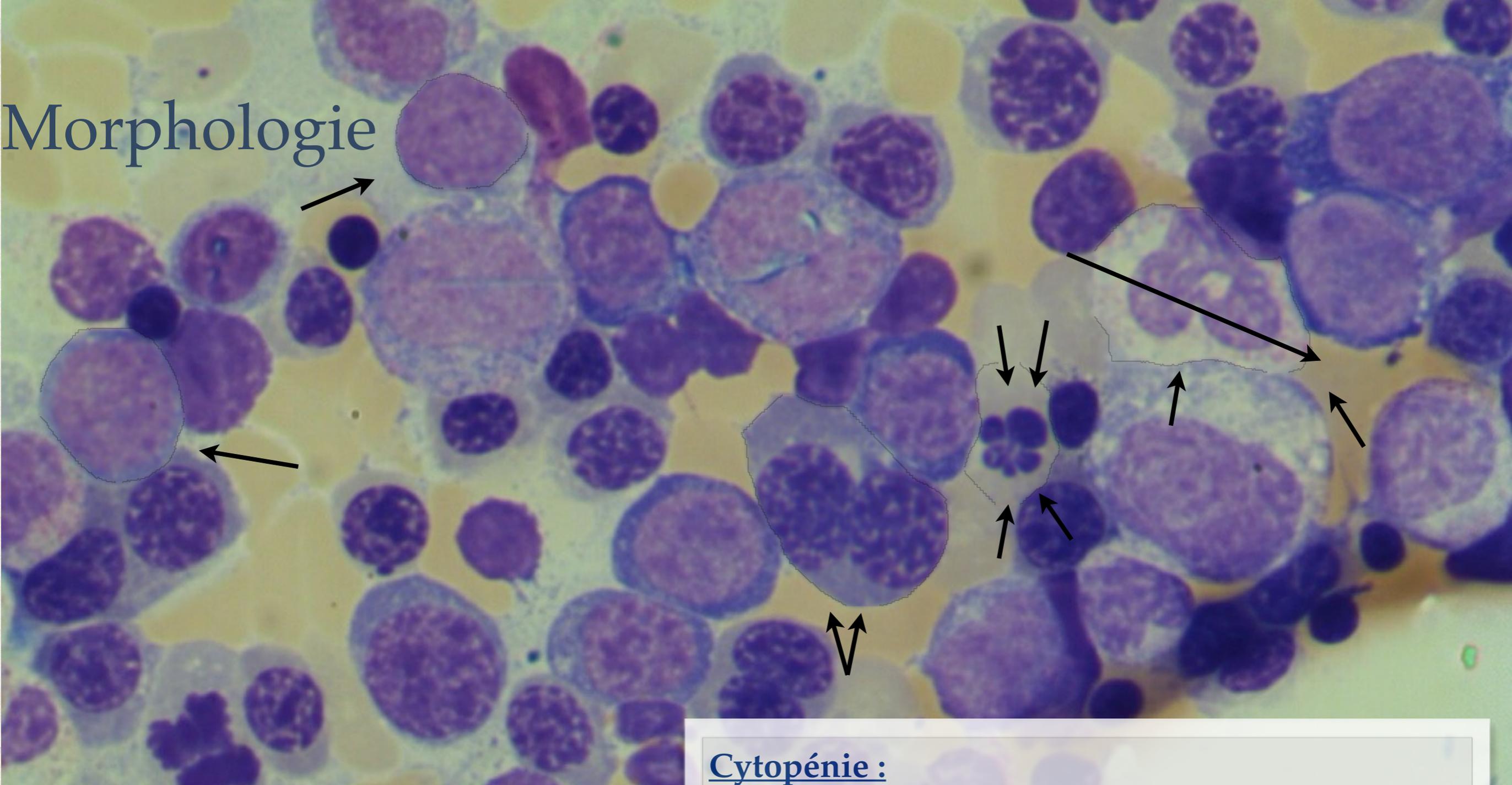
Cytogénétique



Biologie moléculaire

Morphologie

# Morphologie



- SMD bas grade sans anomalies morphologiques
- Toxique (alcool, médicaments, ...), infections virales, carences nutritionnelles.
- Certains cas : hypocellularité ou fibrose.

## Cytopénie :

- Hémoglobine < 10 g/dL
- Neutrophiles <  $1,8 \times 10^9$
- Plaquettes <  $100 \times 10^9$

## Dysplasie :

- $\geq 10\%$  de dysplasie dans au moins 1 lignée

## Blastes médullaires :

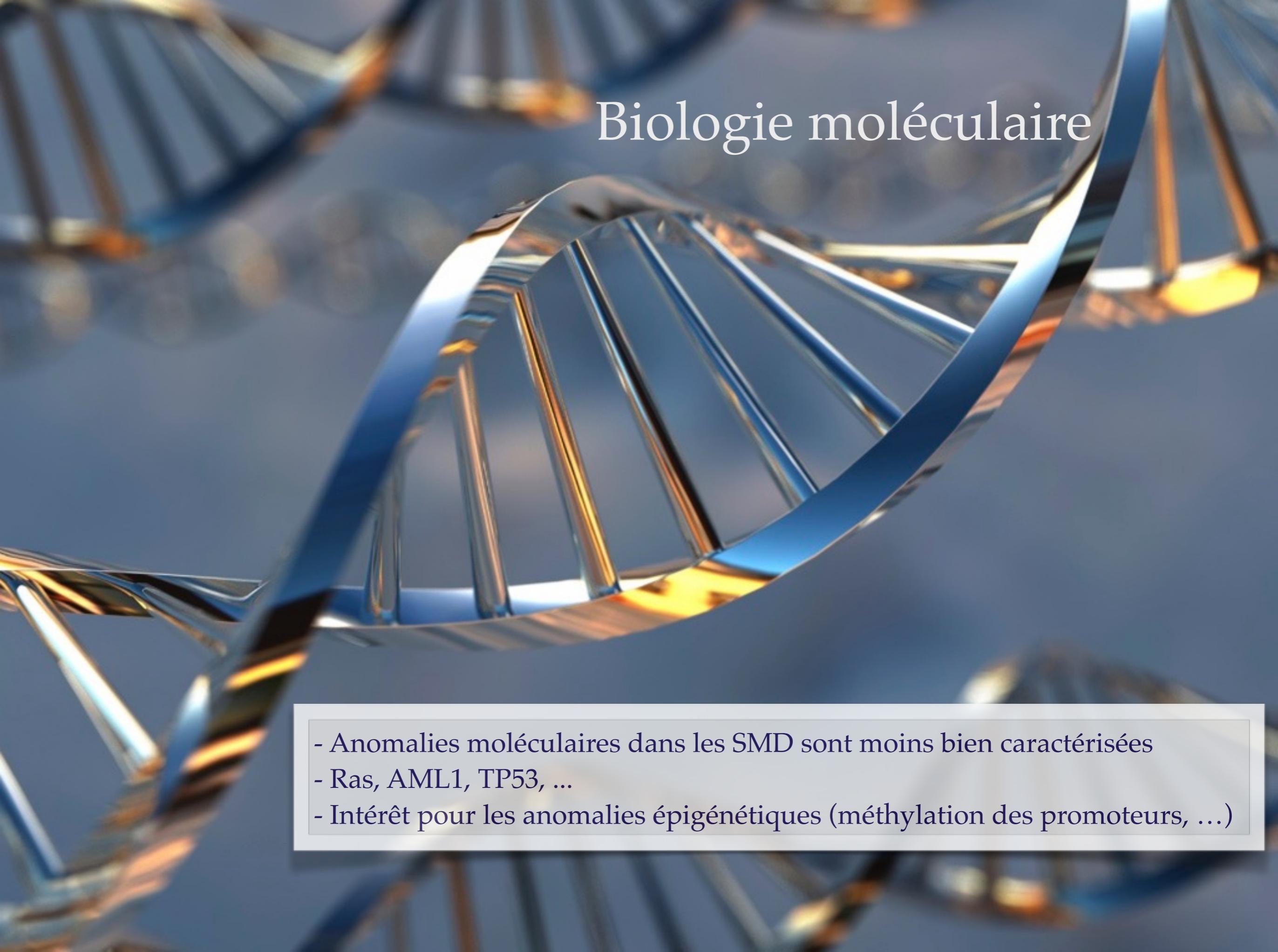
- < 20% (5-9% : AREB-1 et 10-19% : AREB-2)

## Cytogénétique



- Anomalies cytogénétiques clonales ~50% des cas de SMD
- -5 / del(5q), -7 / del(7q), del(20q), 17p, +8, -Y, caryotypes complexes, ...
- **Hétérogénéité** des anomalies génétiques
- Pronostique !
- Nouveaux outils, études plus larges, ...

# Biologie moléculaire

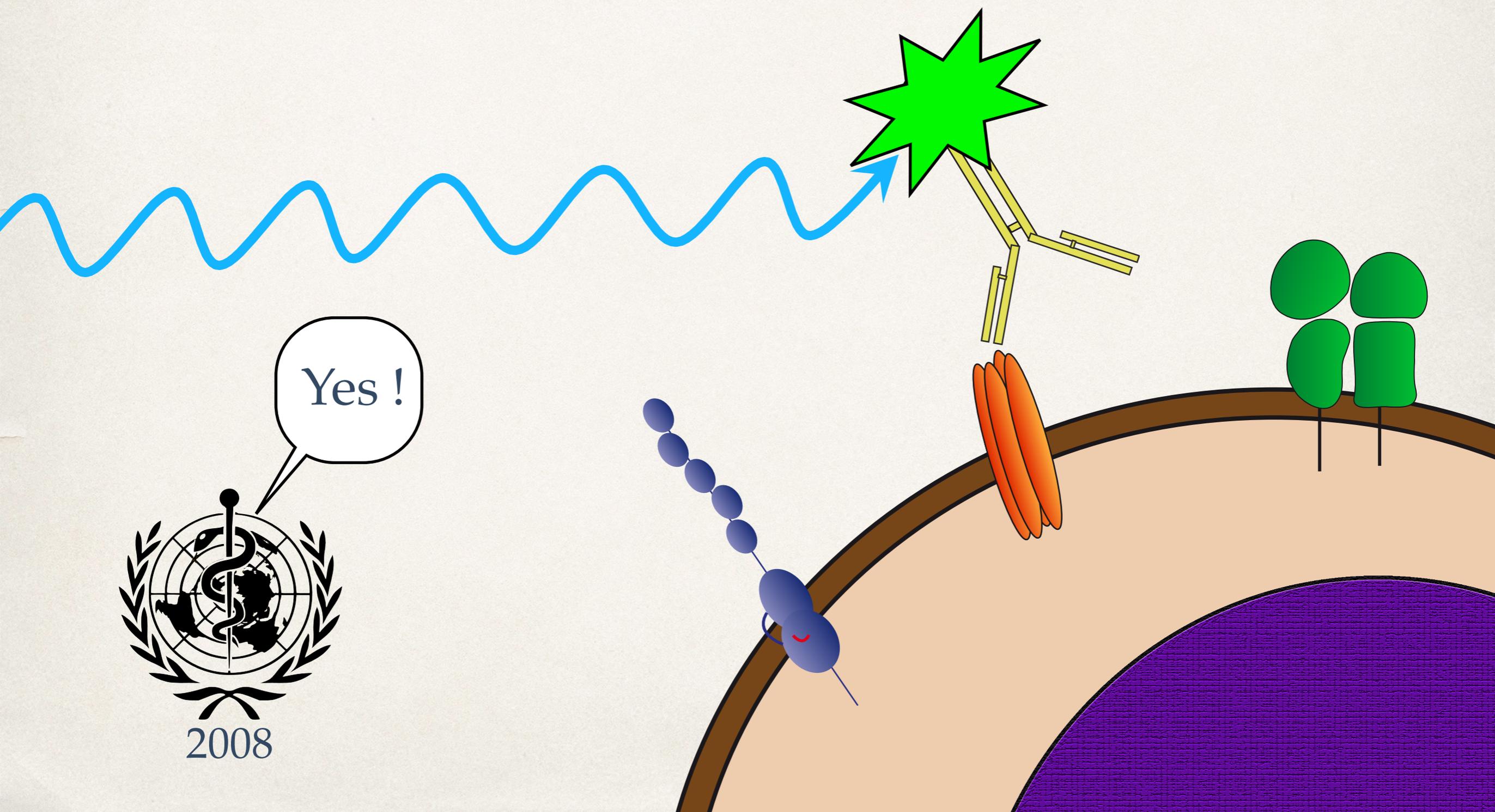


- Anomalies moléculaires dans les SMD sont moins bien caractérisées
- Ras, AML1, TP53, ...
- Intérêt pour les anomalies épigénétiques (méthylation des promoteurs, ...)

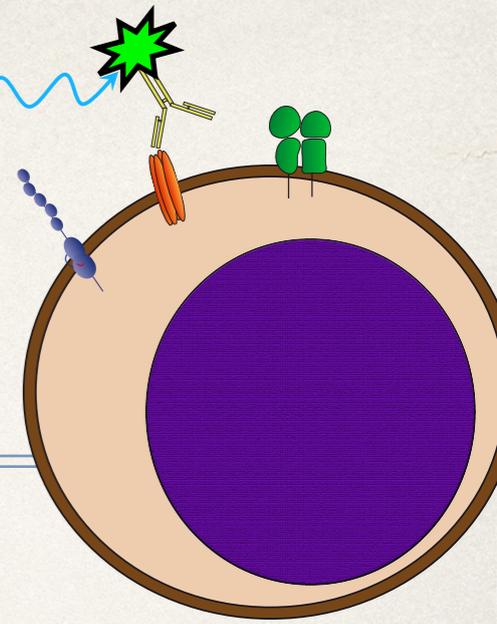
Pas toujours suffisant... particulièrement dans les  
SMD de bas grade !



# Est-ce que la CMF peut aider ?



# Cytométrie en flux dans les SMD



- ❖ Altération de l'expression des antigènes
- ❖ Evaluation du nombre de blastes et/ou détection de blastes à l'immunophénotype aberrant
- ❖ Identification de caractéristiques de dysmaturation sur les cellules de la lignée myéloïde.
- ❖ Exclusion d'autres pathologies
- ❖ Suivi, émergence de nouvelles populations pathologiques --> evolution ?

**WHO 2008** :  $\geq 3$  **aberrant features** by FC and borderline dysplasia by morphology and no cytogenetic abnormalities --> highly suggestive for MDS (--> reevaluation over several months). A single aberrant feature by FC is not enough.

Implementation of flow cytometry in the diagnostic work-up of myelodysplastic syndromes in a multicenter approach: Report from the Dutch Working Party on Flow Cytometry in MDS

Theresia M. Westers<sup>a,\*</sup>, Vincent H.J. van der Velde<sup>a</sup>, Rik A. Brooimans<sup>d</sup>, Claudia Cali<sup>a</sup>, Angelika M. Dräger<sup>a</sup>, Anja de Jong<sup>a</sup>, P. (Ellen) A. Kuiper-Kramer<sup>g</sup>, Marije Jeroen G. te Marvelde<sup>b</sup>, Joke K. van der Molen-Sinl<sup>a</sup>, Frank W.M.B. Preijers<sup>h</sup>, Roger K. Schindhelm<sup>g</sup>, Alit August H. Westra<sup>a</sup>, Arjan A. van de Loosdrecht<sup>a, c</sup>, MDS of the Dutch Society of Cytometry (NVC)

## Diagnostic utility of flow cytometry in low-grade myelodysplastic syndromes: a prospective validation study

Kiyoyuki Ogata,<sup>1</sup> Matteo G. Della Porta,<sup>2</sup> Luca Malcovati,<sup>2</sup> Cristina Picone,<sup>2</sup> Norio Yokose,<sup>3</sup> Akira Matsuda,<sup>4</sup> Taishi Yamashita,<sup>1,5</sup> Hideto Tamura,<sup>1</sup> Junichi Tsukada,<sup>6</sup> and Kazuo Dan<sup>1</sup>

## Myelodysplastic syndromes: the role of flow cytometry in diagnosis and prognosis

M. STETLER-STEVENSON, C. M. YUAN

Validation of a flow cytometric scoring system as a prognostic indicator for posttransplantation outcome in patients with myelodysplastic syndrome

Bart L. Scott,<sup>1,2</sup> Denise A. Wells,<sup>3</sup> Michael R. Loken,<sup>3</sup> David Myerson,<sup>1,2</sup> Wendy M. Leisenring,<sup>1,2</sup> and H. Joachim Deeg<sup>1,2</sup>

## Diagnostic flow cytometry for low-grade myelodysplastic syndromes

Kiyoyuki Ogata\*

Division of Hematology, Department of Medicine, Nippon Medical School, Tokyo, Japan

## Standardization of flow cytometry in myelodysplastic syndromes: report from the first European LeukemiaNet working conference on flow cytometry in myelodysplastic syndromes

Arjan A. van de Loosdrecht,<sup>1</sup> Canan Alhan,<sup>1</sup> Marie Christine Béné,<sup>2</sup> Matteo G. Della Porta,<sup>3</sup> Angelika M. Dräger,<sup>4</sup> Leon Fouillard,<sup>4</sup> Patricia Font,<sup>5</sup> Ulrich Germing,<sup>6</sup> Detlef Haase,<sup>7</sup> Christa H. Homburg,<sup>8</sup> Robin Ireland,<sup>9</sup> Luca Malcovati,<sup>3</sup> Jeroen G. te Marvelde,<sup>12</sup> Ghulam J. Mufti,<sup>9</sup> Kiyoyuki Ogata,<sup>13</sup> A. Porwit,<sup>15</sup> Frank W. Preijers,<sup>10</sup> Stephen J. Richards,<sup>16</sup> Gerrit Jan Schuurman,<sup>18</sup> Vincent H.J. van der Velden,<sup>12</sup> Paresh Vyas,<sup>19</sup> August H. Westra,<sup>1</sup> Michael R. Loken,<sup>20</sup> and Theresia M. Westers<sup>1</sup>

## Myeloid and monocytic dyspoiesis as determined by flow cytometric scoring in myelodysplastic syndrome correlates with the IPSS and with outcome after hematopoietic stem cell transplantation

Denise A. Wells, Martin Benesch, Michael R. Loken, Carlos Vallejo, David Myerson, Wendy M. Leisenring, and H. Joachim Deeg

## Bone Marrow Cells from Myelodysplastic Syndromes Show Altered Immunophenotypic Profiles That May Contribute to the Diagnosis and Prognostic Stratification of the Disease: a Pilot Study on A Series of 56 Patients

Sergio Matarraz,<sup>1</sup> Antonio López,<sup>1</sup> Susana Barrena,<sup>1</sup> Carlos Fernandez,<sup>1</sup> Evan Jensen,<sup>1</sup> Juan Flores-Montero,<sup>1</sup> Ana Rasillo,<sup>1</sup> José María Sayagues,<sup>1</sup> María Luz Sánchez,<sup>1</sup> Paloma Bárcena,<sup>1</sup> Jesús María Hernández-Rivas,<sup>2</sup> Carlos Salvador,<sup>3</sup> Nuria Fernandez-Mosteirín,<sup>3</sup> Manuel Giralt,<sup>3</sup> Luis Perdiguer,<sup>4</sup> Paula Laranjeira,<sup>5</sup> Artur Paiva,<sup>5</sup> and Alberto Orfao<sup>1\*</sup>

<sup>1</sup>Centro de Investigación del Cáncer (Instituto de Biología Molecular y Celular del Cáncer; CSIC-USAL), Servicio General de Citometría and Departamento de Medicina, Universidad de Salamanca, Salamanca, Spain

<sup>2</sup>Servicio de Hematología, Hospital Universitario de Salamanca, Salamanca, Spain

<sup>3</sup>Servicio de Hematología, Hospital Miguel Servet, Zaragoza, Spain

<sup>4</sup>Servicio de Hematología, Hospital de Alcañiz, Teruel, Spain

<sup>5</sup>Unidade de Citometria, Centro de Histocompatibilidade do Centro-Coimbra, Coimbra, Portugal

## Flow cytometric scoring system as a diagnostic and prognostic tool in myelodysplastic syndromes

Sung-Chao Chu<sup>a,b</sup>, Tso-Fu Wang<sup>a</sup>, Chi-Cheng Li<sup>d</sup>, Ruey-Ho Kao<sup>a,c</sup>, Dian-Kun Li<sup>c,f</sup>, Yu-Chieh Su<sup>c,f</sup>, Denise A. Wells<sup>e</sup>, Michael R. Loken<sup>e,\*</sup>

# Comités internationaux Standardisation Guidelines

Ident  
myel  
Ar My  
ar my  
hen  
Denise  
my  
Sun  
Denise A. Wells<sup>e</sup>, ]

<sup>e</sup>Unidade de Citometria, Centro de Histocompatibilidade do Centro-Coimbra, Coimbra, Portugal

-risk

r Velden,<sup>2</sup>

n

in  
nieh Su<sup>c,f</sup>,

2000

2003

2005

2008

2009

2010

2013

2015

?



# Hématopoïèse normale dans la moelle

*Immunophenotypic Differentiation Patterns of Normal Hematopoiesis in Human Bone Marrow : Reference Patterns for Age-Related Changes and Disease-Induced Shifts, EG van Lochem et al, Cytometry Part B (Clinical Cytometry) 60B:1-13 (2004)*

---

❖ **Différenciation des granulocytes**

CD34 / CD117 / CD45 / CD13-33  
CD16 / CD13 / CD45 / CD11b

❖ Différenciation monocytaire

CD14 / CD33 / CD45 / CD34

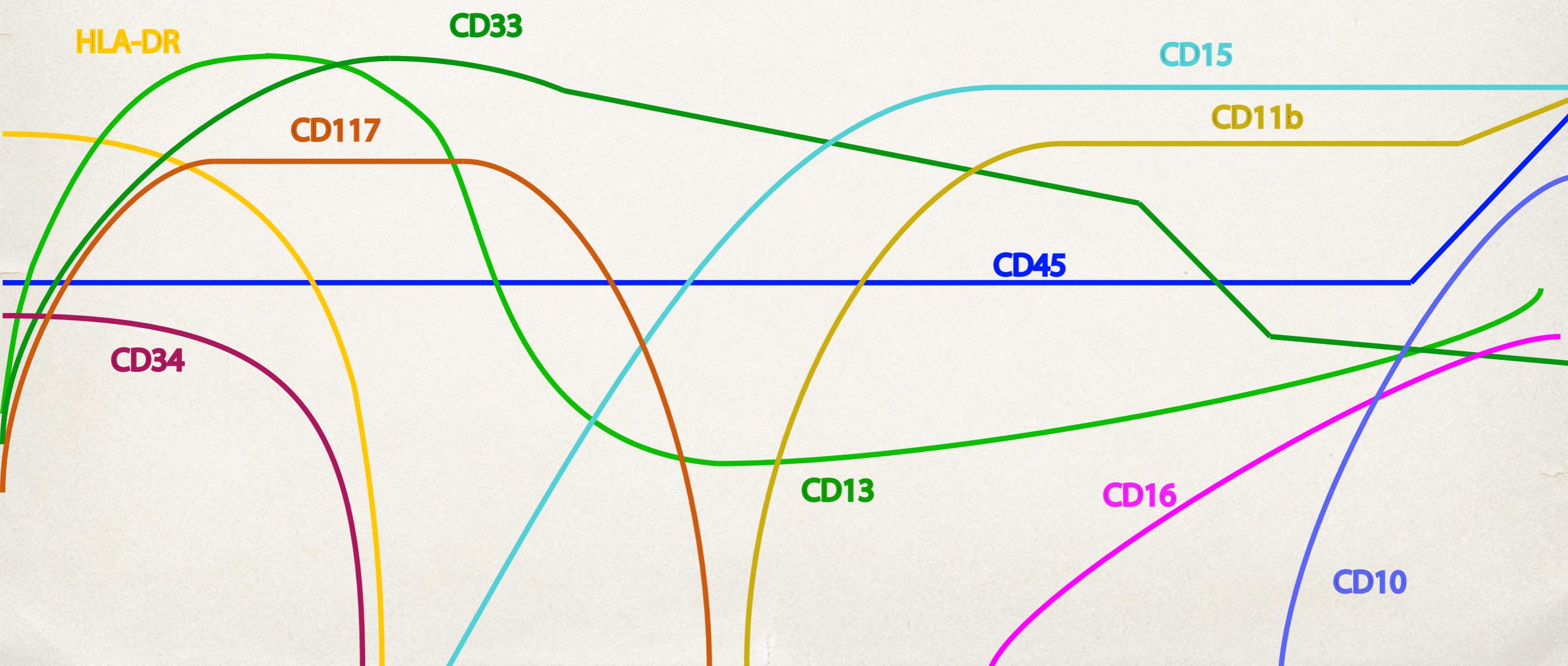
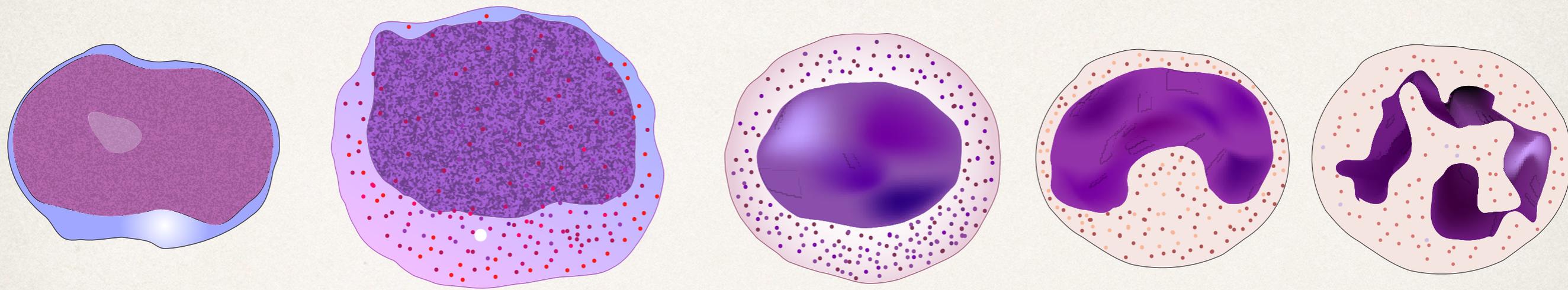
❖ Différenciation cellules B

TdT / CD20 / CD19 / CD10  
CD45 / CD34 / CD19 / CD22

❖ Différenciation érythroïde

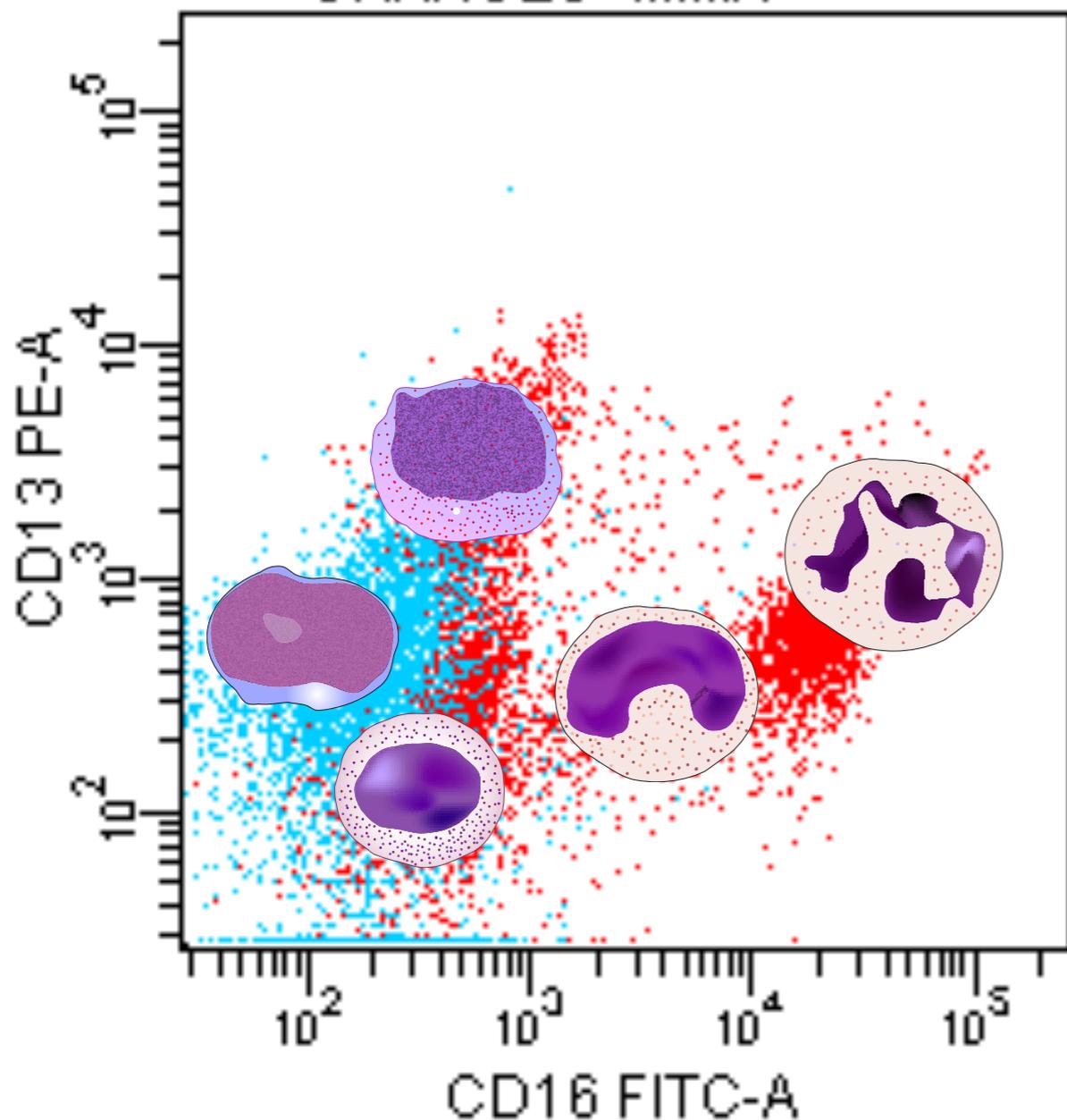
CD71 / CD235a / CD45 / CD34

# Maturation myéloïde

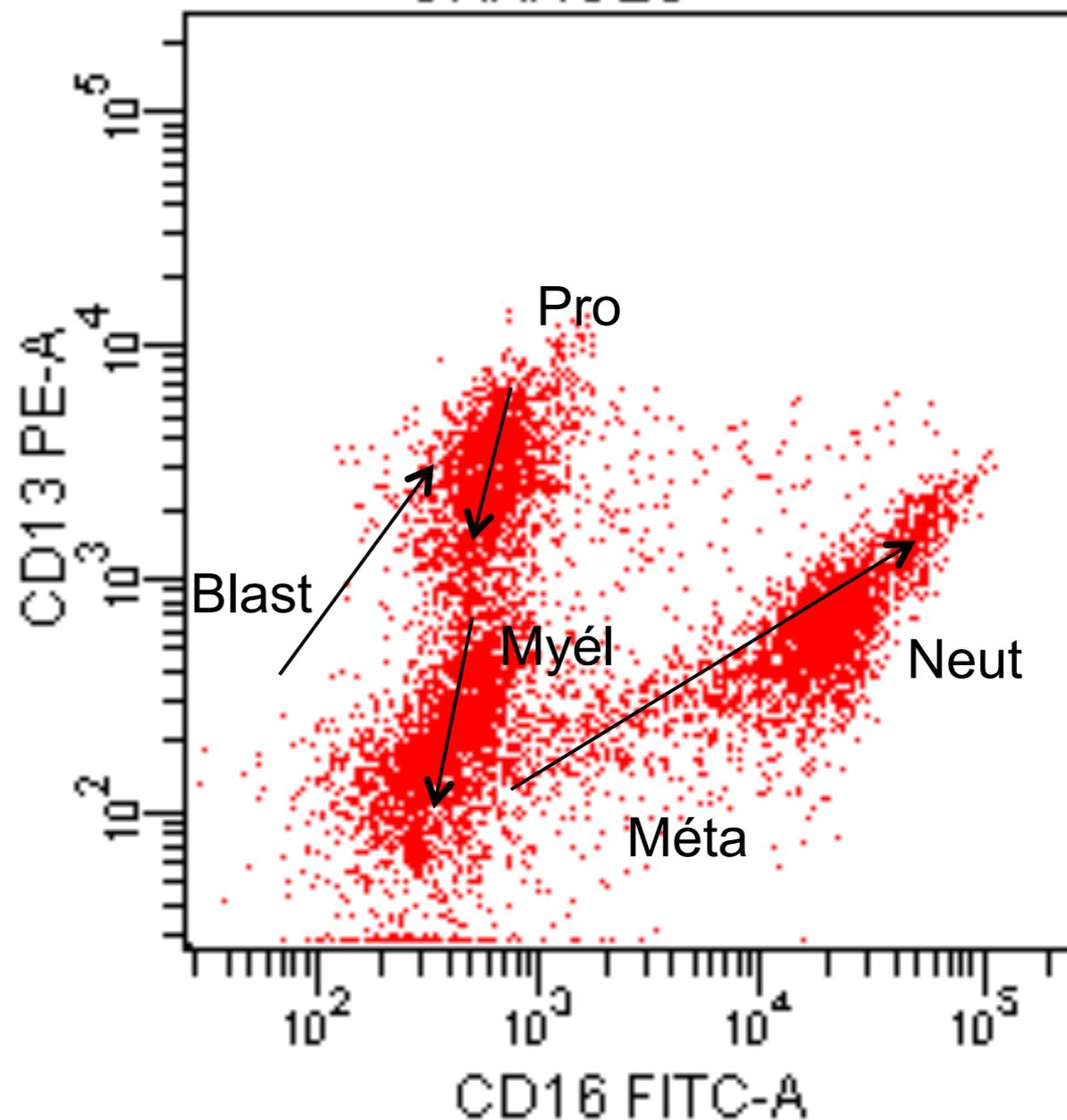


# Différenciation myéloïde normale

GRANULO+IMMA



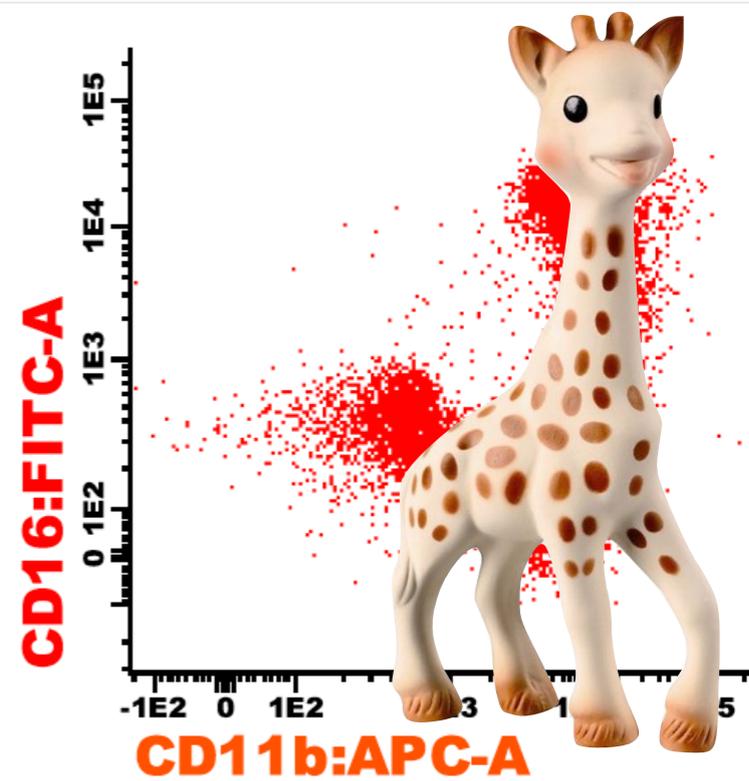
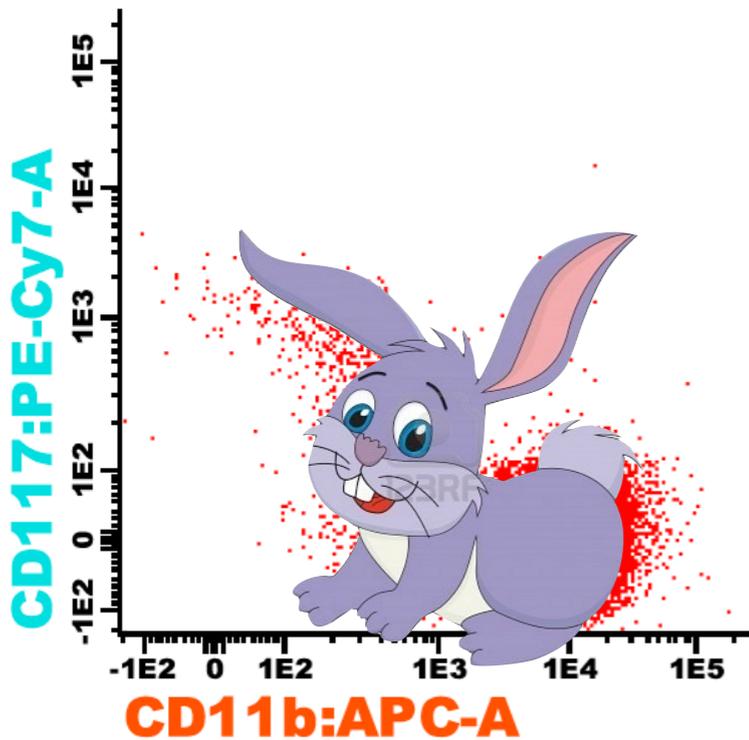
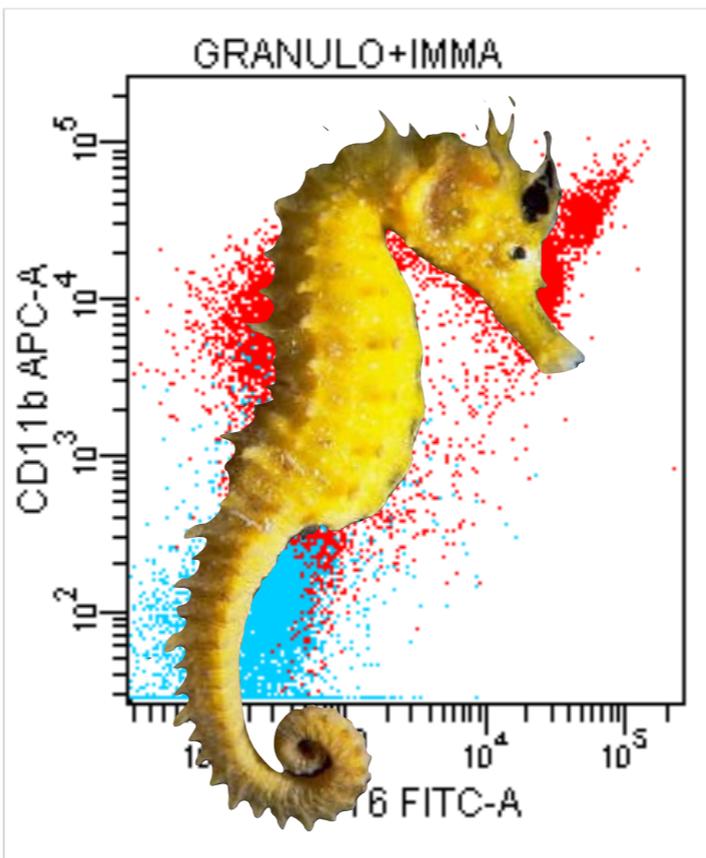
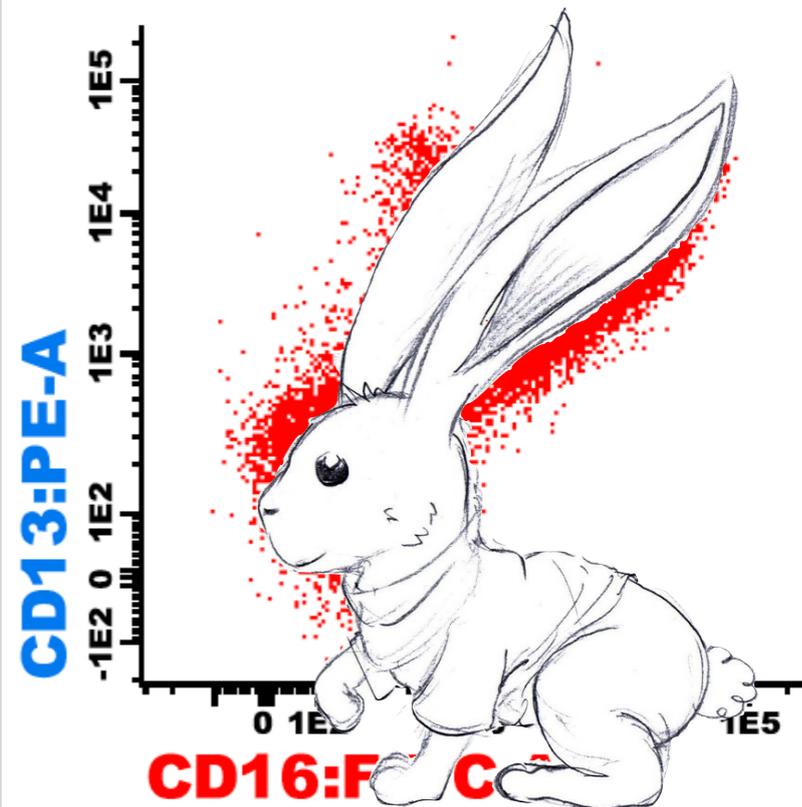
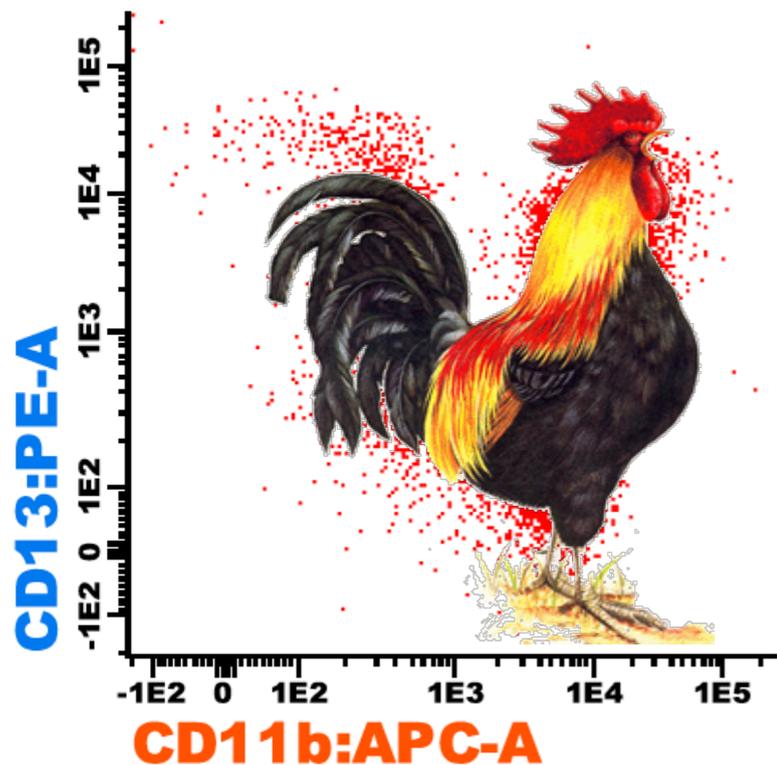
GRANULO



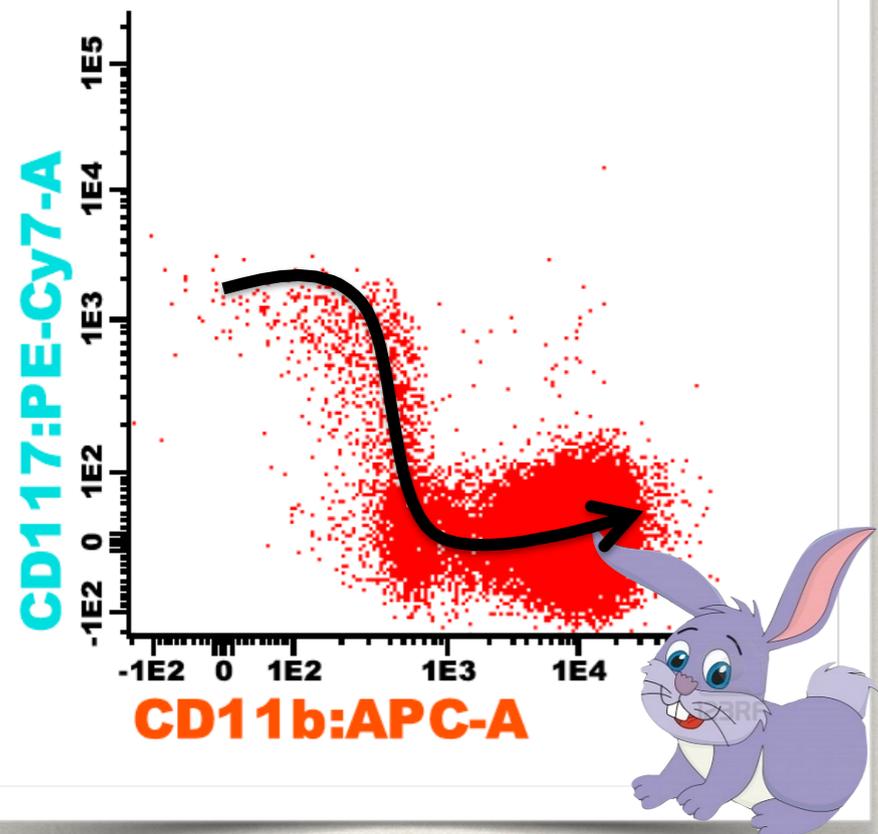
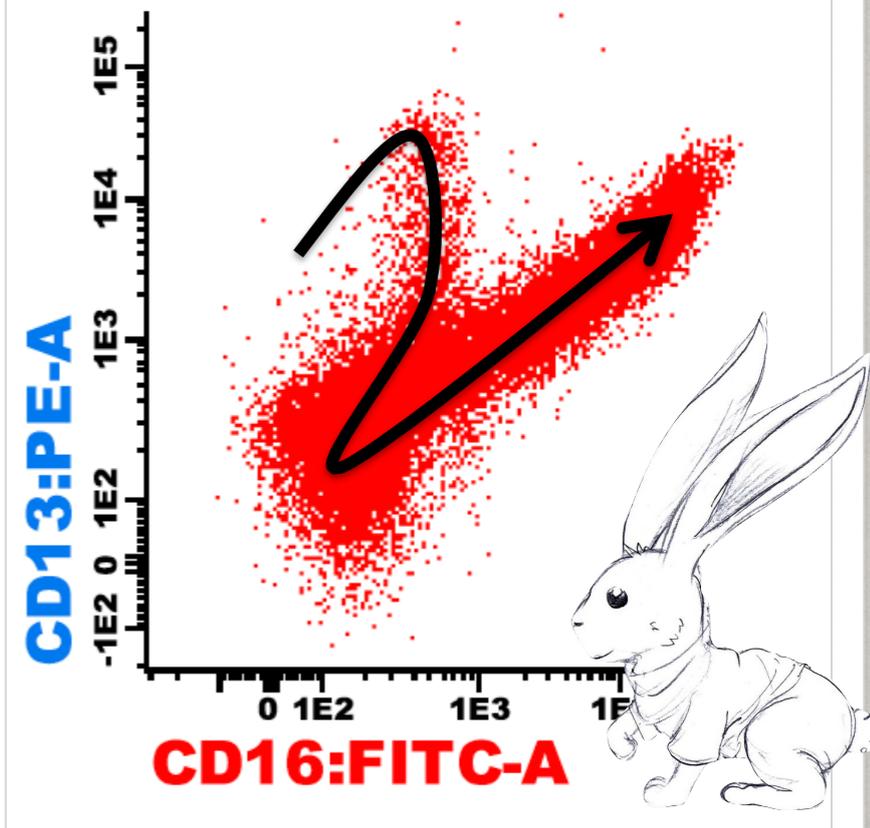
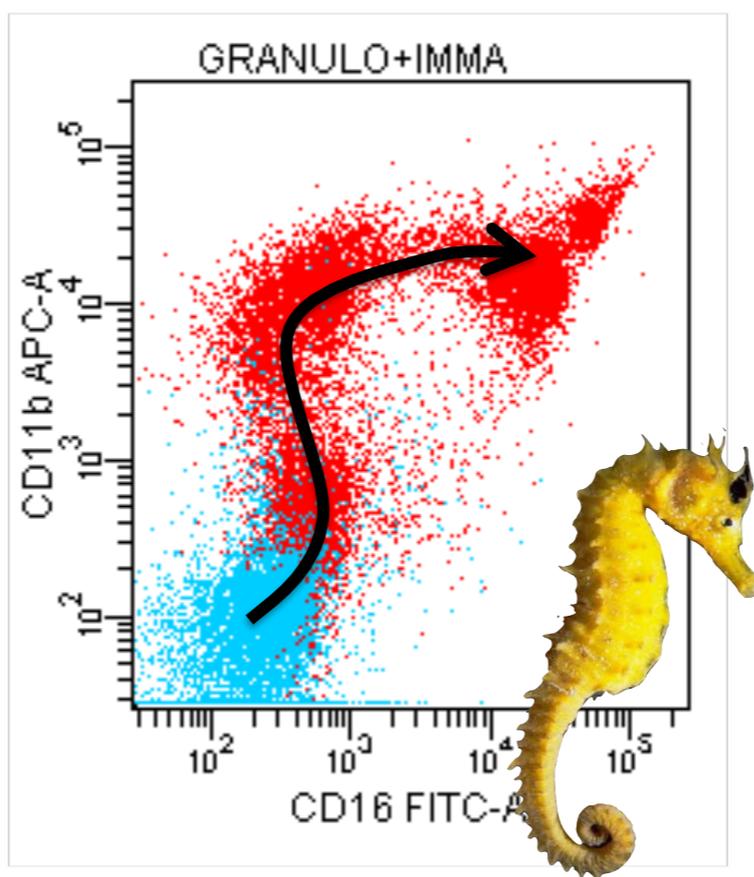
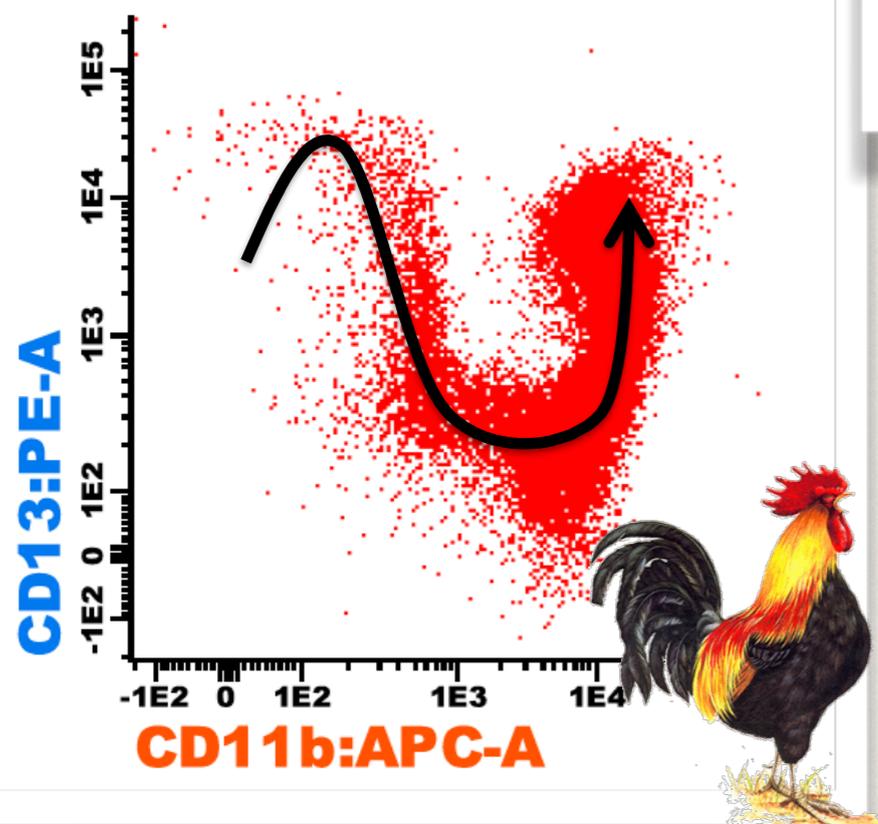
# Différenciation myéloïde normale



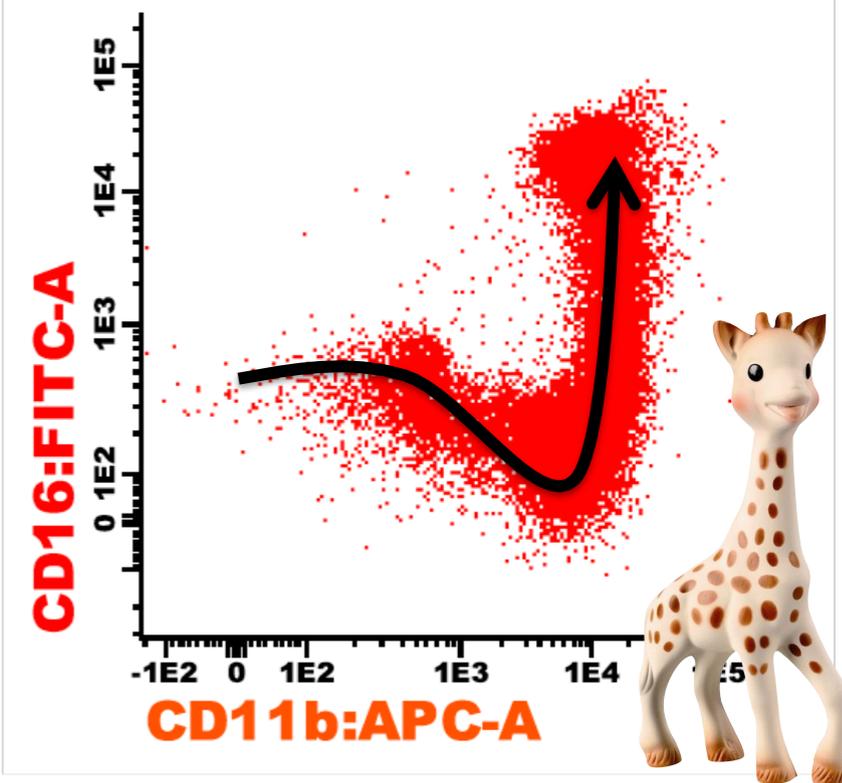
CD13/CD16/CD11b/CD117



CD13/CD16/CD11b/CD117



Connaître ces arcs permet de traquer les anomalies !





*La maturation a du bon...*

↓ SSC des neutrophiles

↑ % CD34+

Niveau d'expression anormal  
de marqueurs

# Que se passe-t-il dans un SMD ?

Expression d'un marqueur  
aberrant

↓ Progéniteurs B  
(parmi les CD34+)

Expression asynchrone de  
marqueurs de maturation



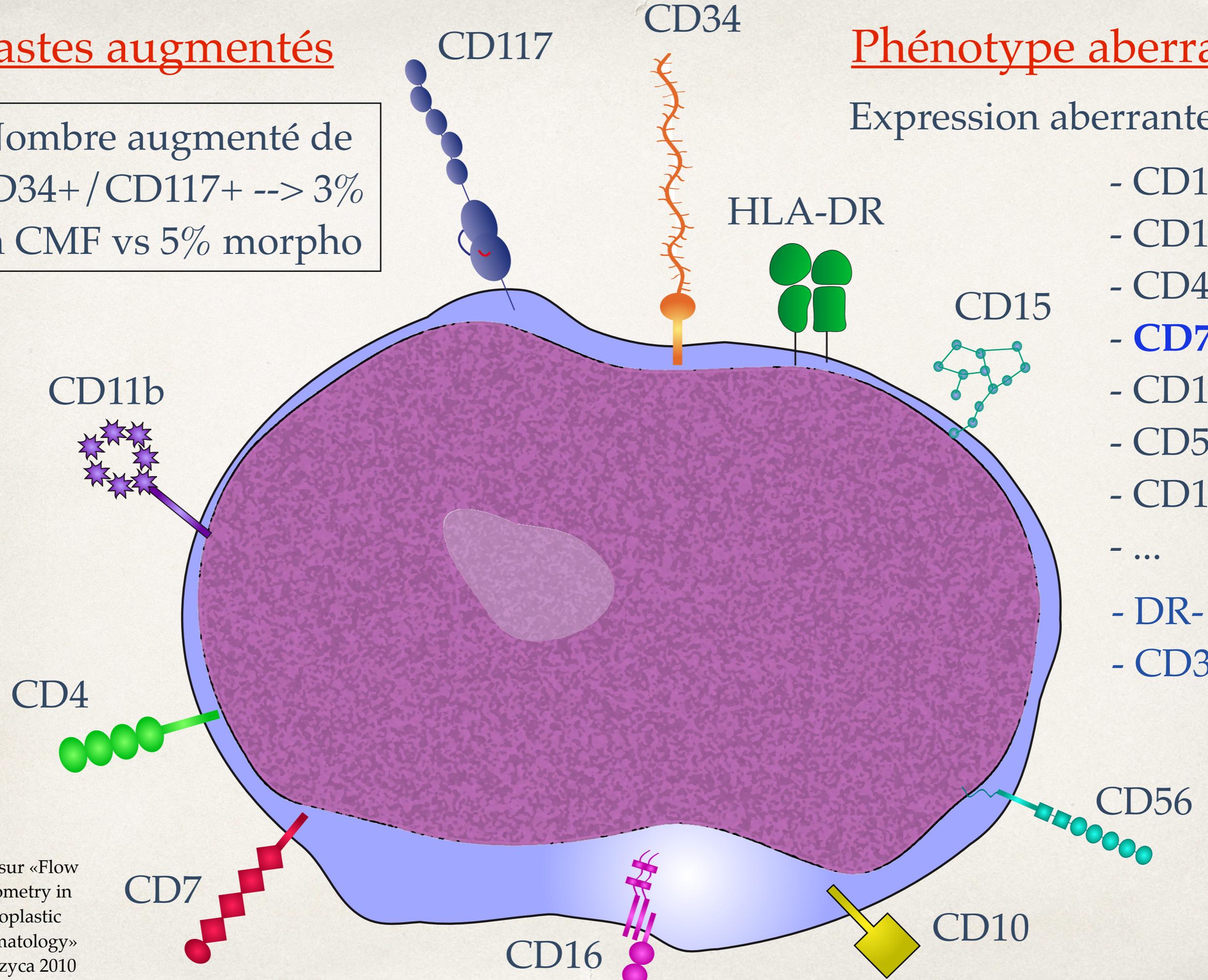
# Blastes augmentés

Nombre augmenté de CD34+ / CD117+ --> 3% en CMF vs 5% morpho

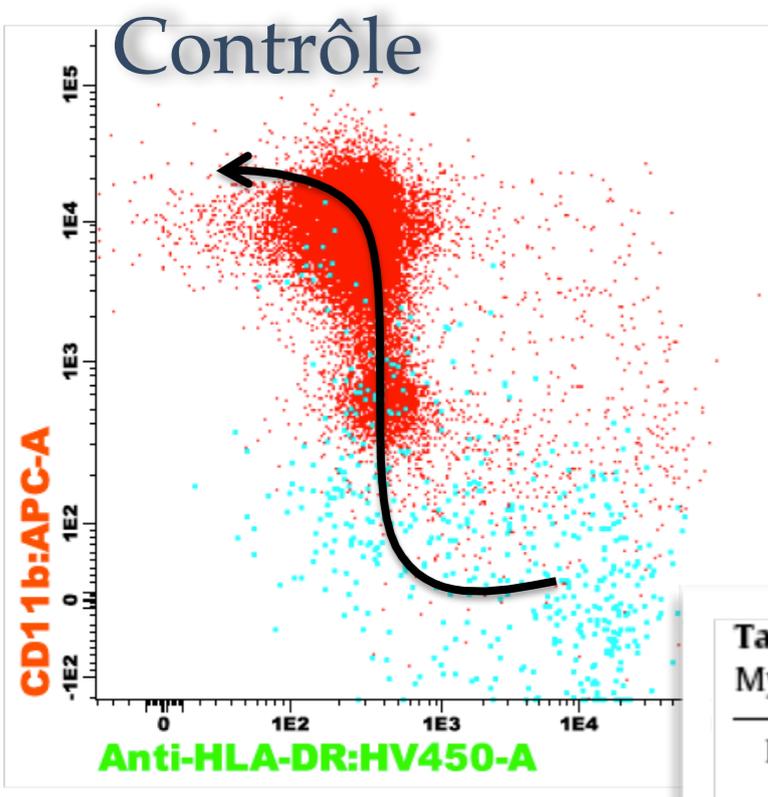
# Phénotype aberrant

Expression aberrante de

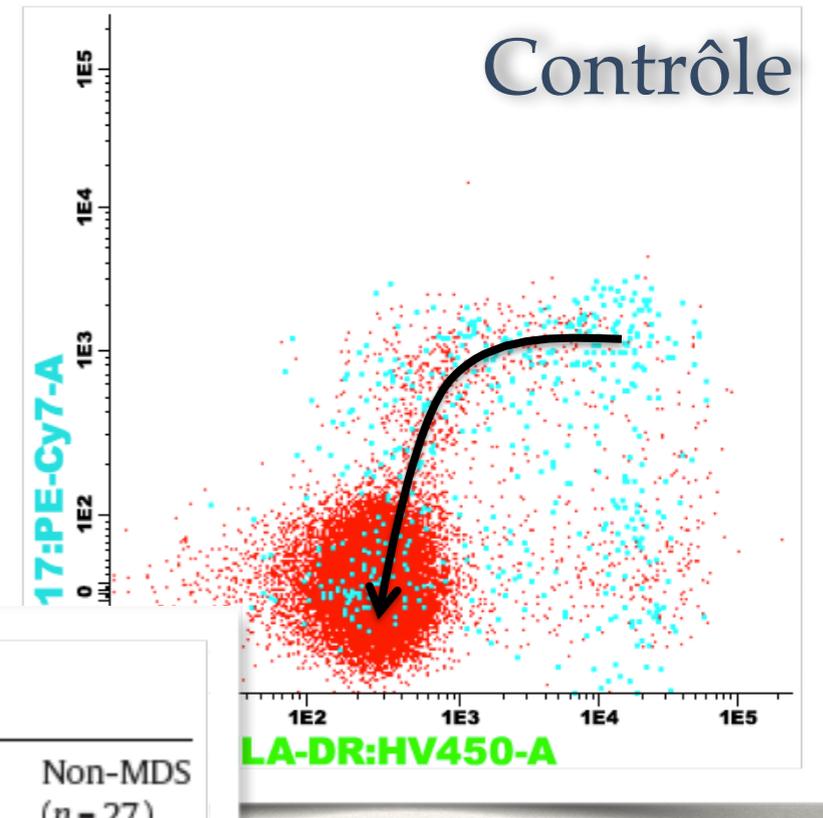
- CD11b
- CD15
- CD4
- **CD7**
- CD10
- CD56
- CD16
- ...
- DR-
- CD38-



Basé sur «Flow Cytometry in Neoplastic Haematology» Gorczyca 2010



Gate : IMMA + granulo

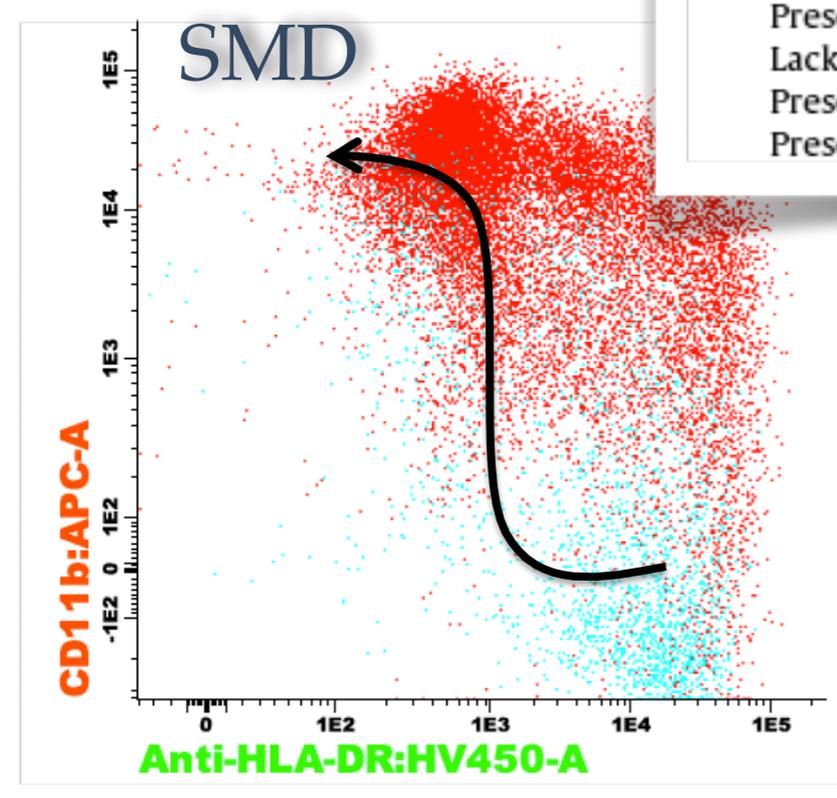


**Table 3**  
Myeloid and monocytic abnormalities detected by flow cytometry.

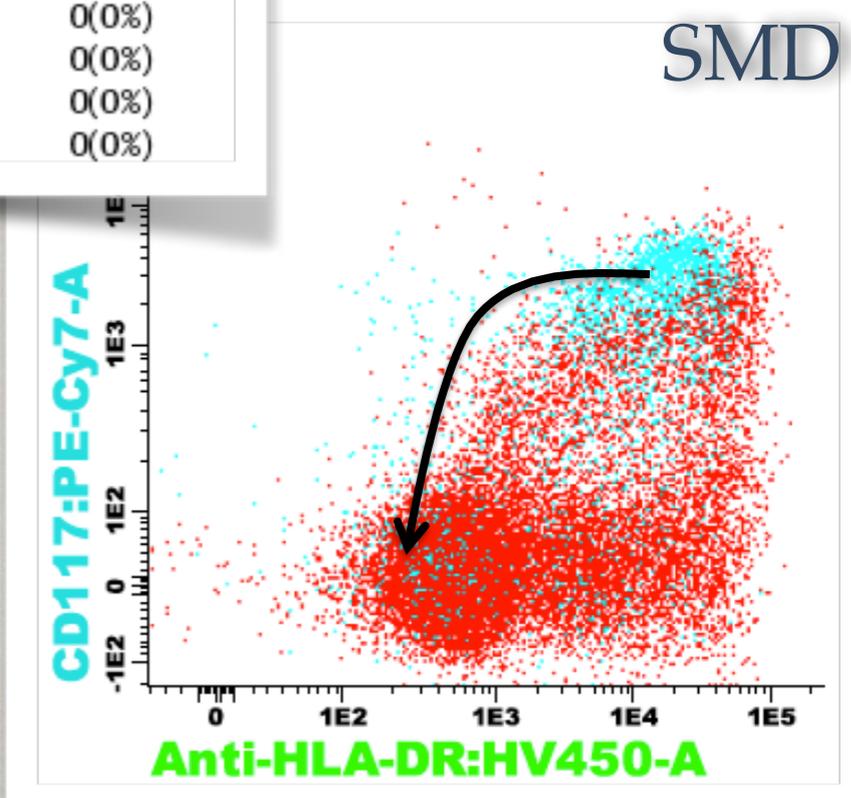
Flow cytometric abnormality	MDS (n = 56)	Non-MDS (n = 27)
Abnormal maturing myeloid, no. (%) <sup>a</sup>		
Abnormal granularity	9(16%)	0(0%)
Abnormal decrease CD45	3(5%)	0(0%)
Abnormal relationship CD13/CD16	18(32%)	2(7%)
Abnormal relationships HLA-DR/CD11b	14(25%)	0(0%)
Asynchronous shift to the left	11(20%)	0(0%)
Presence of CD56	13(23%)	0(0%)
Lack of CD33	0(0%)	0(0%)
Presence of CD34	2(4%)	0(0%)
Presence of lymphoid antigens	0(0%)	0(0%)

HLA-DR / CD11b

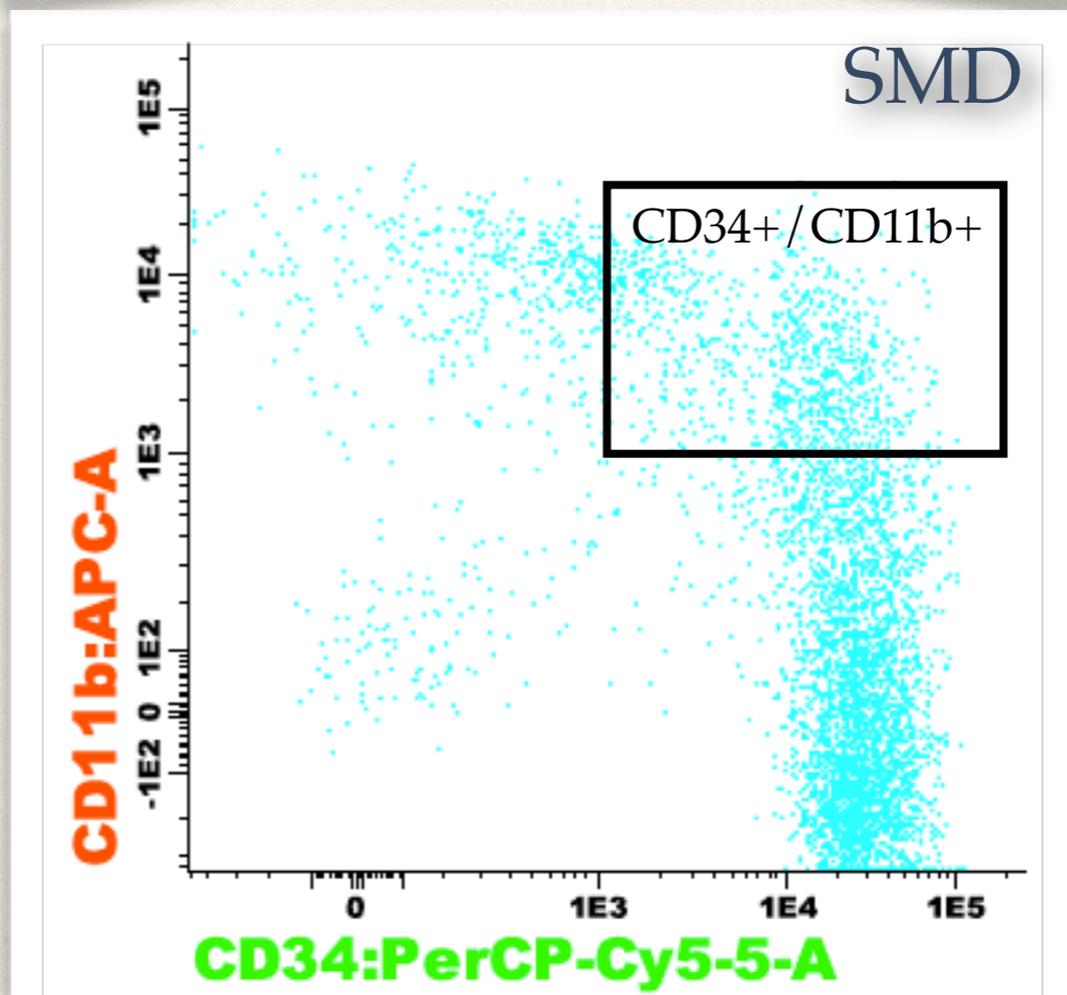
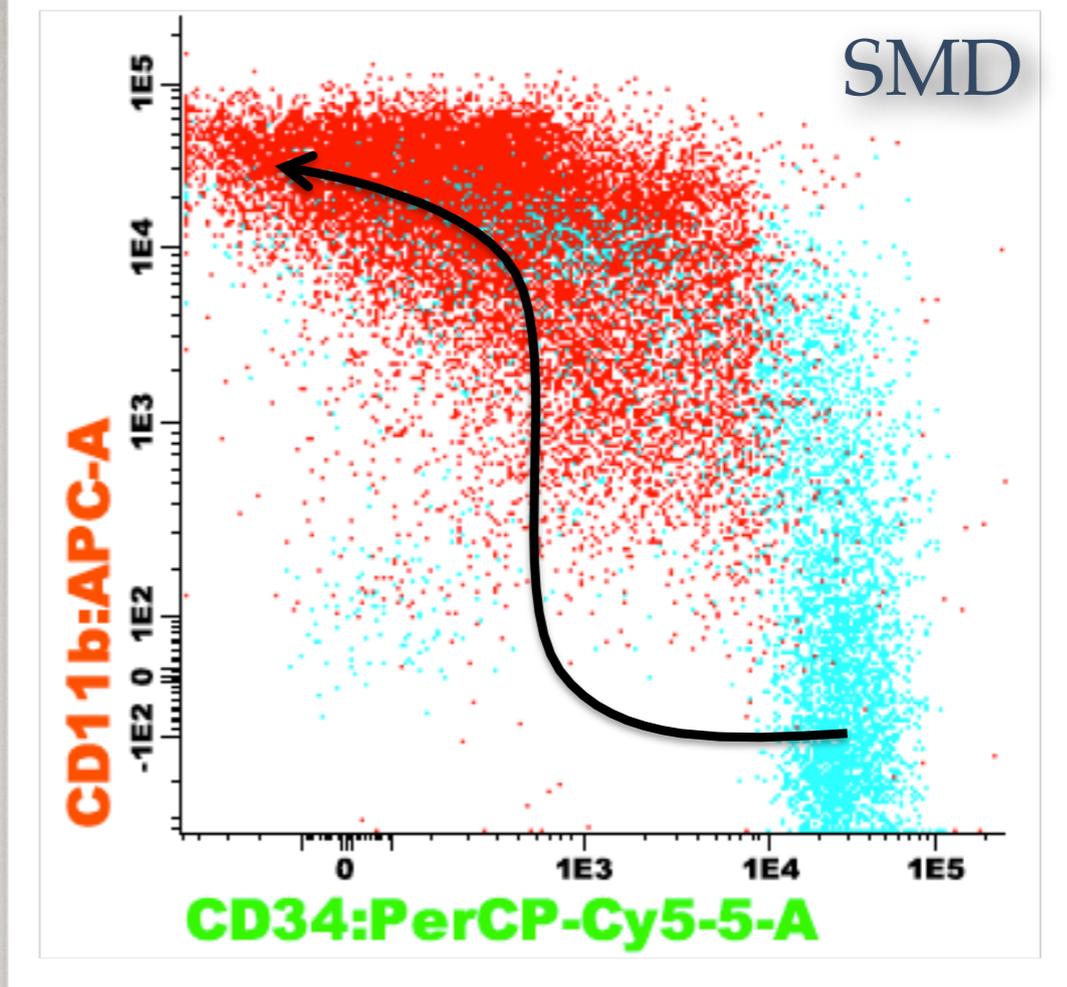
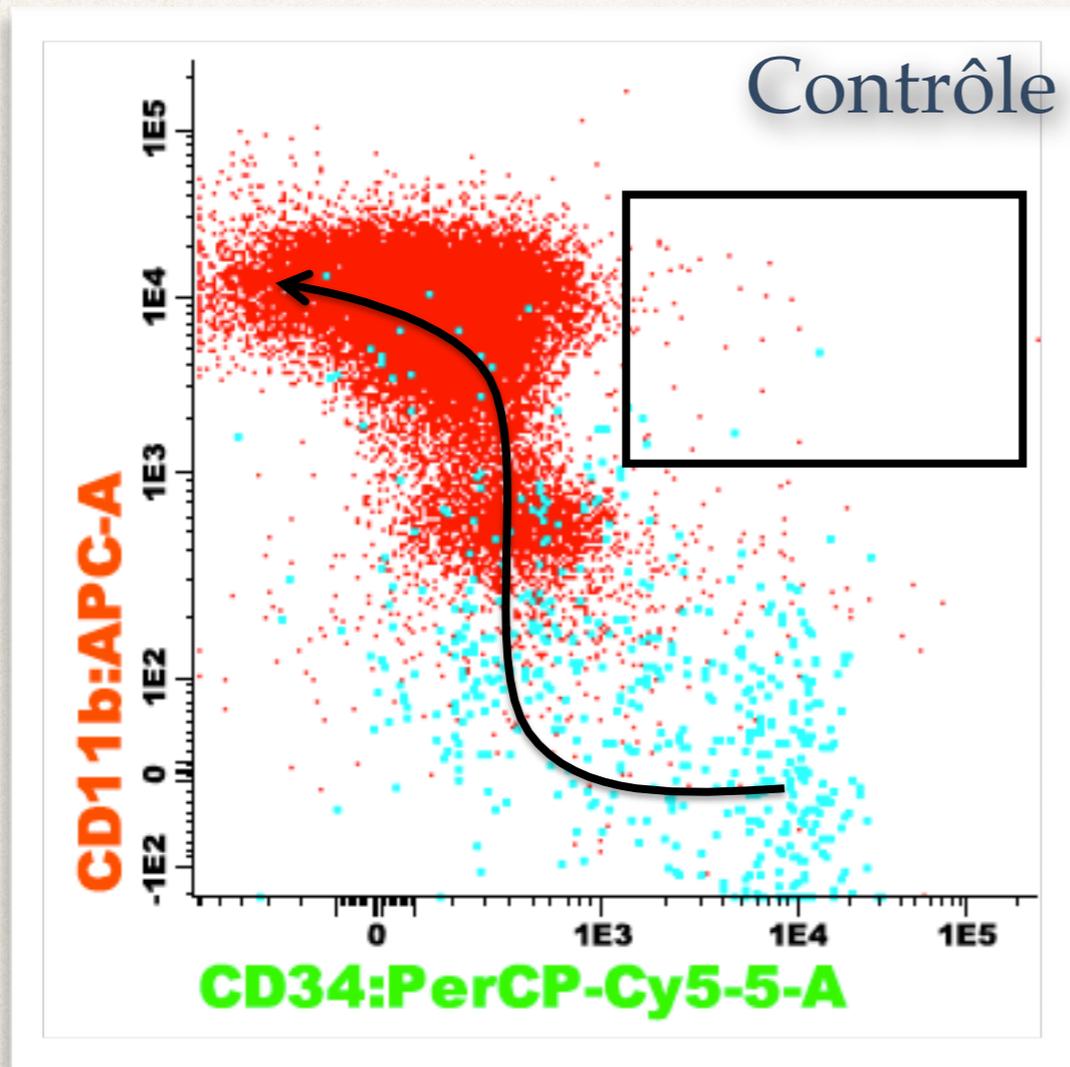
HLA-DR / CD117



*Flow Cytometric scoring system as a diagnostic and prognostic tool in myelodysplastic syndromes, Chu et al, Leukemia Research 35 (2011) 868-873*



CD34 / CD11b



Pas toujours aussi clair !  
En particulier dans les  
formes intermédiaires...

# Phénotype aberrant des blastes ex : CD33 - CD7

«min. 10%»

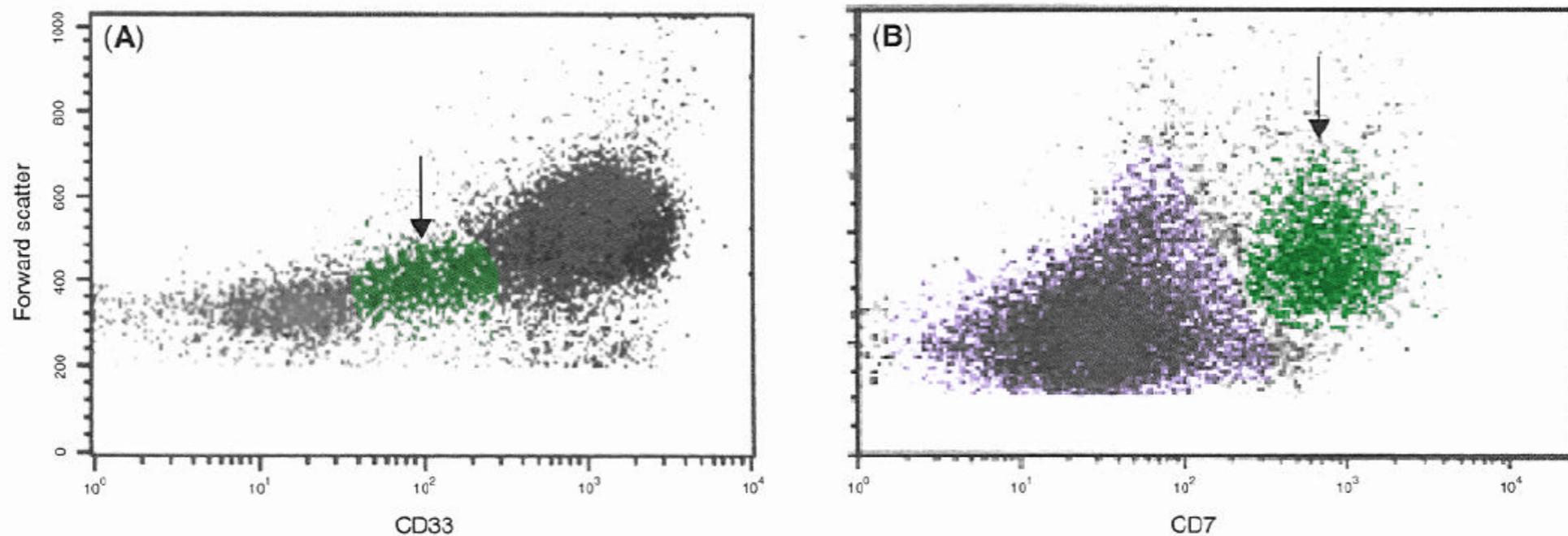
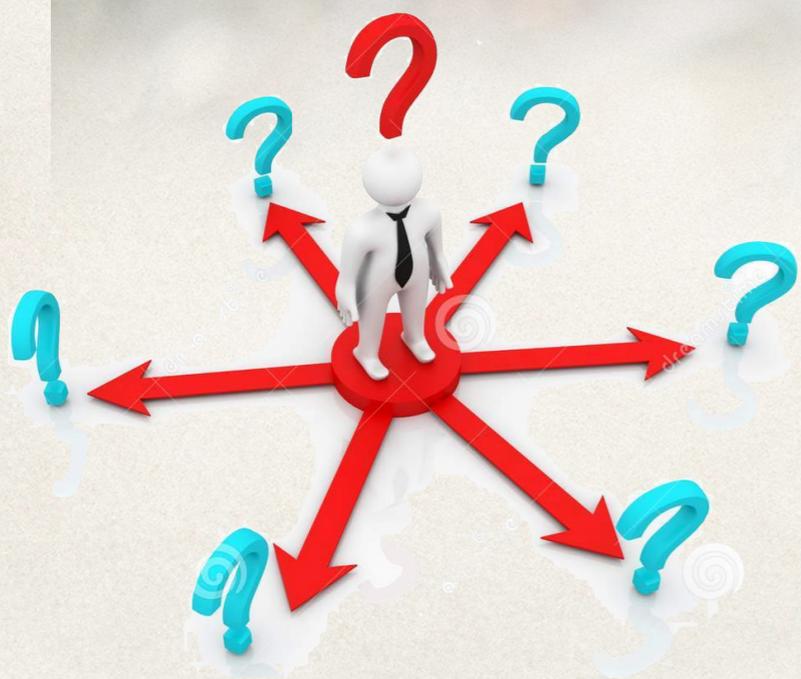
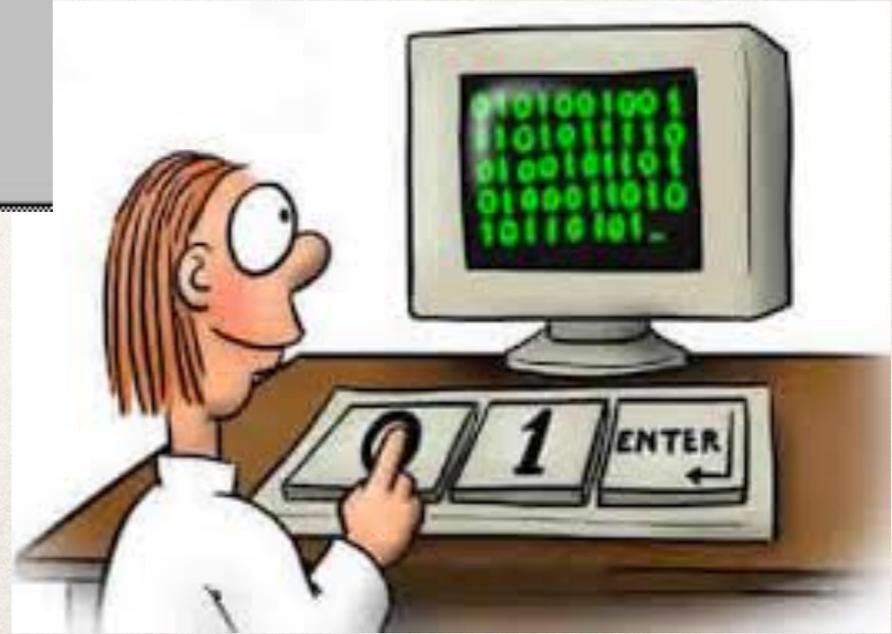
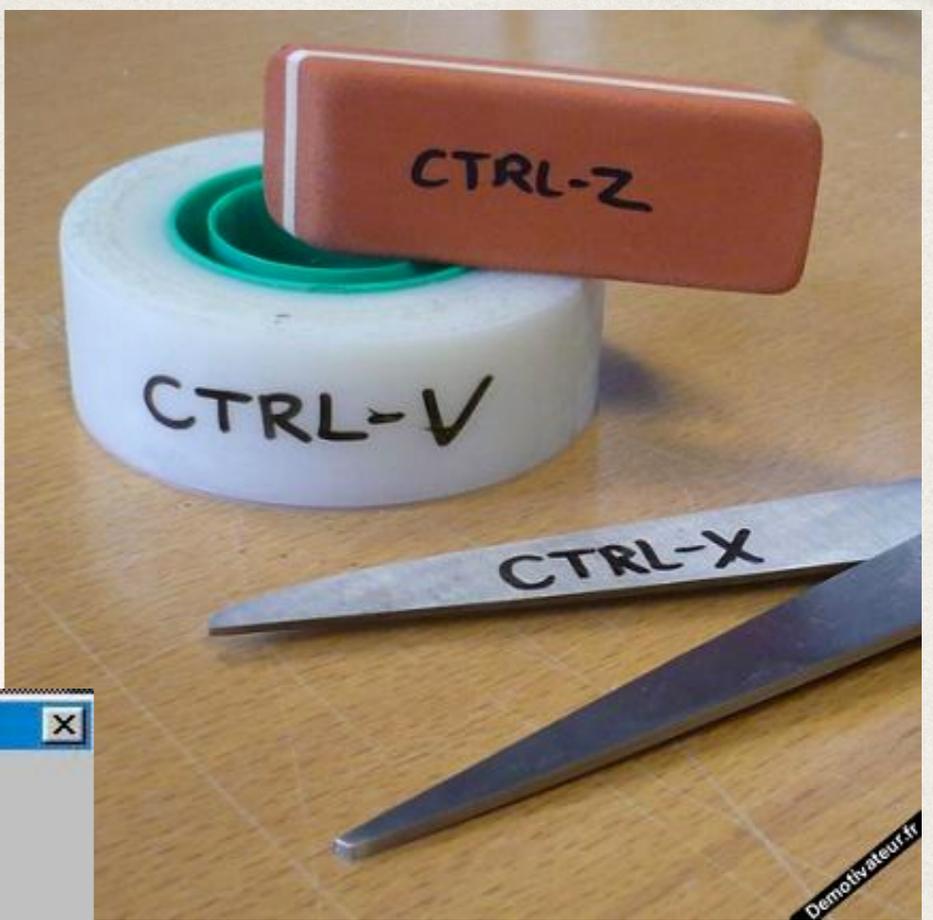
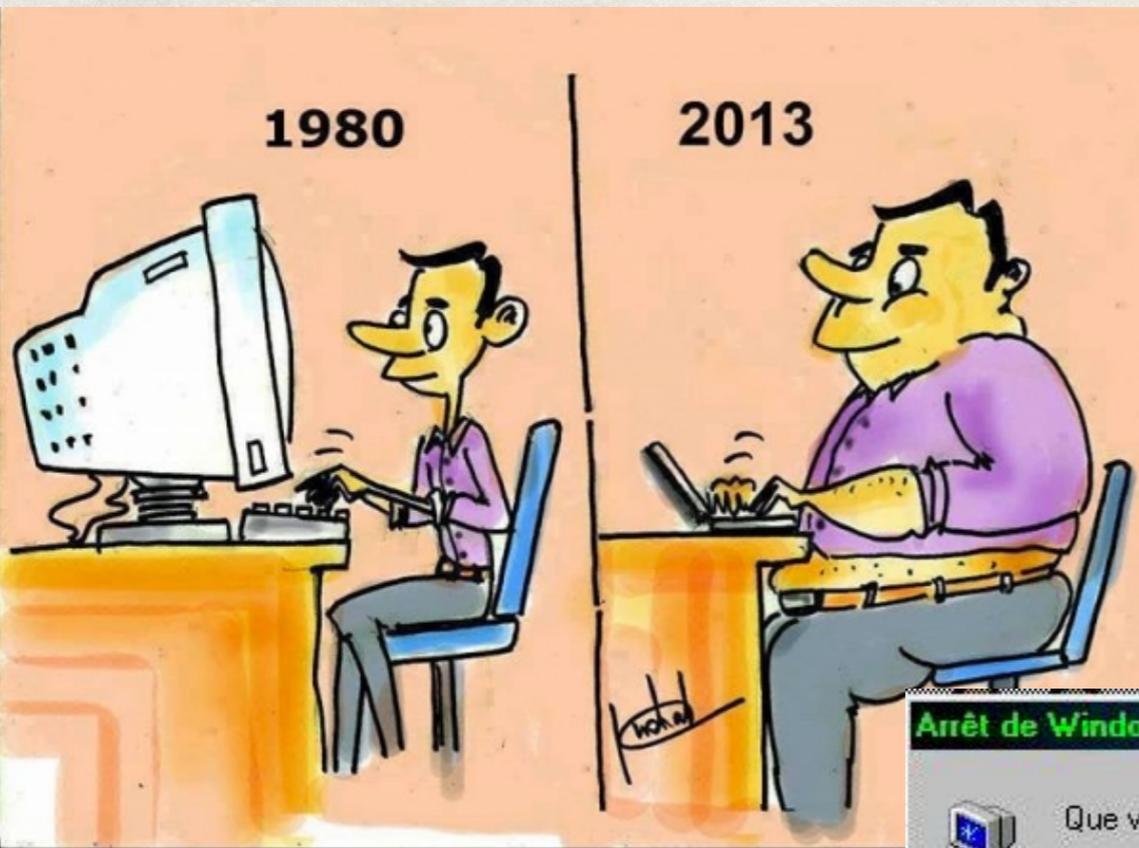


Figure 6.13 Aberrant phenotype of blasts in myelodysplastic syndrome (two cases). A shows blasts with aberrant (dim) expression of CD33 (arrow) and B shows blasts with aberrant expression of CD7 (arrow).

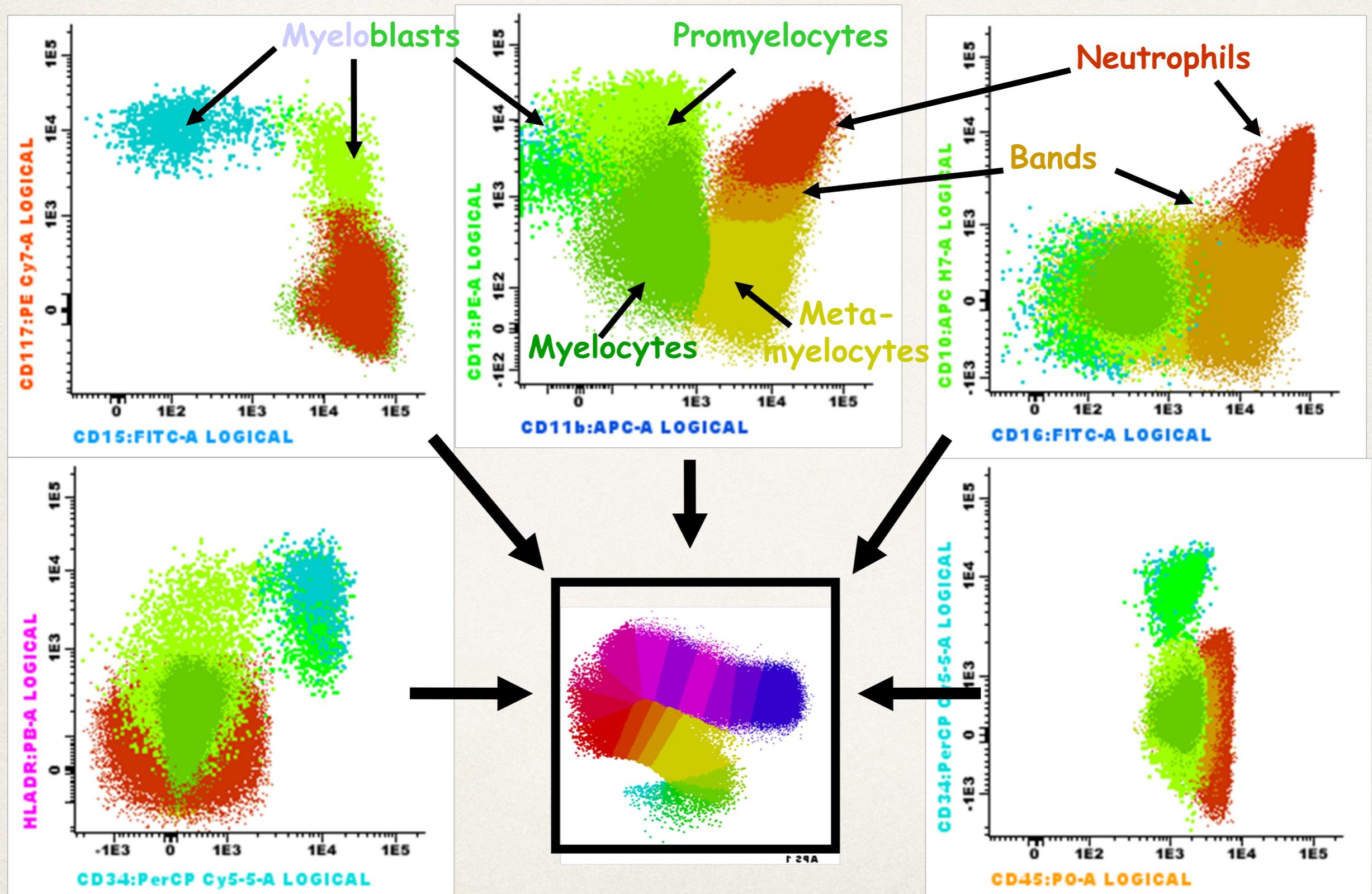
*Marqueurs d'infidélité de lignées : CD5, CD7, CD19, CD56...*

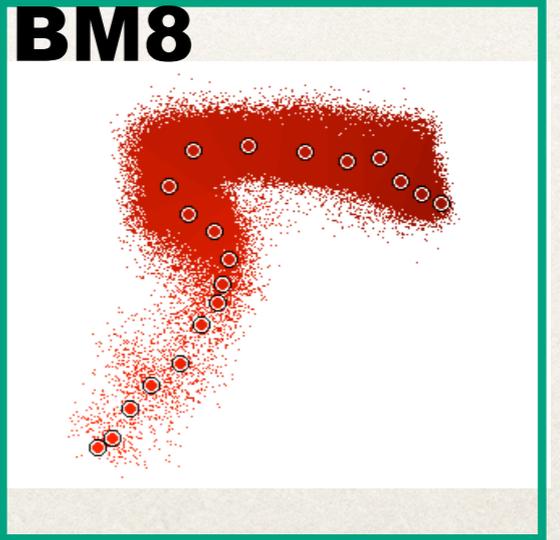
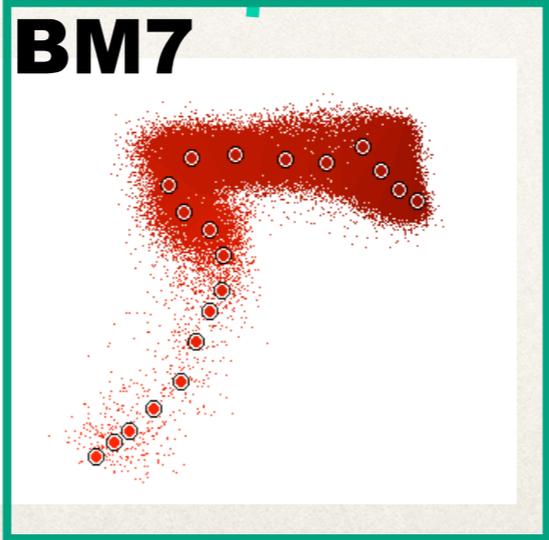
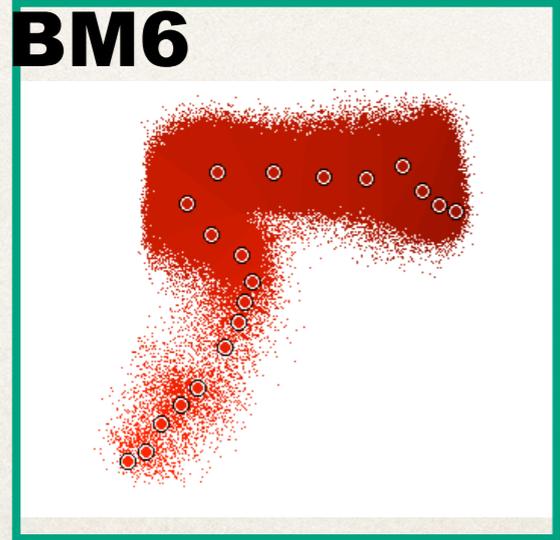
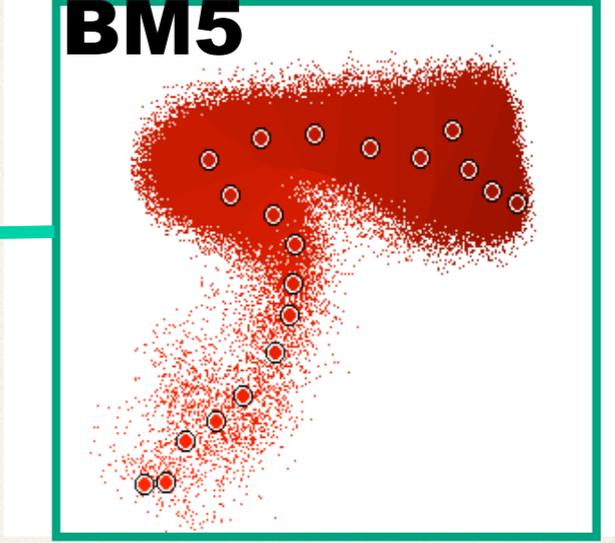
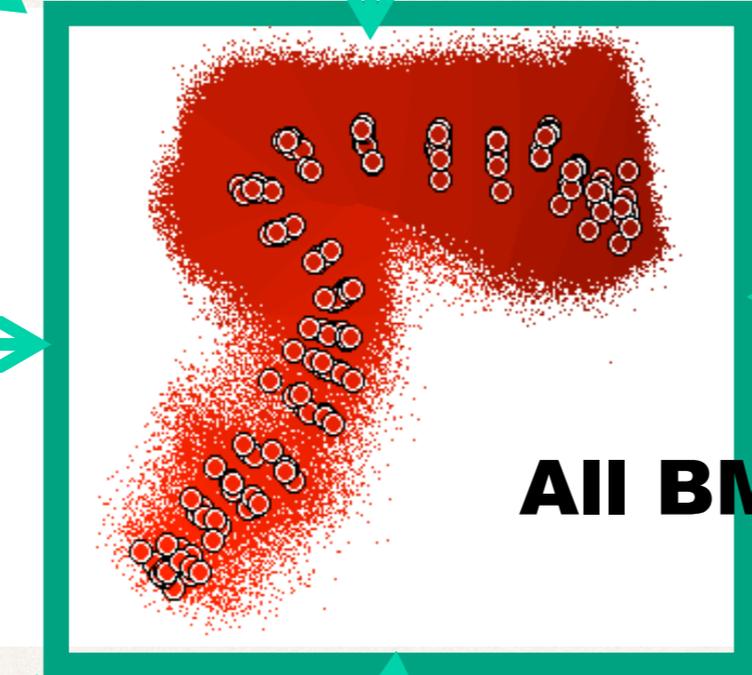
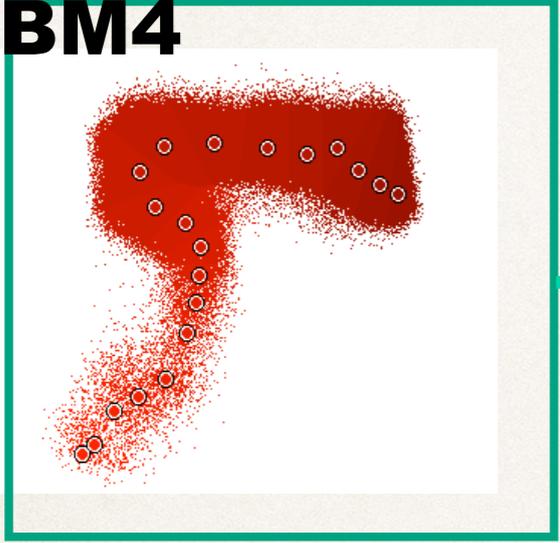
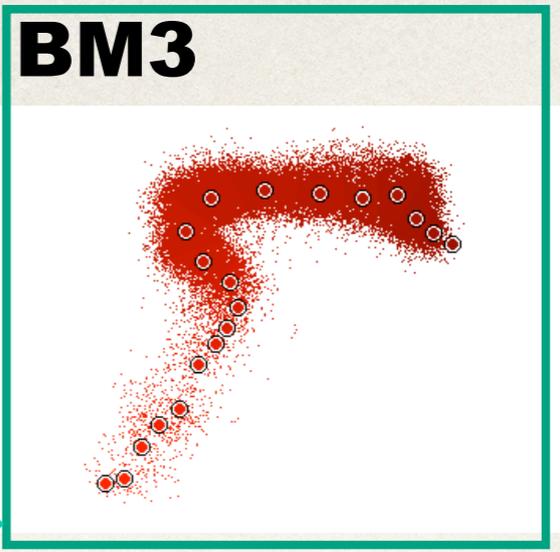
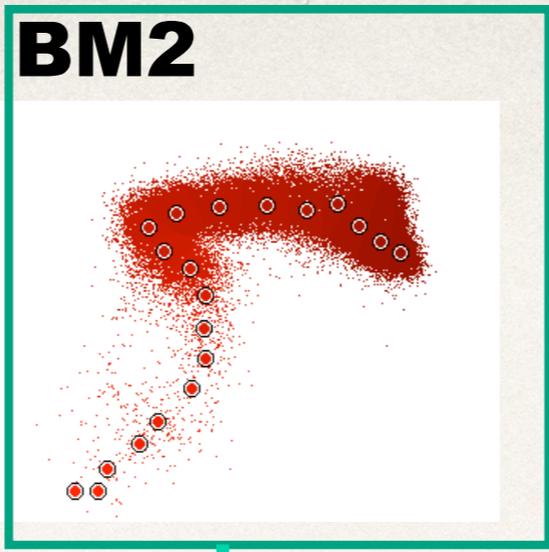
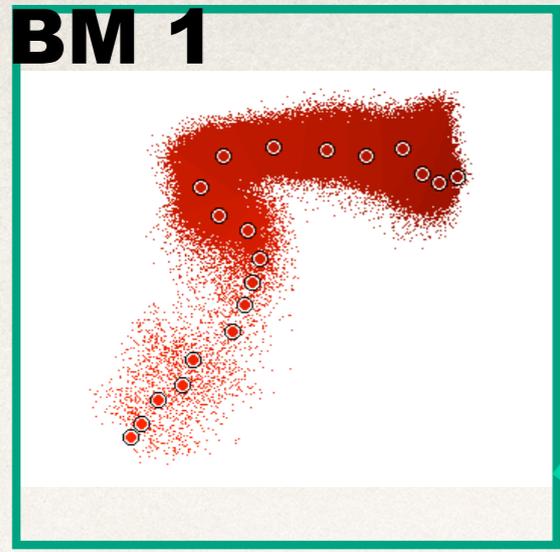
! CD7 est exprimé sur un petit contingent de blastes précoces et normaux, en particulier dans une hématopoïèse régénérative

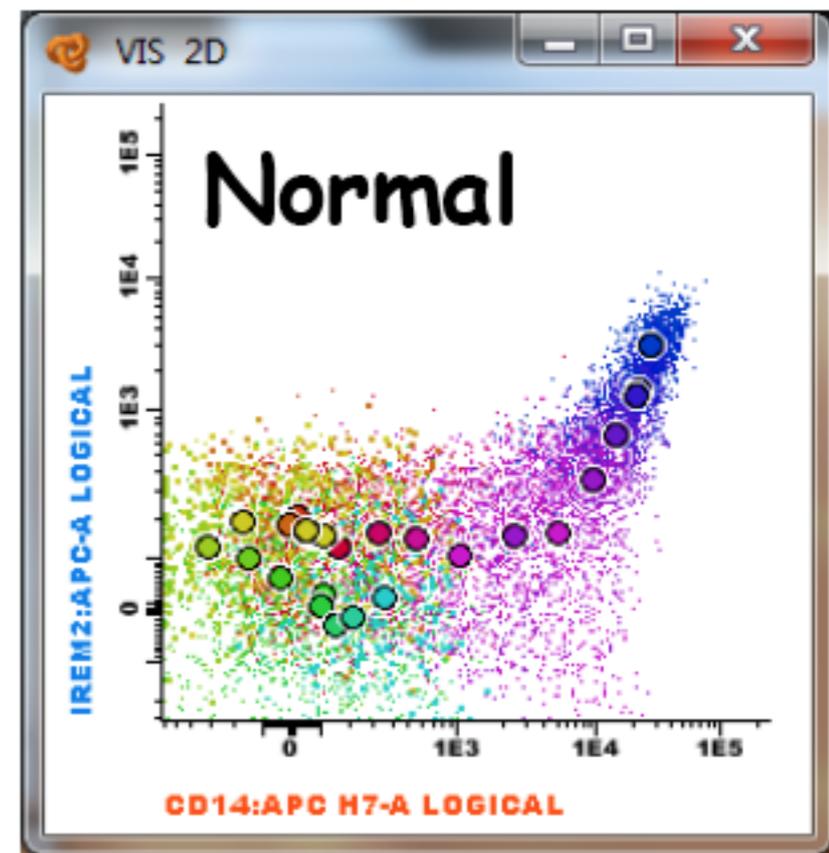
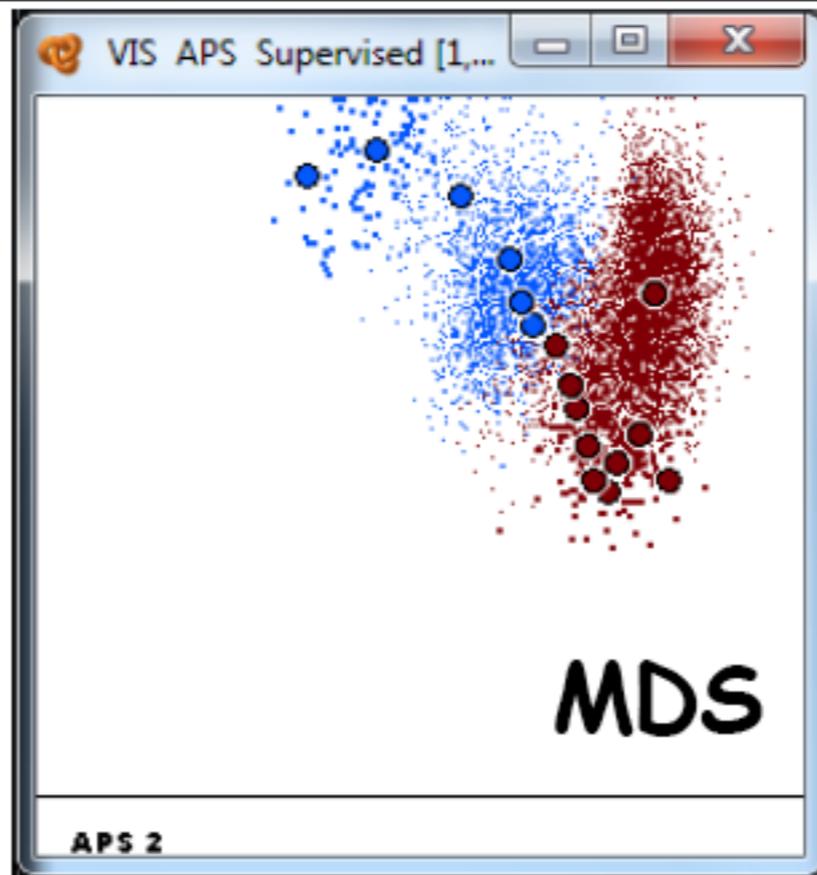
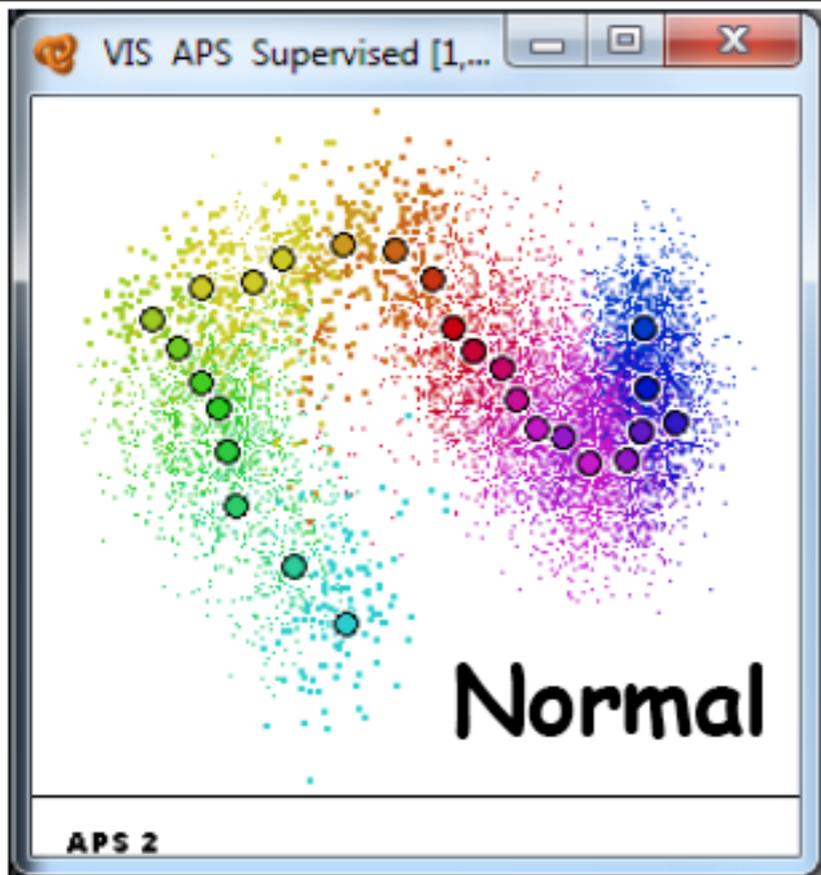




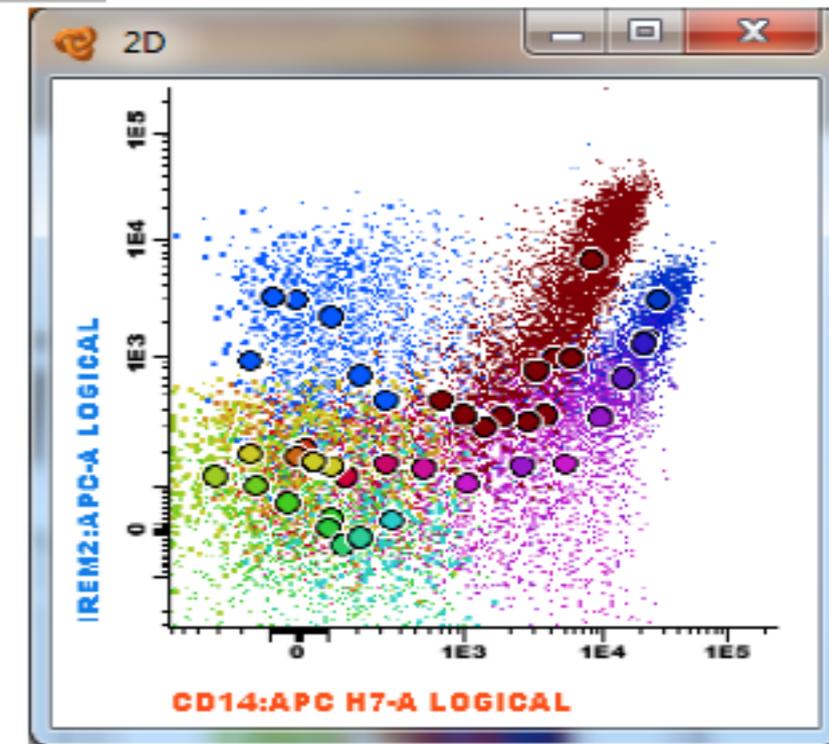
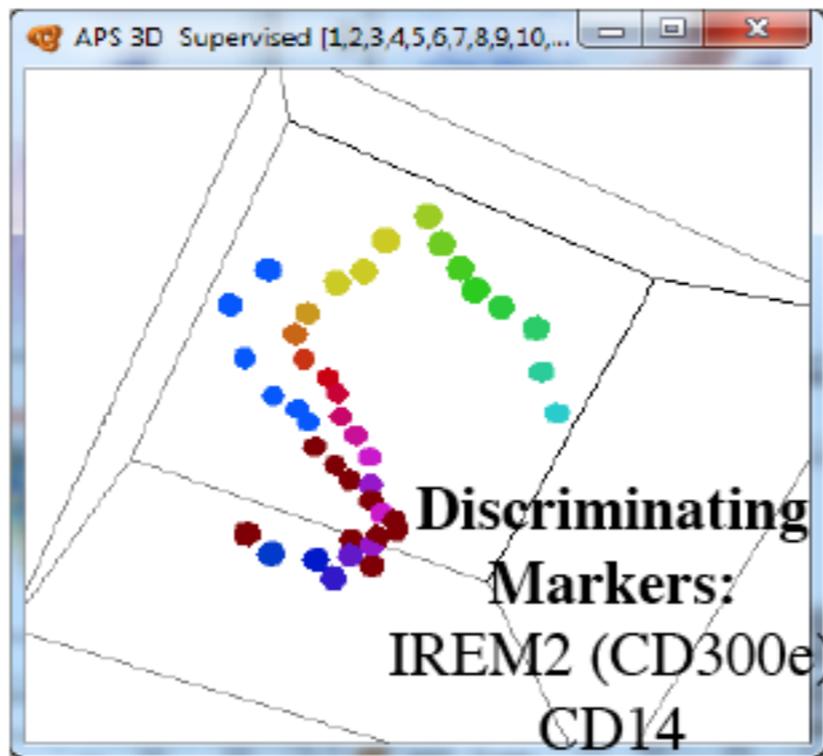
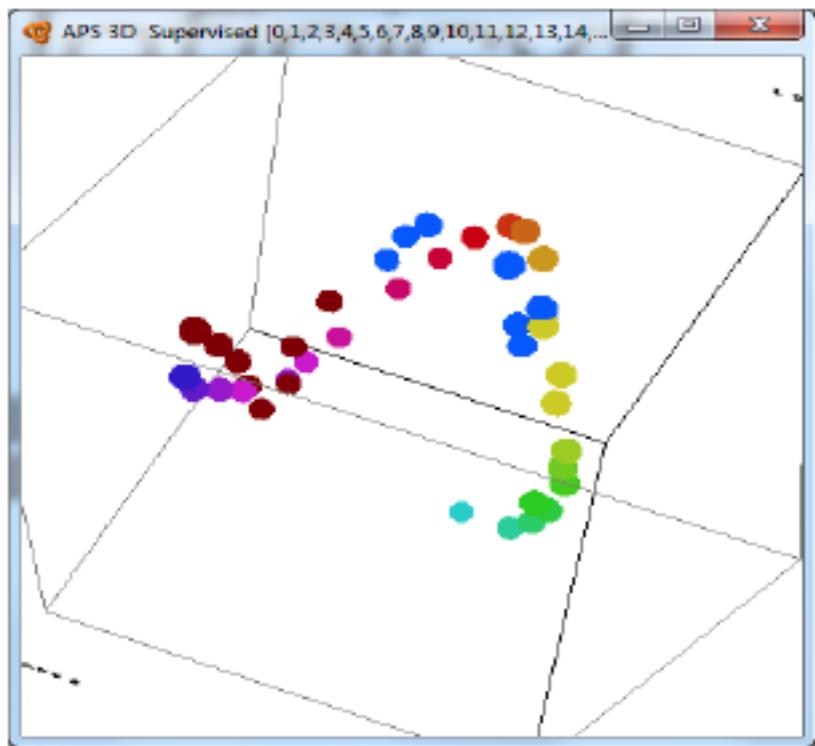
# N-DIMENSIONAL NEUTROPHIL MATURATION IN NORMAL BM





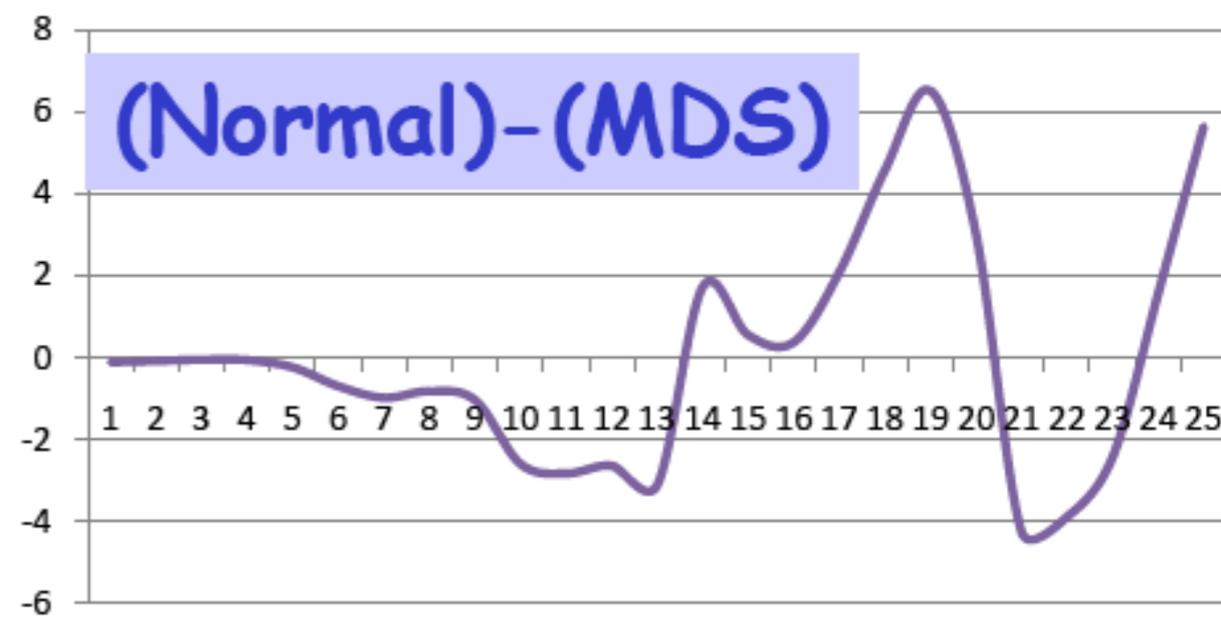
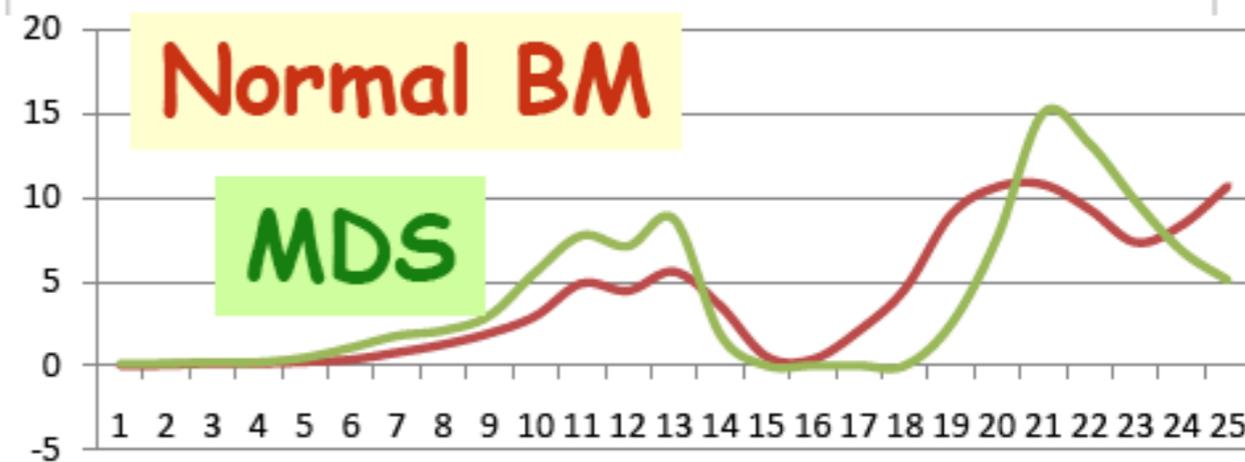
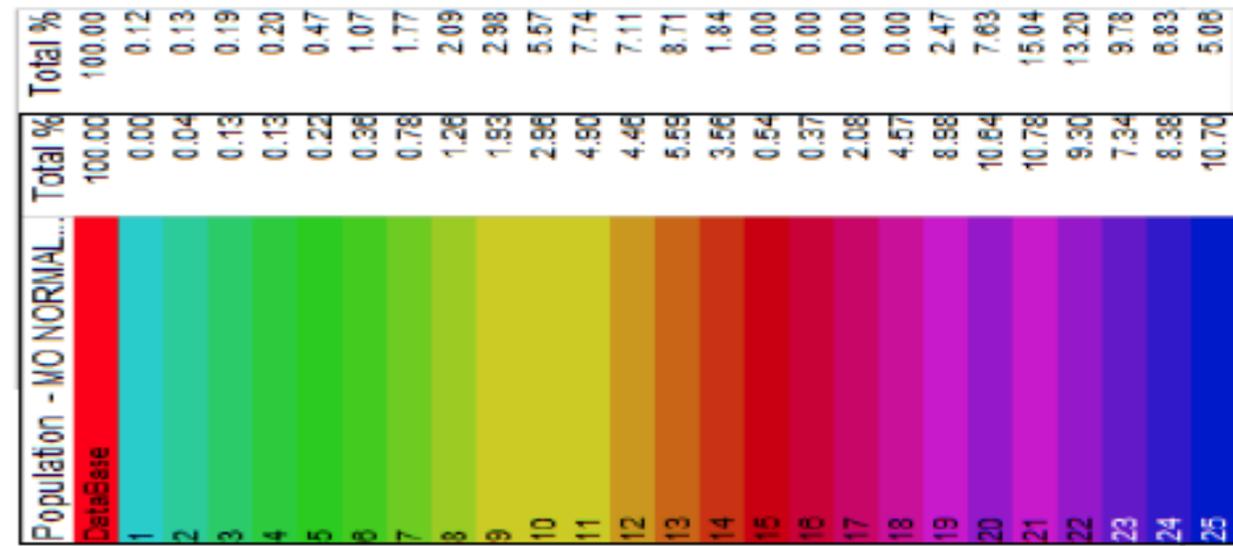
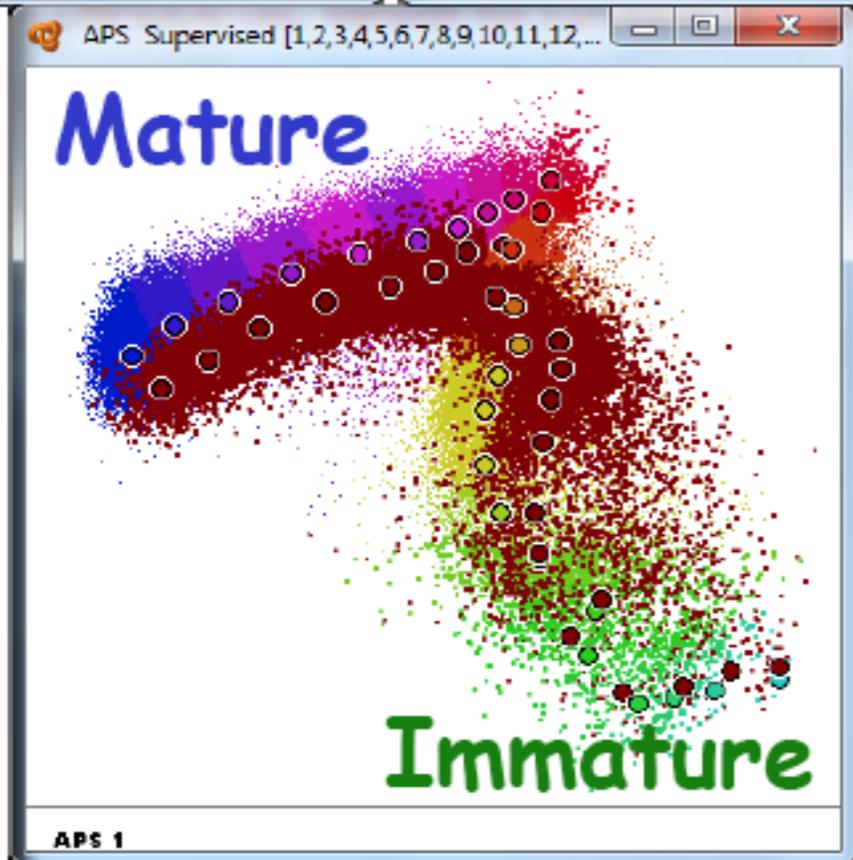
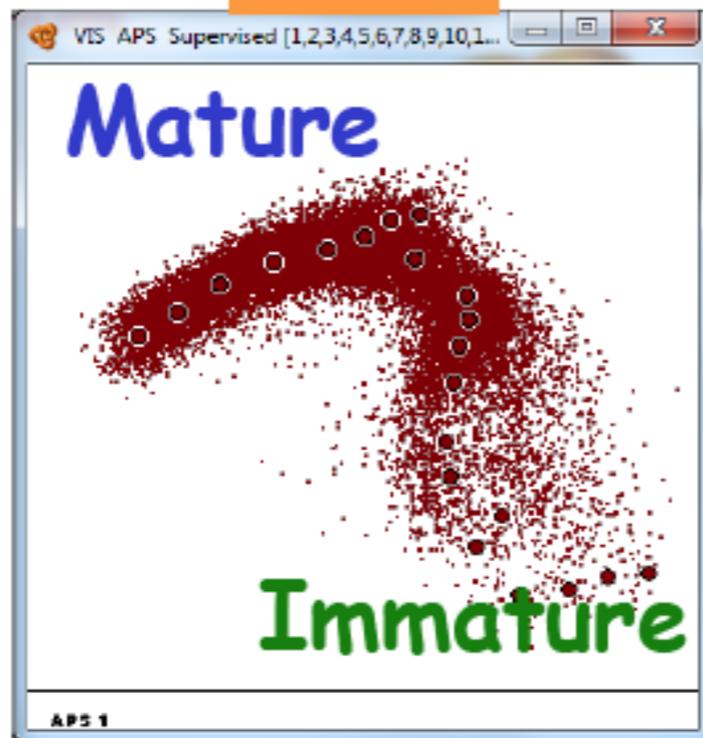
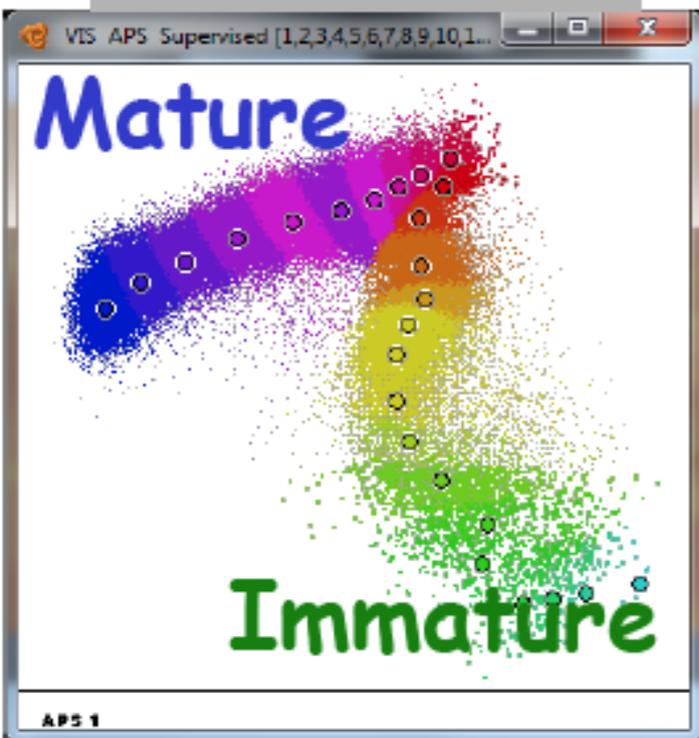


**NORMAL + MDS OVERLAY**



Normal BM

MDS



Outils informatiques performants...

# Multiplés stratégies Identification des précurseurs myéloïdes

---

- ❖  $CD45^{dim} SSC^{low/int}$
- ❖  $CD45^{dim} SSC^{low/int} CD34^{+}$
- ❖  $CD45^{dim} SSC^{low/int} HLA-DR^{+} CD11b^{-}$
- ❖  $CD45^{dim} SSC^{low/int} HLA-DR^{+} CD117^{+}$

! Expressions aberrantes  
Importance de combinaisons redondantes !

↓ SSC des neutrophiles

↑ % CD34+

Niveau d'expression anormal  
de marqueurs

# Que se passe-t-il dans un SMD ?

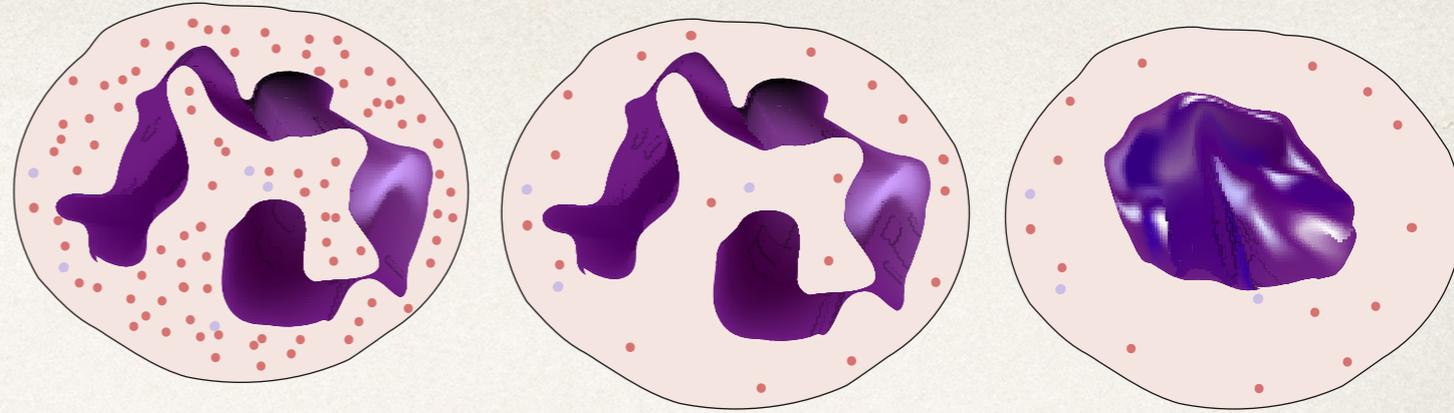
Expression d'un marqueur  
aberrant

↓ Progéniteurs B  
(parmi les CD34+)

Expression asynchrone de  
marqueurs de maturation

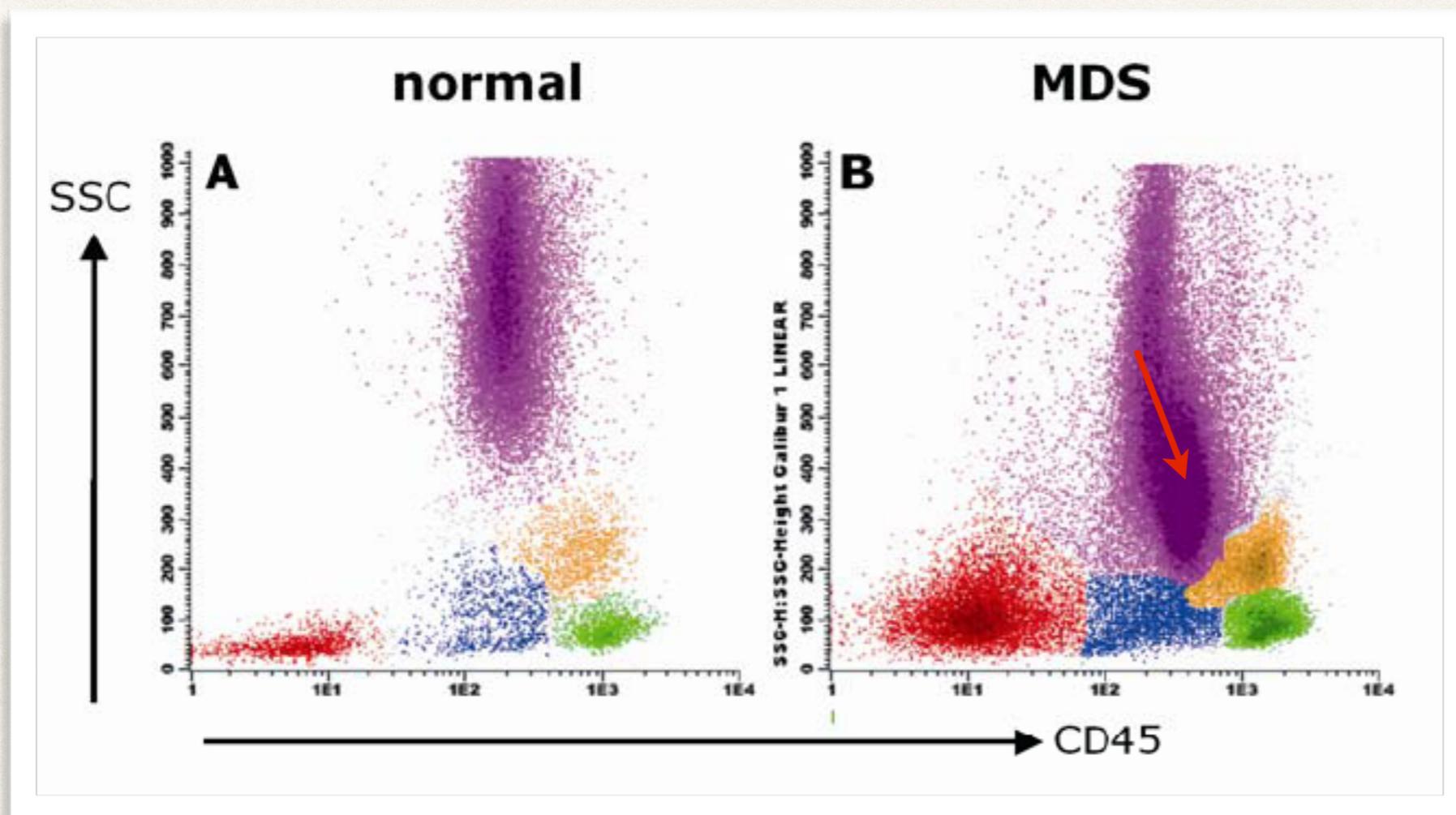


# SSC diminué



- \* SSC diminué dû à l'hypogranularité
- \* Possible manque de CD10 sur les neutrophiles matures
- \* **Le plus fréquent** : relations aberrantes entre **CD13/CD11b** et/ou **CD13/CD16**.
- \* Autres marqueurs :
  - \* Expression altérée de CD45 ou CD33 (! polymorphismes !)
  - \* Expression asynchrone de CD34
  - \* Expression de CD56, ...

# SSC abaissé (granulocytes)



*Standardization of flow cytometry in myelodysplastic syndromes : a report from an international consortium and the European leukemianet working group, TM Westers et al, Leukemia 2012*

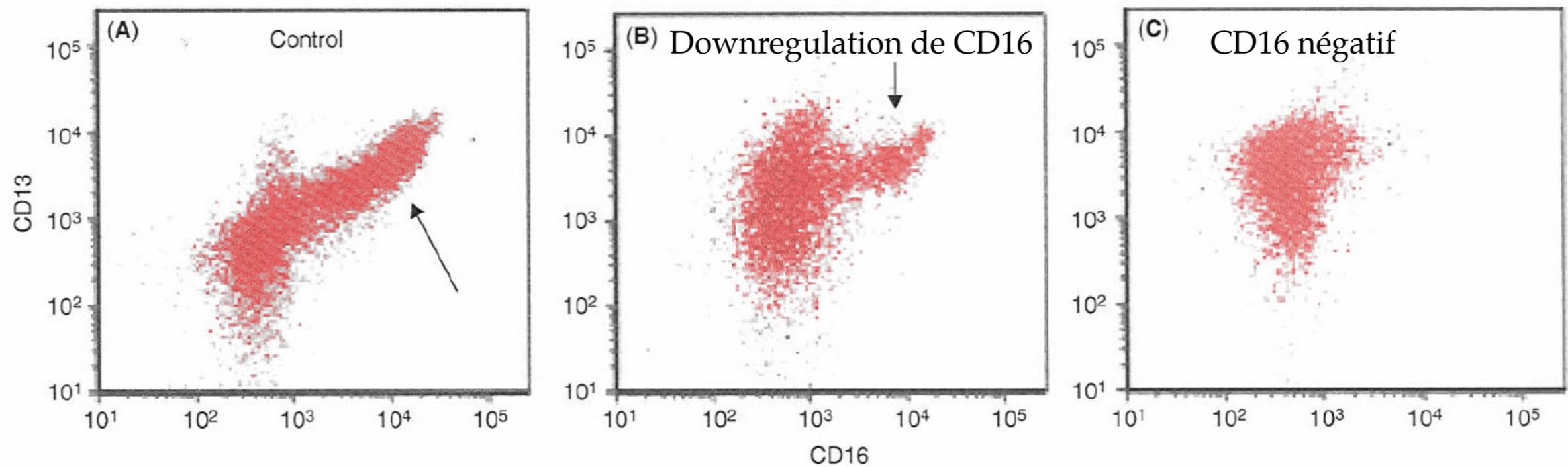


Figure 6.7 Flow cytometric feature of myelodysplastic syndrome—aberrant expression of CD16. (A) Normal (control) sample with variable expression of CD16 [mature elements are positive (arrow)]. Panel B shows prominent downregulation of CD16. Only minute subset of granulocytes is positive (arrow). Panel C shows negative CD16 in all granulocytes.

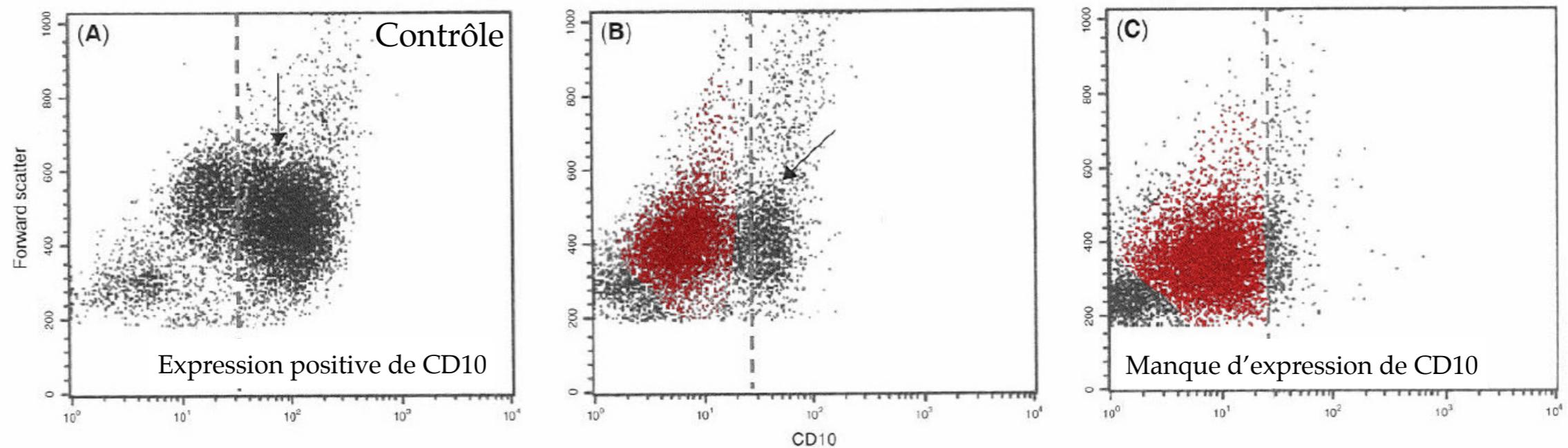


Figure 6.4 Flow cytometric feature of myelodysplastic syndrome (MDS)—aberrant expression of CD10 by granulocytes. (A) Normal (control) sample with positive CD10 expression (arrow). (B) MDS with partial loss of CD10 expression; only minute subset of granulocytes (arrow) is CD10<sup>+</sup>. (C) MDS with complete lack of CD10 expression by granulocytes.

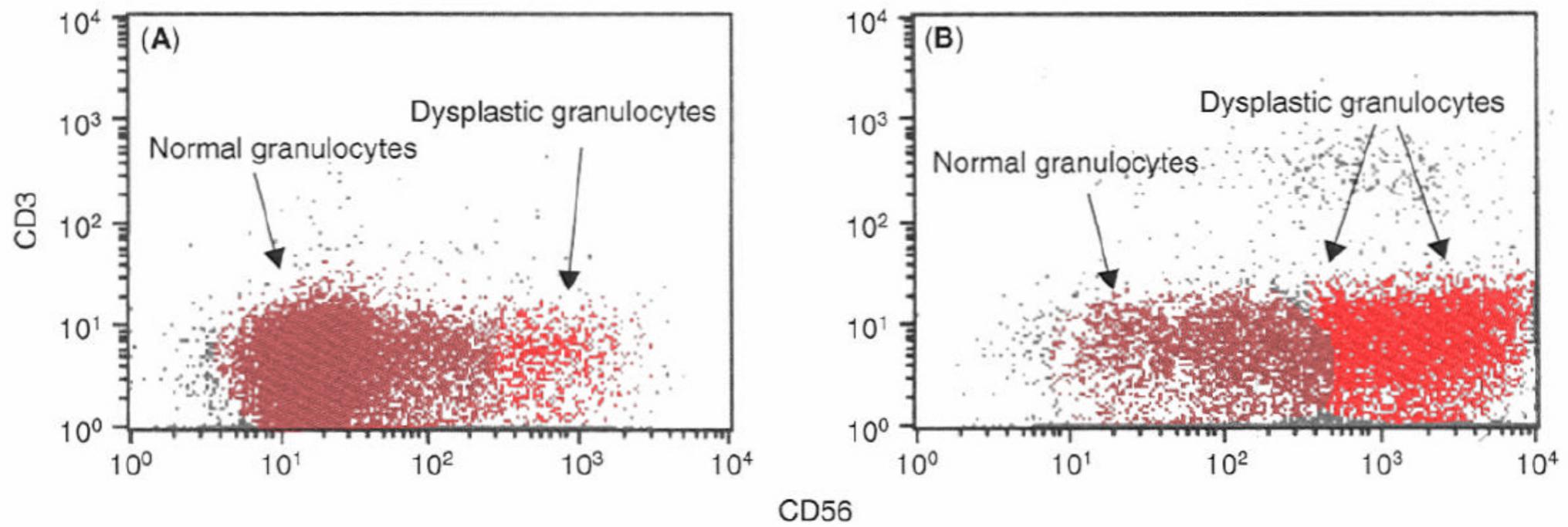
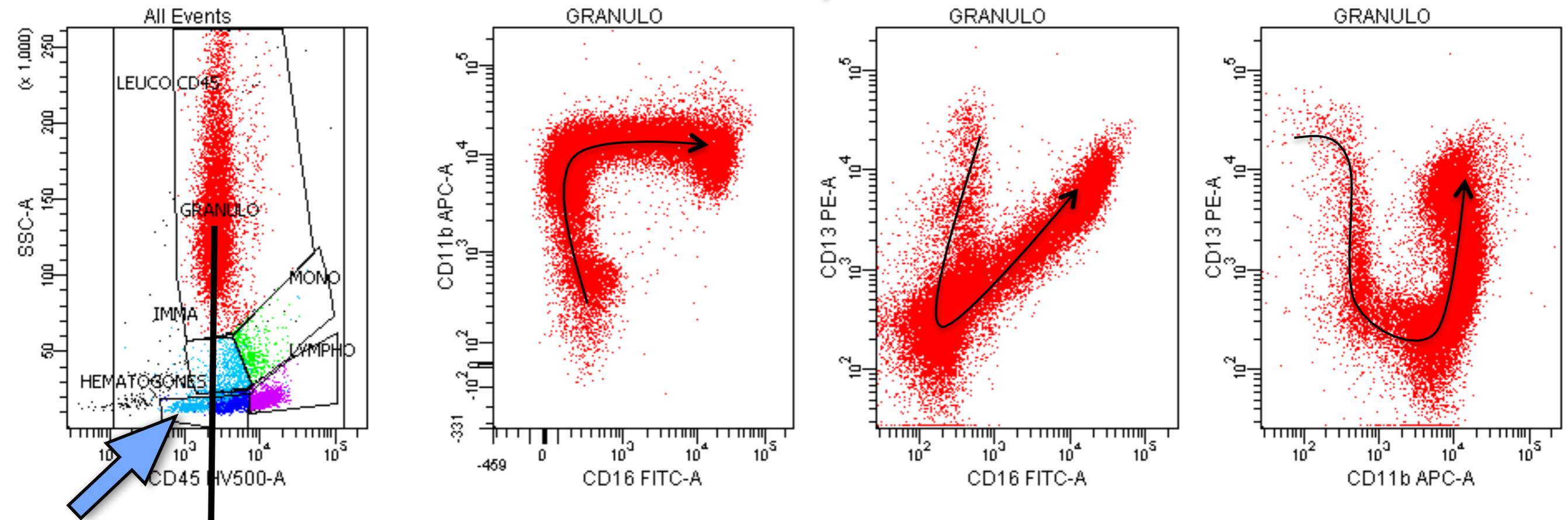
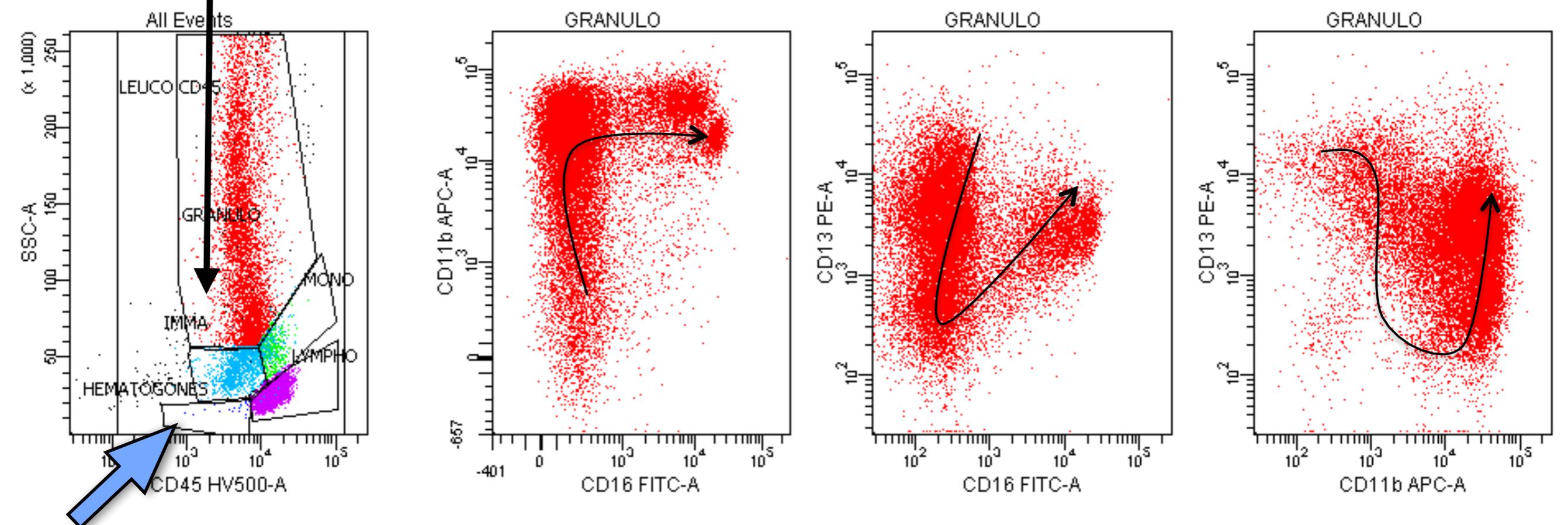


Figure 6.8 Flow cytometric feature of myelodysplastic syndrome—aberrant expression of CD56 by granulocytes (A, mild upregulation; B, marked upregulation).

# Normal pattern



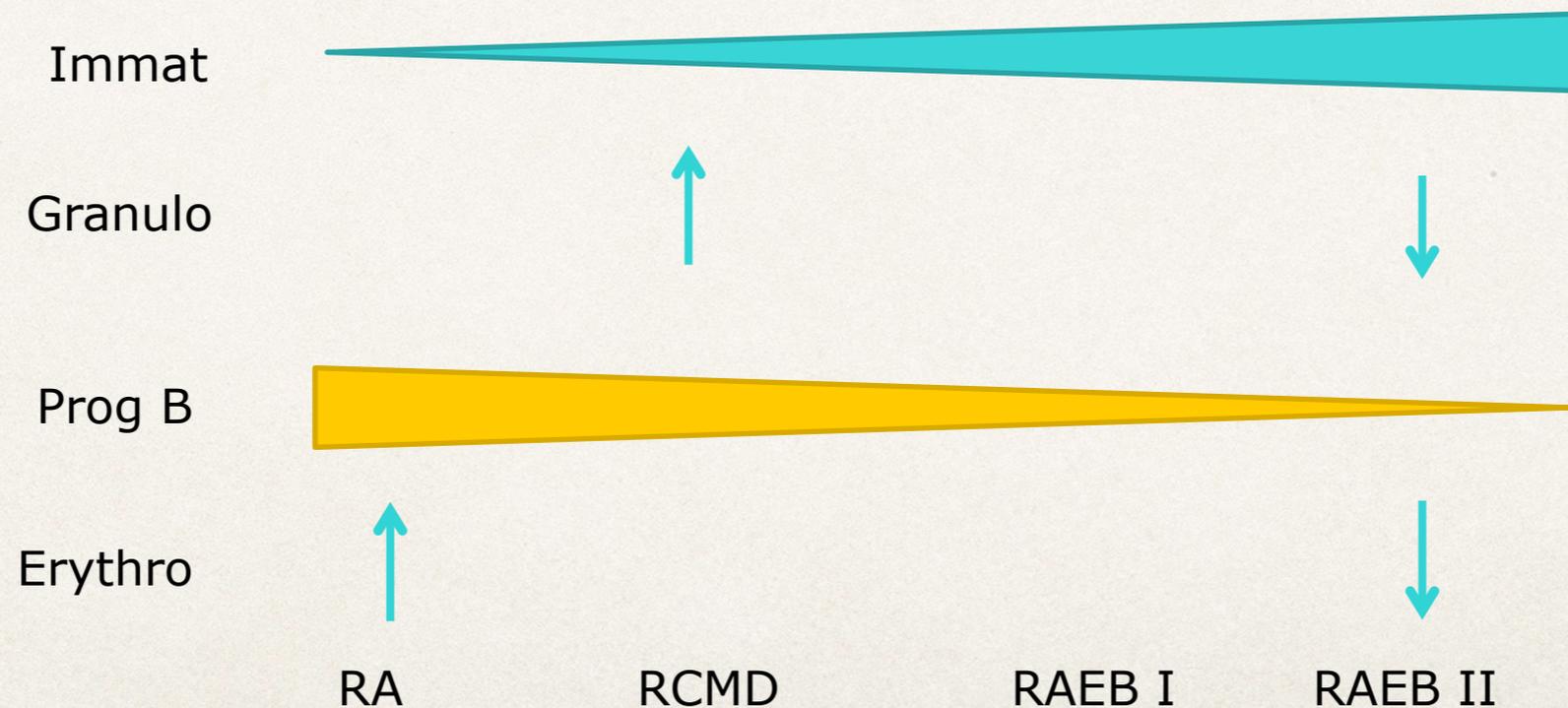
# SMD : motifs anormaux de maturation



# Diminution des progéniteurs lymphoïdes B (hematogones)

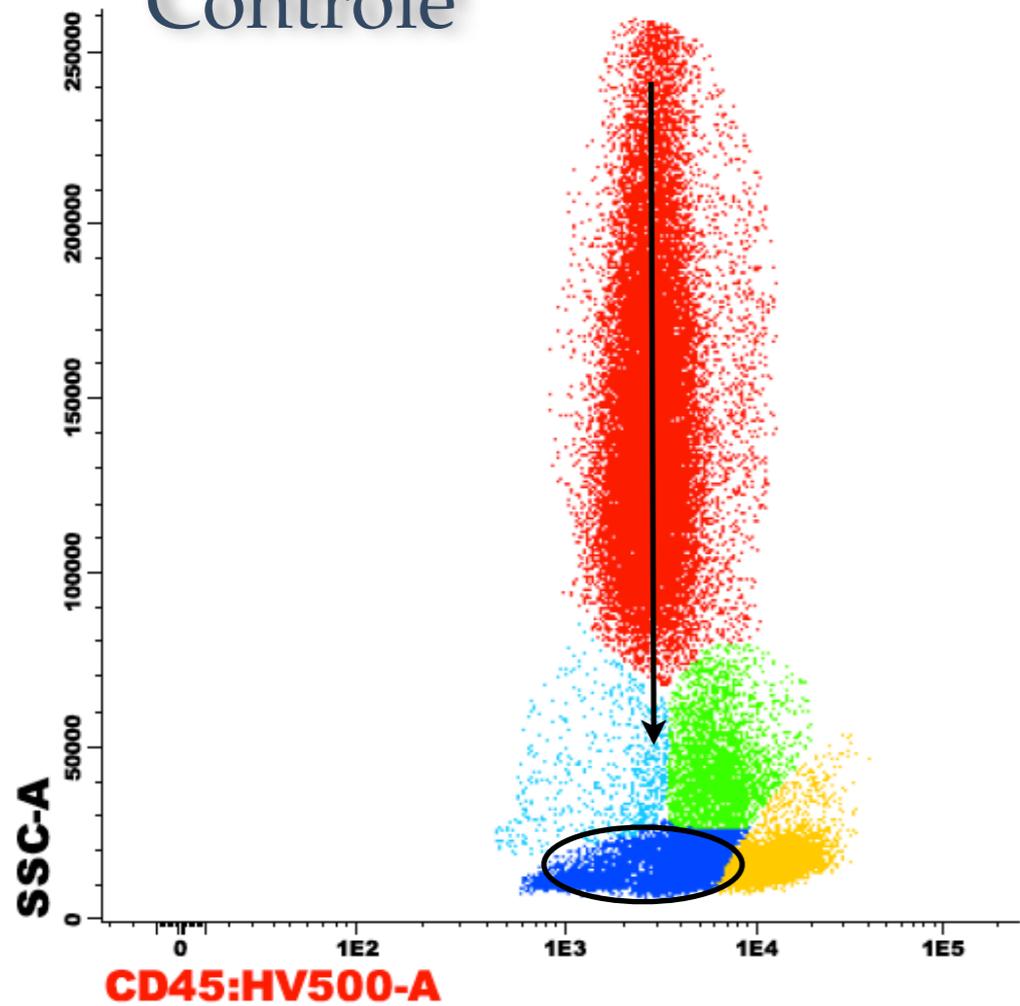
Matarraz et al, Leukemia, 2008

- \* CD34/TdT/CD10 et habituellement CD19 et CD22.
- \* Particulièrement informatif dans les SMD de bas grade
- \*  $CD45^{dim/low} SCC^{low}$  et  $CD34+/CD19+$  ou **CD34+/CD10+** (*Ogata et al*) --> **ProgB / CD34tot**
- \* Fraction des CD34+ :

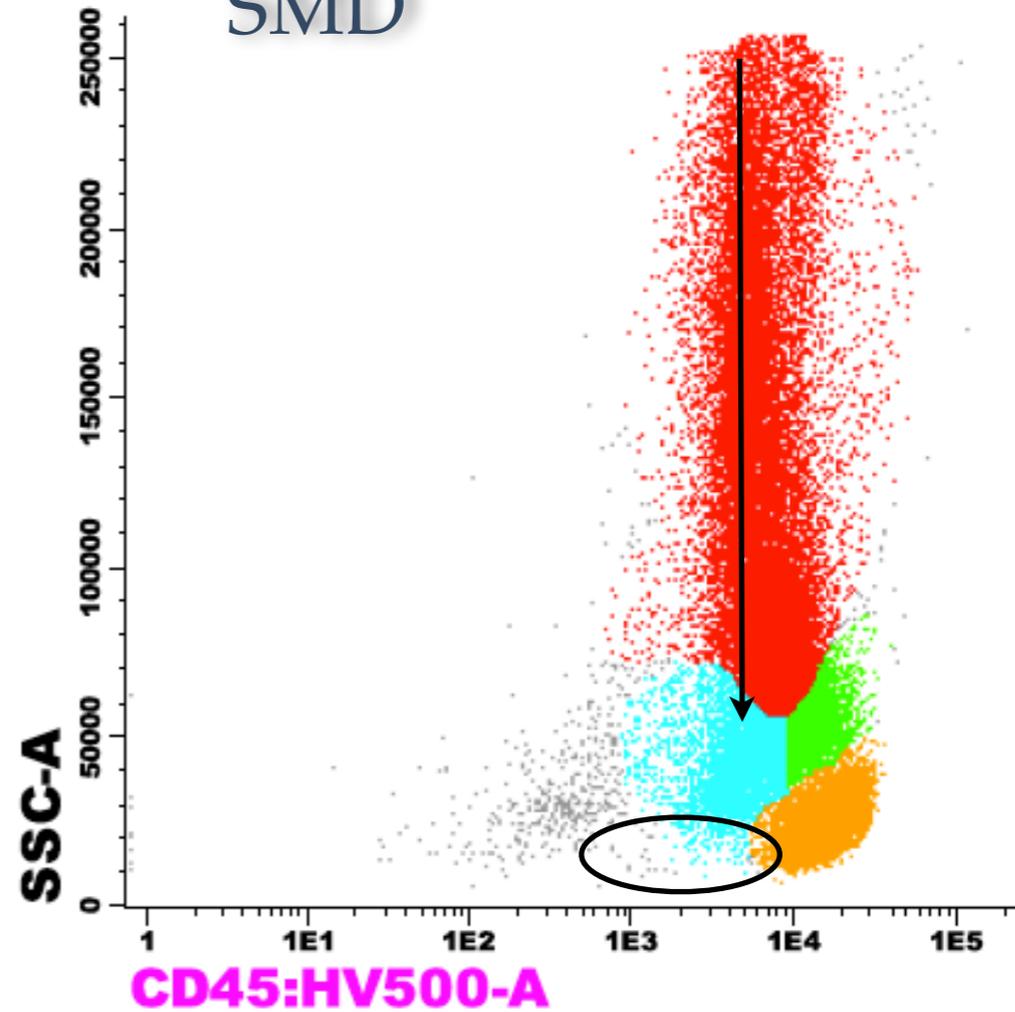


! Diminution non spécifique des SMD : age, LMC, immunodéficiences, ...

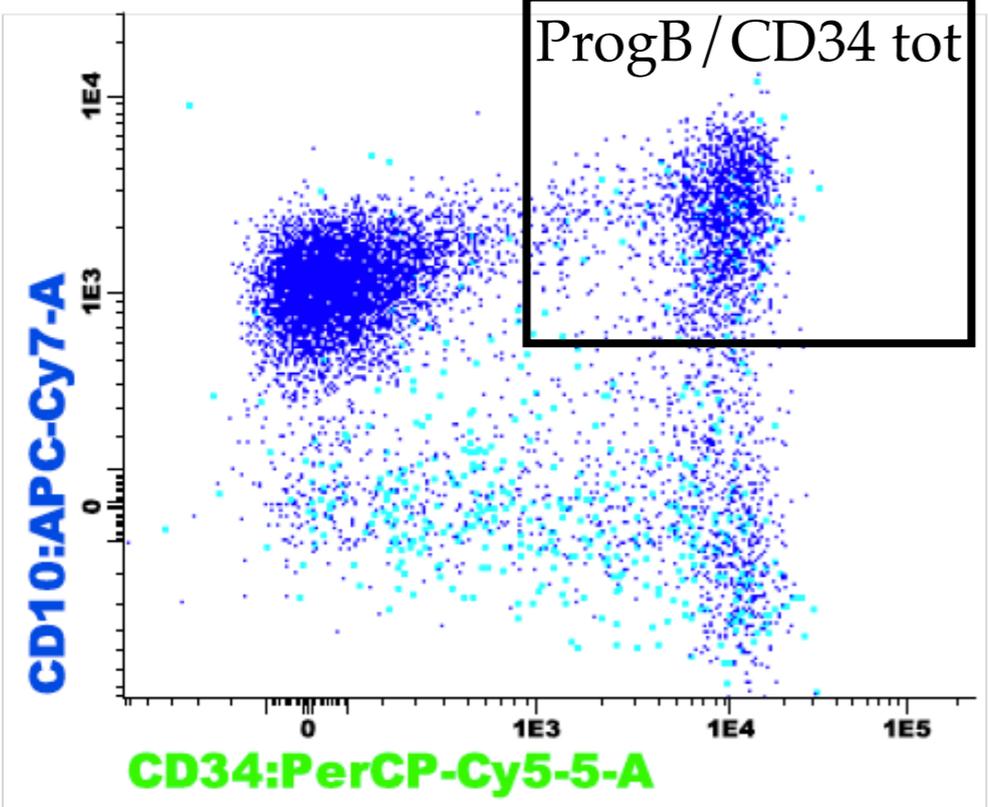
Contrôle



SMD

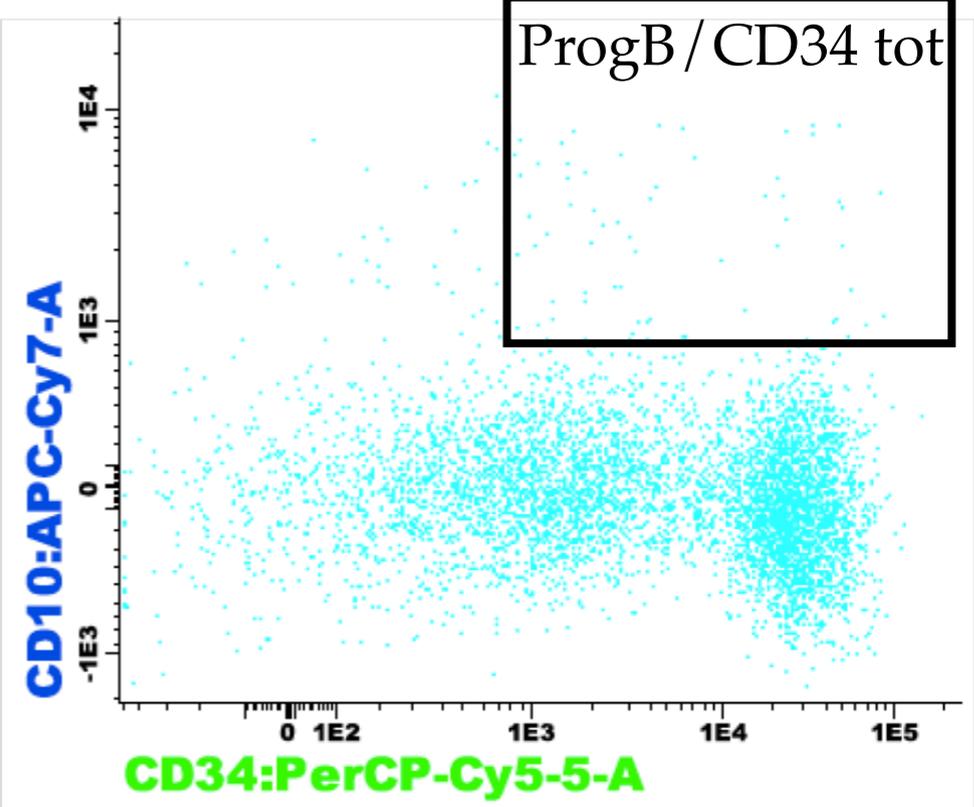


ProgB / CD34 tot



Gate sur CD34+

ProgB / CD34 tot



# Que peut-on faire au laboratoire ?

---

- \* Le diagnostic des SMD n'est pas si évident lorsque les patients manquent de marqueurs spécifiques (excès de blastes, caryotype, sidéroblastes en couronne).
- \* C'est parfois un challenge pour les SMD de bas grade qui ne possèdent pas ces spécificités.
- \* Monitorer le traitement et la progression de la pathologie.
- \* Pattern recognition analysis --> déviation de l'expression normale des antigènes... Bon outil pour les « opérateurs experts »... Mais reproductibilité ? Besoin de paramètres hautement reproductibles !
- \* Etude des caractéristiques cytométriques —> scores (pour discriminer les SMD de bas grade des cytopénies non-clonales).

# FCSS (*Flow Cytometry Scoring System*)

2012

Standardisation



**Standardization of flow cytometry in myelodysplastic syndromes: a report from an international consortium and the European leukemianet working group**

TM Westers, R Ireland, W Kern, C Alhan, JS Balleisen, P Bettelheim, K Burbury, M Cullen, JA Cutler, MG Della Porta, AM Dräger, J Feuillard, P Font, U Germing, D Haase, U Johansson, S Kordasti, MR Loken, L Malcovati, JG te Marvelde, S Matarraz, T Milne, B Moshaver, G J Mufti, K Ogata, A Orfao, A Porwit, K Psarra, SJ Richards, D Subirá, V Tindell, T Vallespi, P Valent, VHJ van der Velden, TM de Witte, DA Wells, F Zettl, MC Béné and AA van de Loosdrecht

Score

**Multicentre validation of a reproducible flow cytometric score for the diagnosis of low-risk myelodysplastic syndromes: results of a European LeukemiaNET study**

by Matteo G. Della Porta, Cristina Picone, Cristiana Pascutto, Luca Malcovati, Hideto Tamura, Hiroshi Handa, Magdalena Czader, Sylvie Freeman, Paresh Vyas, Anna Porwit, Leonie Saft, Theresia M. Westers, Canan Alhan, Caludia Cali, Arjan A. Van de Loosdrecht, and Kiyoyuki Ogata

*Haematologica* 2012 [Epub ahead of print] *étude multi-centrique*

# Standardisation ?

---

- \* CMF est de plus en plus reconnue pour jouer un rôle important dans diagnostic et le pronostic des SMD.
- \* European LeukemiaNet (ELN) MDS workshops.
- \* **4 paramètres hautement reproductibles :**
  - \* Pourcentage augmenté des progéniteurs CD34+ (cellules nucléées)
  - \* Nombre diminué de progéniteurs B parmi les CD34+
  - \* Diminution ou augmentation de l'expression du CD45 sur les cellules myéloïdes progénitrices
  - \* Diminution du SSC des neutrophiles

# Scoring system

**Multicentre validation of a reproducible flow cytometric score for the diagnosis of low-risk myelodysplastic syndromes: results of a European LeukemiaNET study**

by Matteo G. Della Porta, Cristina Picone, Cristiana Pascutto, Luca Malcovati, Hideto Tamura, Hiroshi Handa, Magdalena Czader, Sylvie Freeman, Paresh Vyas, Anna Porwit, Leonie Saft, Theresia M. Westers, Canan Alhan, Caludia Cali, Arjan A. Van de Loosdrecht, and Kiyoyuki Ogata

*Haematologica* 2012 [Epub ahead of print]

797 patients

- Devrait être reproductible entre les différents opérateurs
- Les résultats doivent être facilement compris par les cliniciens

Score SMD	Valeurs cut-off (normalité)
Myeloblast-related cluster size (%) <sup>*</sup>	< 2 %
B-progenitor-related cluster size (%) <sup>**</sup>	> 5 %
Lymphocytes to myeloblasts CD45 ratio	4 - 7,5
Granulocytes to lymphocytes SSC ratio	> 6

\* in all nucleated cells

\*\* in all CD34+

# Diagnostic des SMD bas grade

Multicentre validation of a reproducible flow cytometric score for the diagnosis of low-risk myelodysplastic syndromes: results of a European LeukemiaNET study

by Matteo G. Della Porta, Cristina Picone, Cristiana Pascutto, Luca Malcovati, Hideto Tamura, Hiroshi Handa, Magdalena Czader, Sylvie Freeman, Paresh Vyas, Anna Porwit, Leonie Saft, Theresia M. Westers, Canan Alhan, Caludia Cali, Arjan A. Van de Loosdrecht, and Kiyoyuki Ogata

Haematologica 2012 [Epub ahead of print]

Positifs  
n=281

Learning cohort	FCM-score					positive cases	sensitivity (%)	specificity (%)
	0	1	2	3	4			
Low-risk MDS	33	50	107	67	24	198/281	70%	
Specific markers of dysplasia*								
MDS with specific markers	17	26	58	49	16	123/166	74%	
MDS without specific markers	16	24	49	18	8	75/115	65%	
WHO category								
RCUD	8	12	26	6	3	35/55	64%	
RARS	7	5	13	5	0	18/30	60%	
RCMD/RS	15	31	61	51	20	132/178	74%	
MDS with del5q	1	1	6	5	1	12/14	86%	
MDS-U	2	1	1	0	0	1/4	-	
Controls	132	107	16	2	0	18/257		93%
Idiopathic or iatrogenic hypoplasia**	6	7	7	0	0	7/20		35%
Anemia***	42	57	5	0	0	5/104		95%
Cytopenia associated with BM infiltration	6	5	1	2	0	2/14		86%
Cytopenia in transplant recipients	0	3	0	0	0	0/3		-
Infective cytopenia	2	4	0	0	0	0/6		-
Immune cytopenia****	76	31	3	0	0	3/110		97%

Negatifs  
n=257





Arlon



les  
Cliniques  
du Sud  
Luxembourg

Alfaber

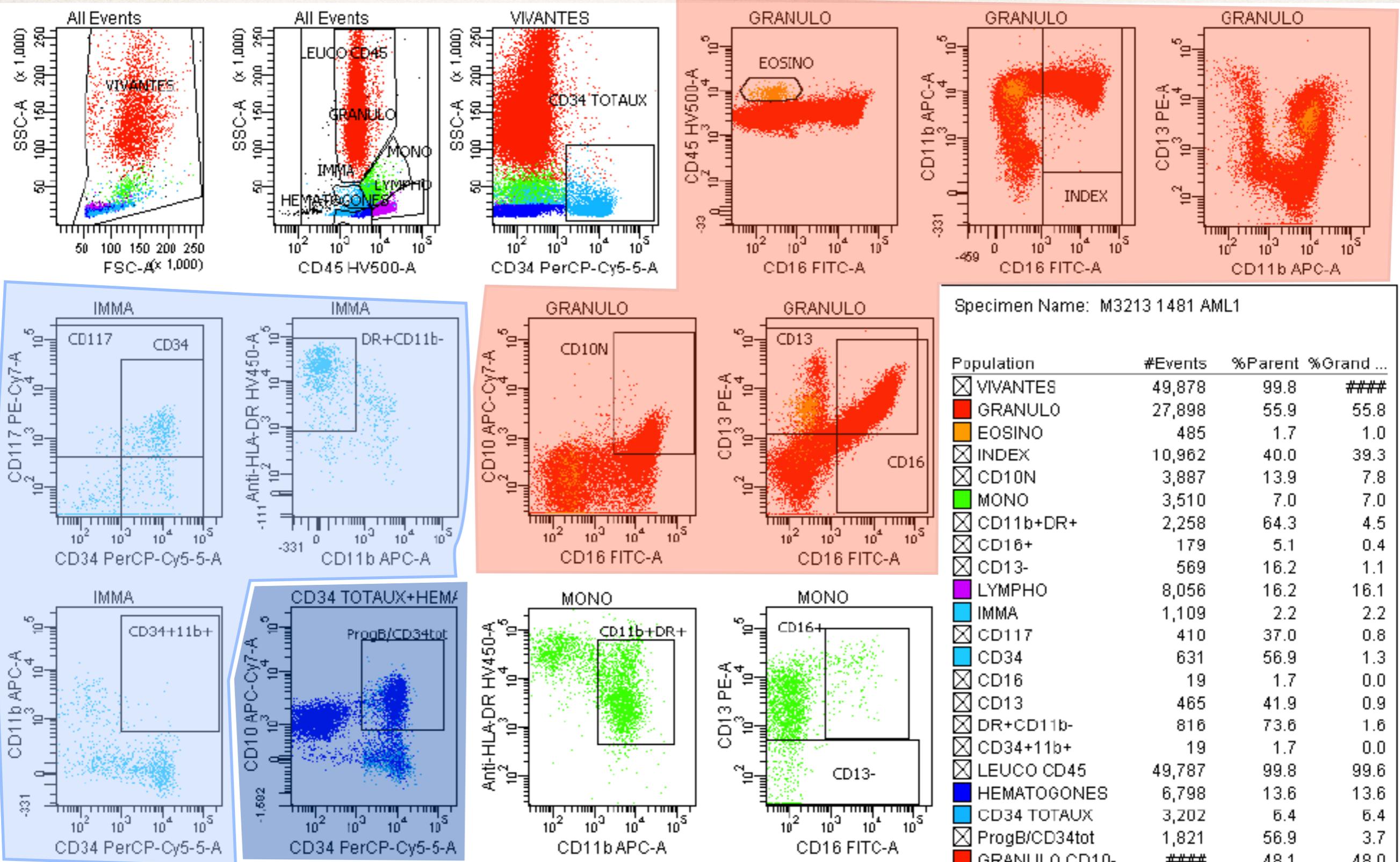
# Notre expérience



- \* **433 patients** --> Analyse médullaire (Aspiration, Biopsie, FC)
- \* 8 couleurs... *AML-1 tube EuroFlow* :
  - \* **HLA-DR / CD45 / CD16 / CD13 / CD34 / CD117 / CD11b / CD10**
- \* Scoring System :

Score SMD	Valeurs cut-off (normal)
% Myeloblasts	< 2 %
% ProgB/CD34 <sub>tot</sub>	> 5 %
Mean CD45 FMI Ly/FMI CD34	4 - 7,5
SSC Mode CD10-/LYMPHO	> 6

# Notre pattern - tube AML1



Specimen Name: M3213 1481 AML1

Population	#Events	%Parent	%Grand ...
VIVANTES	49,878	99.8	###
GRANULO	27,898	55.9	55.8
EOSINO	485	1.7	1.0
INDEX	10,962	40.0	39.3
CD10N	3,887	13.9	7.8
MONO	3,510	7.0	7.0
CD11b+DR+	2,258	64.3	4.5
CD16+	179	5.1	0.4
CD13-	569	16.2	1.1
LYMPHO	8,056	16.2	16.1
IMMA	1,109	2.2	2.2
CD117	410	37.0	0.8
CD34	631	56.9	1.3
CD16	19	1.7	0.0
CD13	465	41.9	0.9
DR+CD11b-	816	73.6	1.6
CD34+11b+	19	1.7	0.0
LEUCO CD45	49,787	99.8	99.6
HEMATOGONES	6,798	13.6	13.6
CD34 TOTAUX	3,202	6.4	6.4
ProgB/CD34tot	1,821	56.9	3.7
GRANULO CD10-	###	48.1	48.0

Based on Euroflow recommendations

# Résultats patients (exclu : LMA de novo, ...)

	Score = 4	Score = 3	Score = 2	Score = 1	Score = 0
n = 433	10	32	44	105	242

- ❖ Score = 4 :

- ❖ AREB-I, AREB-II, AREB/LMA

- ❖ Score = 3 :

- ❖ AREB, RCMD, LMMC, MDS del(7), AREB/LMA, ...

- ❖ Score = 2 :

- ❖ (AREB), RA, 5q-, **aplasie toxique, moelle régénérative (en particulier post-LLA), déficience B12, SMP (peu hématog)**

- ❖ Score = 1 or 0 :

- ❖ Moelle normale ou régénérative, MGUS, Myelome, Vaquez, Hypoplasie, LLC, LNH, LMC, **ARSI, MK, ...**

# Résultats patients

	Score = 4	Score = 3	Score = 2	Score = 1	Score = 0
n = 433	10	32	44	105	242
SMD	10	32	28	9	6
non-SMD	0	0	16	96	236

Sensibilité : 77%

Spécificité : 97%

VPP : 87%

VPN : 94 %

- \* Score of 4 ou 3 : spécificité de 100% ! chez nos patients.
- \* Score of 1 ou 0 : argument contre une SMD, mais pas exclu...
- \* Score of 2 : attention ! Suivre le patient.

# Un exemple...

- \* Homme de 55 ans, pancytopenie.

- Hb : 9,1 g/dL  
- GR : 2,47 millions/ $\mu$ L  
- *MCV* : 105,7 fl  
- *Retic* : 20.300/ $\mu$ L (0,8%)  
- GB : 3.640/ $\mu$ L  
- *Neutro* : 19,3% (700/ $\mu$ L)  
- *Blastes* : 0,8%

- \* Moelle : dysplasie multi-lignée, excès de blastes --> **AREB II**

- \* Cytogénétique : mauvais pronostic

- \* clone 1 : 44XY, ADD(2)(P21), **-7**, DEL(12)(P11)

- \* clone 2 : 44, SL, DER(5) T(2;5) (P12;Q24)

Cytométrie en flux ?

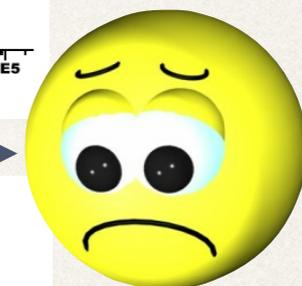
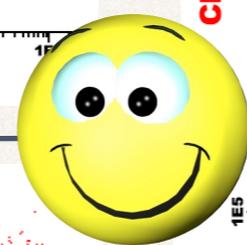
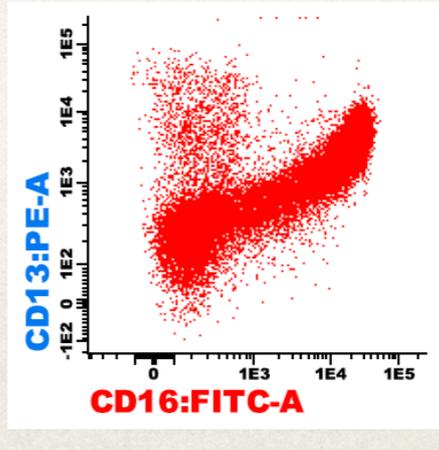
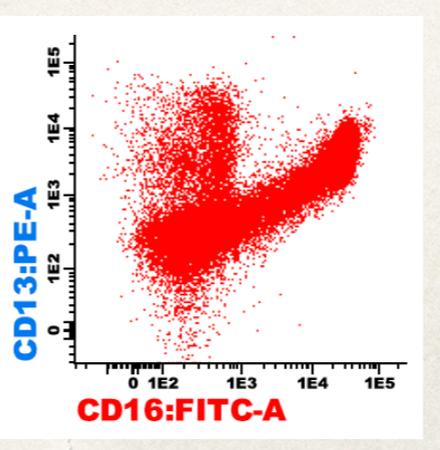
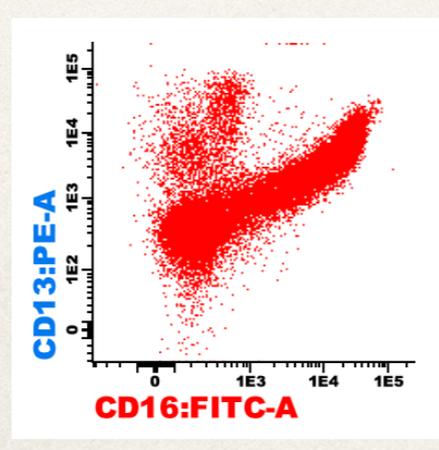
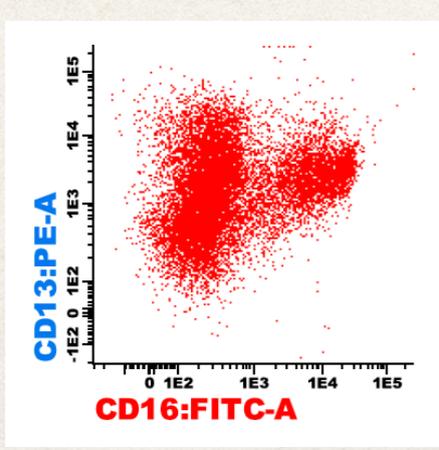
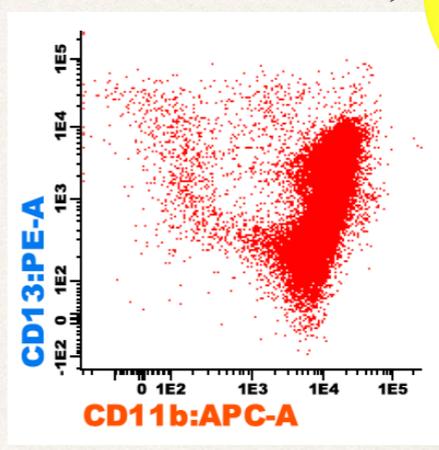
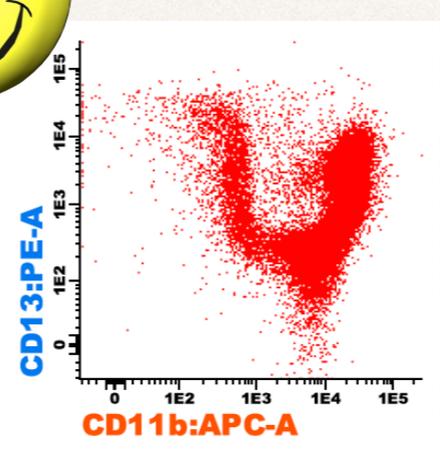
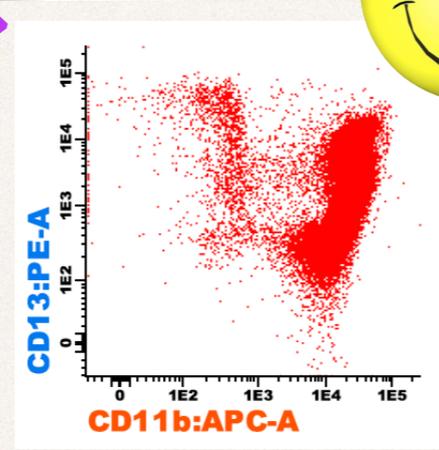
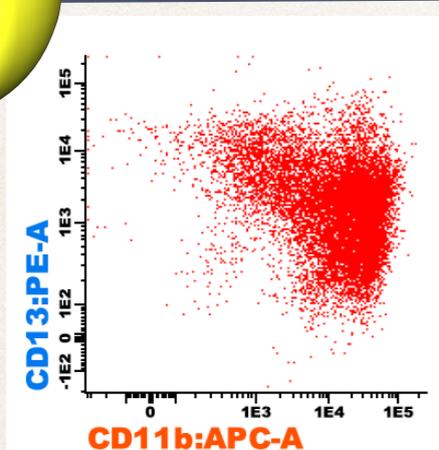
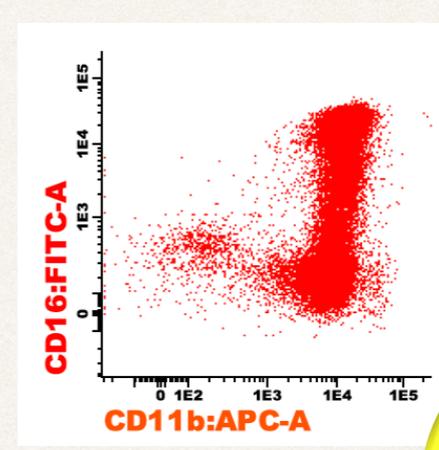
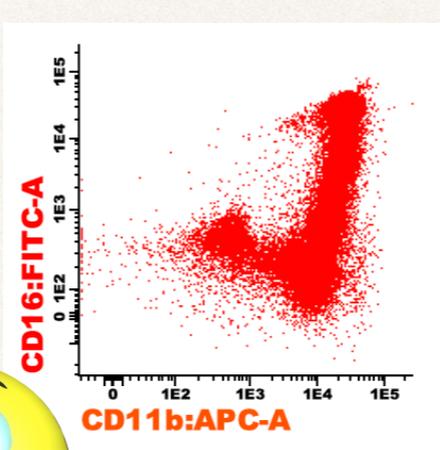
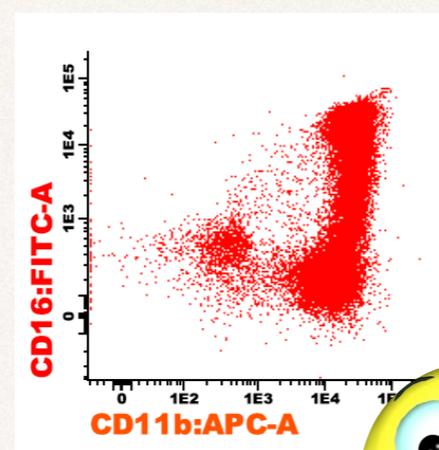
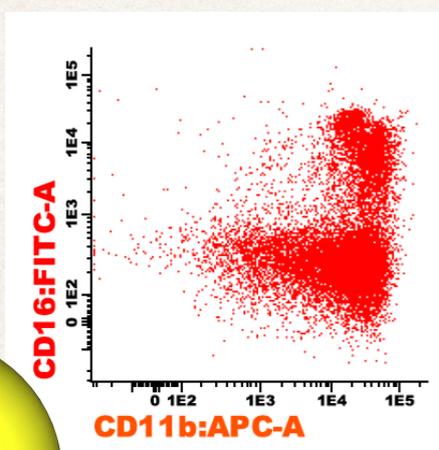
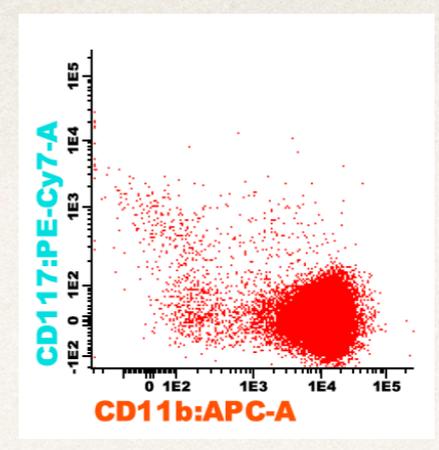
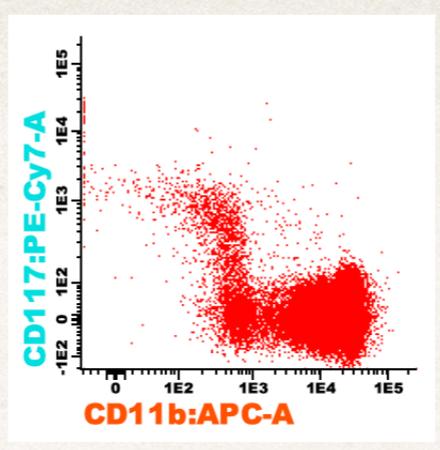
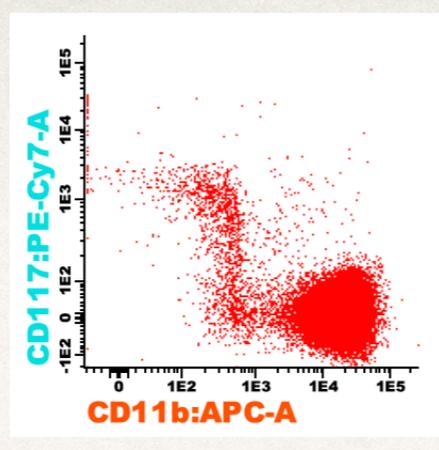
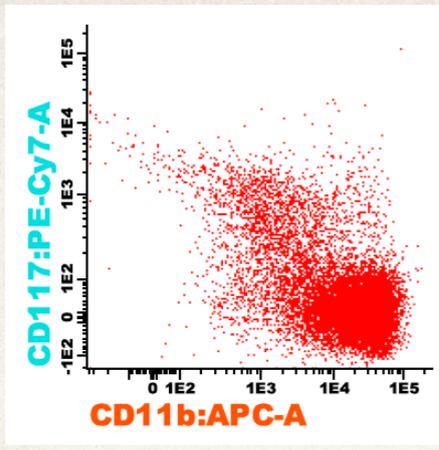
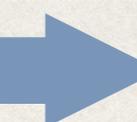


11/2012

02/2012

03/2012

04/2012



Traitement

	11/2011	02/2012	03/2012	04/2012
<b>FCSS</b>	<b>Score = 4</b>	<b>Score = 0</b>	<b>Score = 0</b>	<b>Score = 2</b>
% Myeloblasts ( <b>&lt; 2%</b> )	<b>8</b>	<b>1,1</b>	<b>1,9</b>	<b>3,7</b>
% ProgB/ CD34 <sub>tot</sub> ( <b>&gt; 5%</b> )	<b>3,5</b>	<b>10,5</b>	<b>7,5</b>	<b>5,3</b>
Mean CD45 FMI Ly/FMI CD34 ( <b>4-7,5</b> )	<b>2,9</b>	<b>5,6</b>	<b>5</b>	<b>5,5</b>
SSC Mode CD10-/ LYMPHO ( <b>&gt; 6</b> )	<b>4</b>	<b>7</b>	<b>8</b>	<b>6</b>



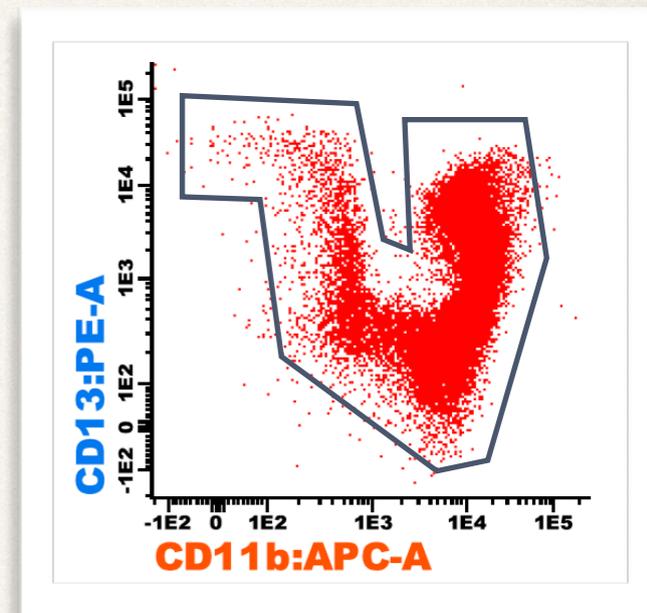
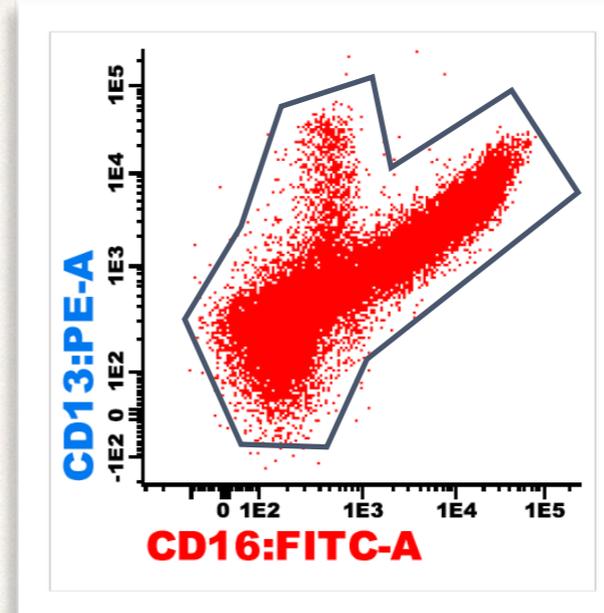
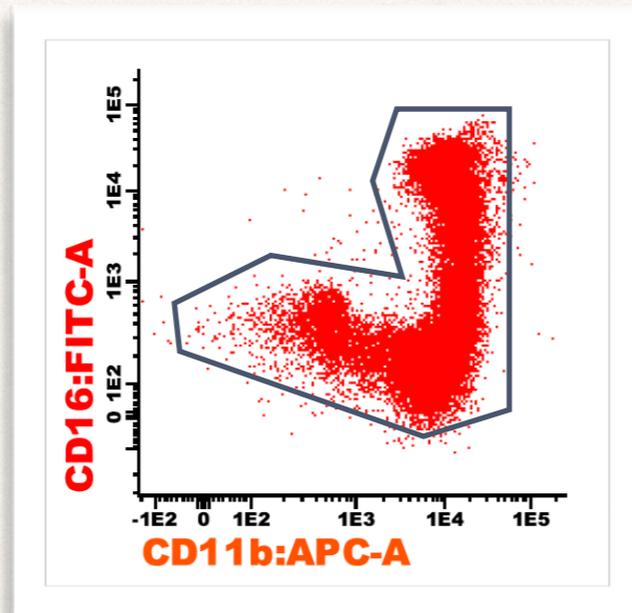
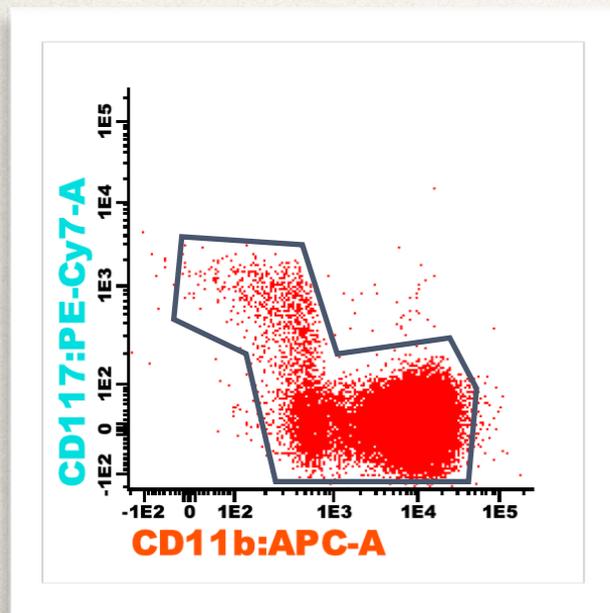
# Que peut-on observer ?

---

- ❖ ! Il s'agit d'un patient !
- ❖ Score = 4 au diagnostic --> traitement --> score = 0
- ❖ 5 mois après le diagnostic: rechute --> score = 2
- ❖ Le FCSS semble suivre l'évolution des patients ?

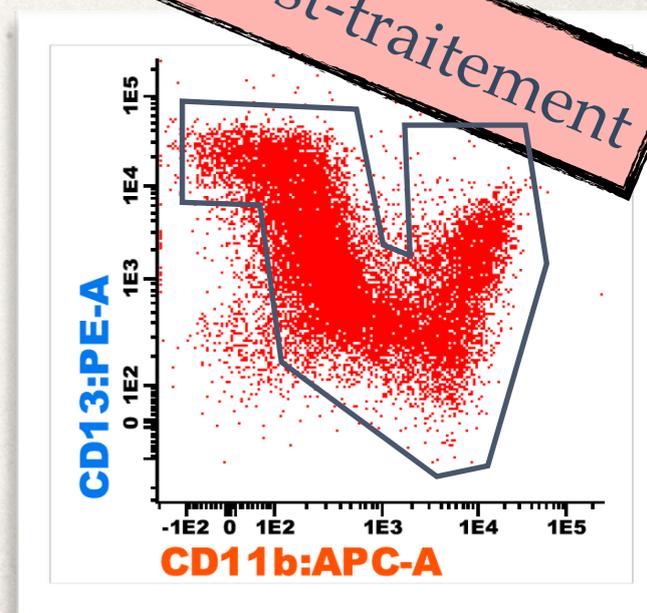
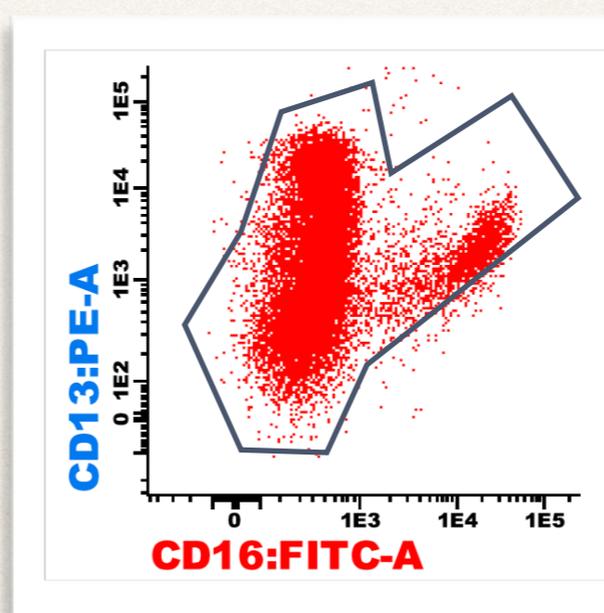
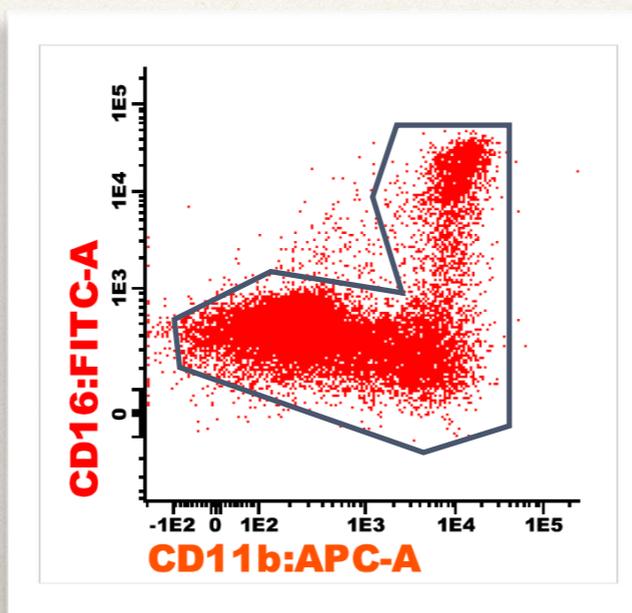
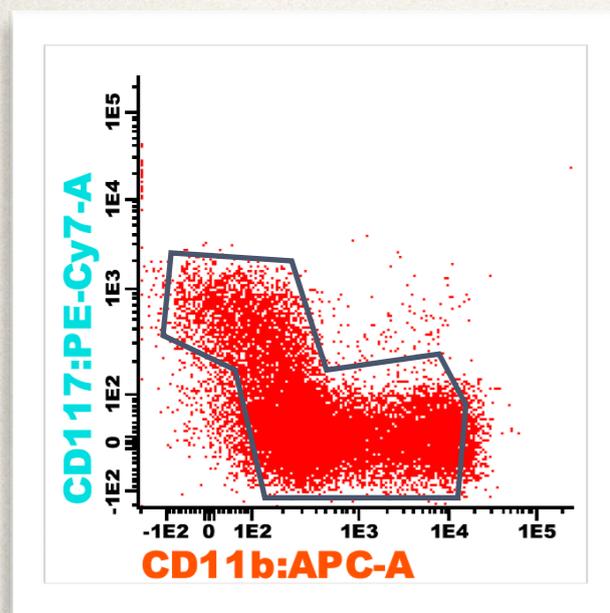
*Et maintenant ?*

Normal

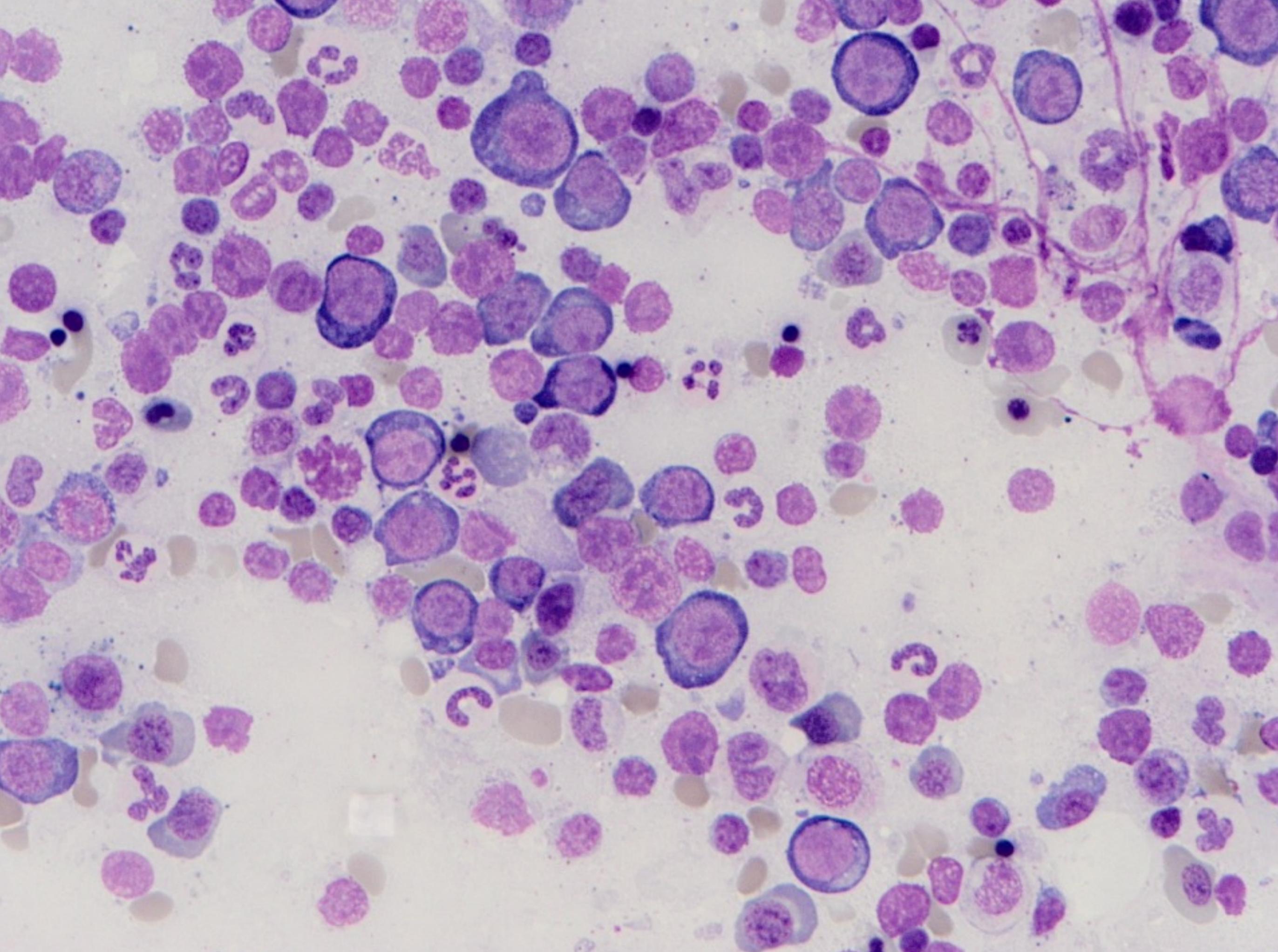


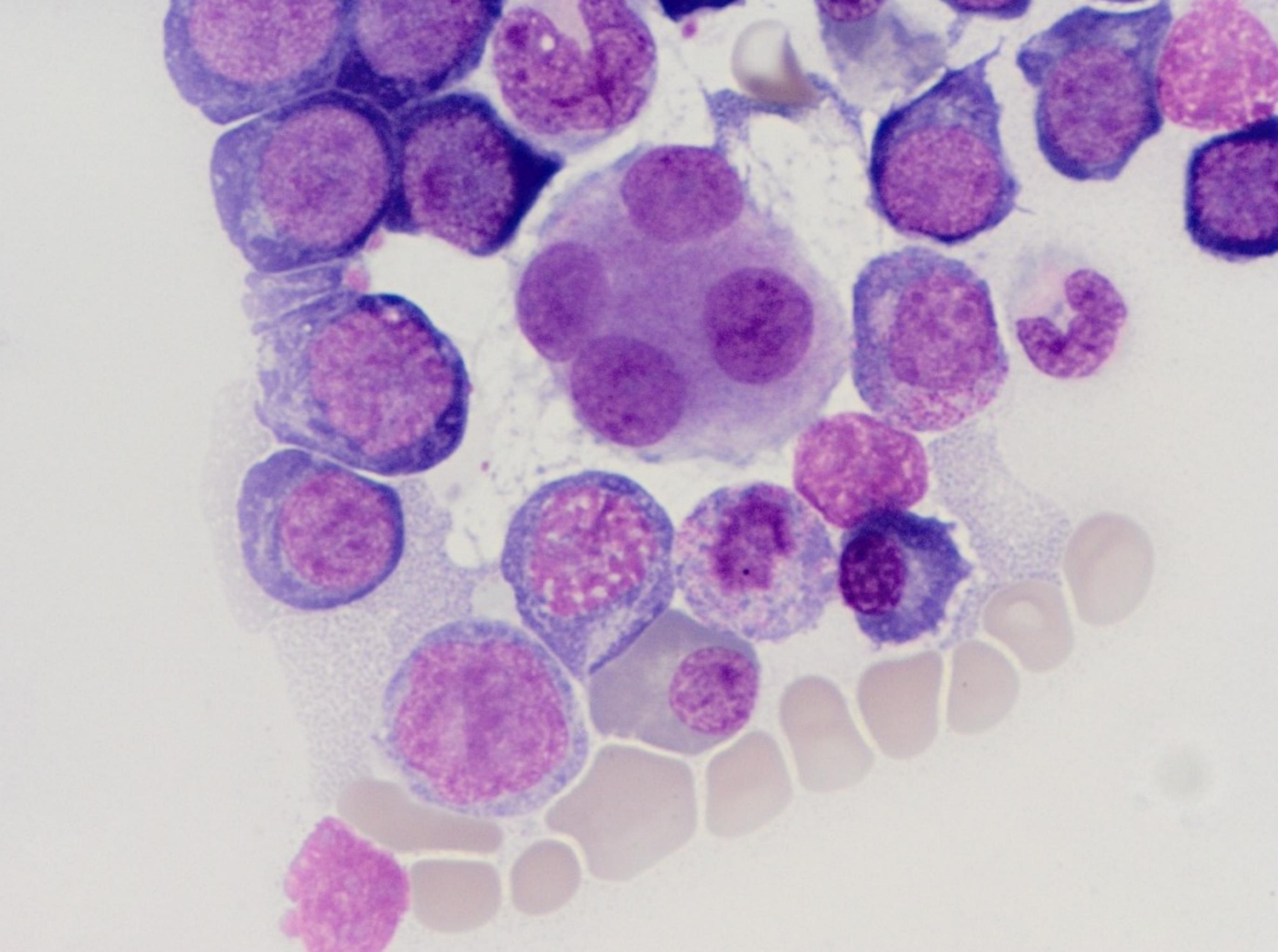
Blocage maturation

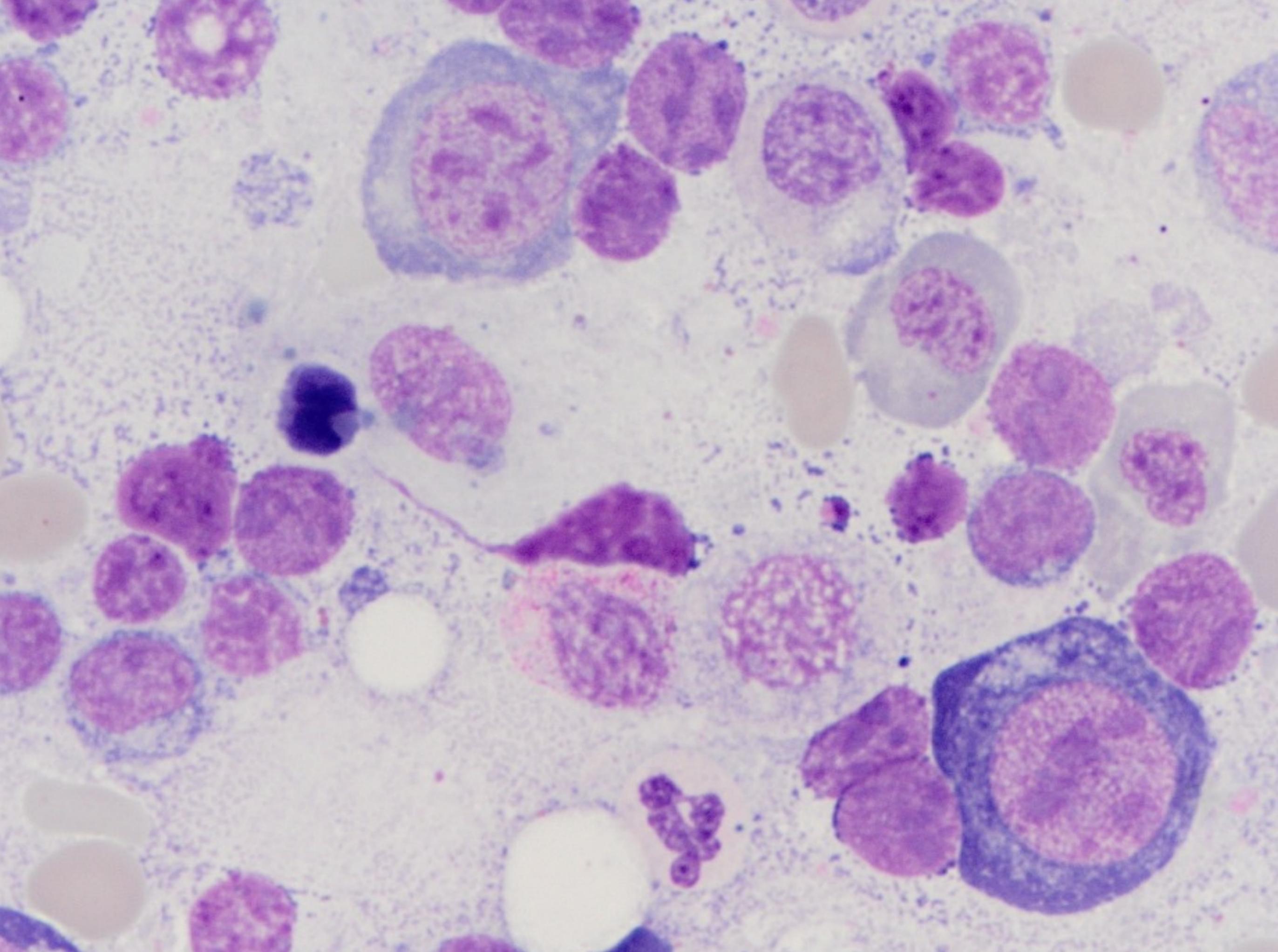
LLC post-traitement

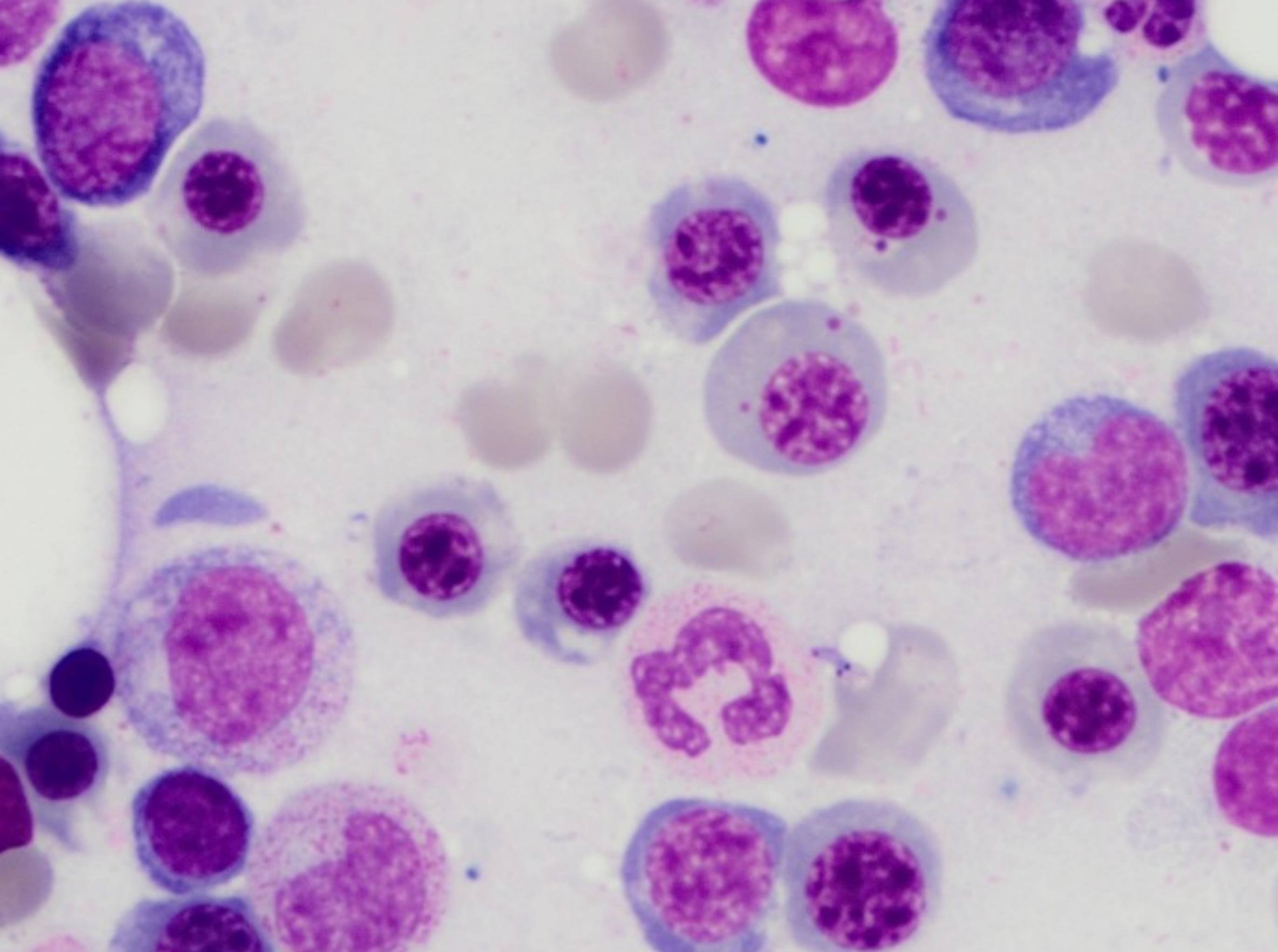


Score = 1

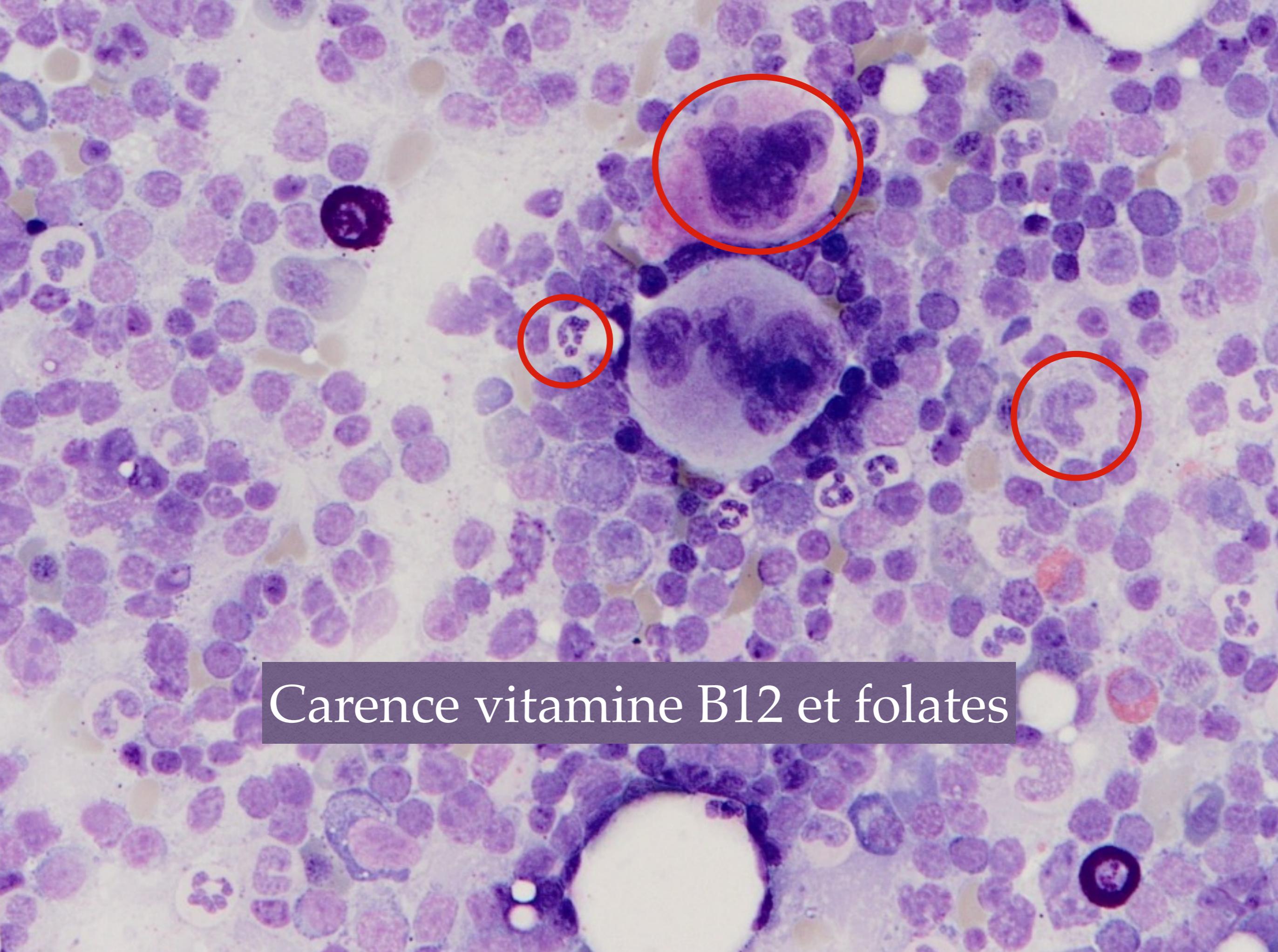












Carence vitamine B12 et folates



Cytogénétique ?  
50% ...

Morphologie  
«Standard» mais...

Quel panel ?  
*EuroFlow*

FC Scoring Systems  
*Oui, car manque de marqueurs  
discriminants dans les SMD*

Quelle est la place de la  
CMF dans les SMD ?  
*Certainement d'importance grandissante*

# Conclusions

**Multicentre validation of a reproducible flow cytometric score for the diagnosis of low-risk myelodysplastic syndromes: results of a European LeukemiaNET study**

by Matteo G. Della Porta, Cristina Picone, Cristiana Pascutto, Luca Malcovati, Hideto Tamura, Hiroshi Handa, Magdalena Czader, Sylvie Freeman, Pares Vyas, Anna Porwit, Leonie Saft, Theresia M. Westers, Canan Alhan, Caludia Cali, Arjan A. Van de Loosdrecht, and Kiyoyuki Ogata

*Haematologica* 2012 [Epub ahead of print]

- ❖ Il a été montré que la CMF est capable d'apporter une aide précieuse dans le diagnostic différentiel des syndromes myélodysplasiques.
- ❖ **Avantages du score :**
  - ❖ Peut être facilement implanté dans beaucoup de laboratoires et ne demande pas un panel d'anticorps extensif !
  - ❖ Les quatre paramètres possèdent une variabilité inter-opérateurs très faible, et leur expression est corrélée à la sévérité de la maladie.
  - ❖ Résultats facilement compris par les cliniciens.
  - ❖ Si le score est positif ( $\geq 2$ ), probabilité post-test d'avoir une pathologie est  $> 90\%$ .
  - ❖ Si le score est négatif, on ne peut exclure un diagnostic de SMD  $\rightarrow$  observation clinique pendant 6 mois suivie par une nouvelle évaluation médullaire ?

# Peut-on utiliser le scoring system pour diagnostiquer un SMD ?

- \* Notre étude montre que le score donne une bonne performance pour le diagnostic des syndromes myélodysplasiques —> utilisé dans notre laboratoire...
- \* Avec un seul tube (AML-1) ! Rapide, « pas cher » et standardisé...
- \* Améliorations diagnostiques :
  - \* Etudier plus de populations (monocytes, ,érythrocytes, ...)
  - \* Intérêt pour des marqueurs additionnels : CD7, CD15, CD56, ...  
**En seconde intention : HLA-DR / CD45 / CD15 / CD56 / CD117 / CD34 / CD7 / CD19**
  - \* Besoin de logiciels informatiques permettant une analyse multidimensionnelle —> peut aider à repérer les irrégularités dans les patterns de maturation, et aider lorsque l'on se retrouve devant des population se superposant.

L'immunophénotype peut aider à établir le diagnostic d'un SMD, en particulier quand la morphologie et la cytogénétique sont non discriminantes

# Merci

- ❖ **Nicolas Hougardy**
- ❖ **Nathalie Mernier**
- ❖ **Vincent Gobin**
- ❖ **Luc Guiot**

